Harmonisation of Regulatory Oversight in Biotechnology

Safety Assessment of Transgenic Organisms in the Environment, Volume 8

OECD CONSENSUS DOCUMENT OF THE BIOLOGY OF MOSQUITO *AEDES AEGYPTI*

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Foreword

Modern biotechnologies are applied to plants species (crops, flowers, trees), animals and micro-organisms. The safety of the resulting transgenic organisms when released in the environment for their use in agriculture, forestry, the food and feed industry or for other applications represents a challenging issue. Genetically engineered products are rigorously assessed by their developers during their elaboration, and by governments when ready for release, to ensure high safety standards. This remains essential with new biotechnology developments using insects to fight against disease outbreaks: engineered mosquitoes need to be evaluated through a scientifically sound approach to risk/safety assessment that will inform biosafety regulators and support the decision concerning the release of these novel organisms in the environment.

The OECD offers a long-standing recognised expertise in biosafety and contributes to facilitating an harmonised approach. The OECD's Working Group on Harmonisation of Regulatory Oversight in Biotechnology (WG-HROB) was established in 1995. The WG-HROB gathers national authorities responsible for the environmental risk/safety assessment of products of modern biotechnology in OECD countries and other economies. International organisations and experts involved in biosafety activities are associated with this programme.

The primary goals of the WG-HROB are to promote international regulatory harmonisation, to ensure that methods used in the risk/safety assessment of genetically engineered products are as similar as possible. This may open the way to possible recognition and even acceptance of information from the assessments of other countries. The benefits of harmonisation are multiple: it strengthens mutual understanding among countries, avoids duplication, saves resources and increases the efficiency of the risk assessment process. Overall, it improves safety while reducing unnecessary barriers to trade.

The environmental risk/safety assessment of transgenic organisms (biosafety assessment) is usually based on the information collected on the characteristics of the host organism, the introduced traits, the environment into which the organism will be released, the interaction between these, and the intended use of the organism for agriculture, forestry, food and feed industry, health improvement or other purposes. Since its establishment, the WG-HROB decided to focus its work on identifying parts of this information which could be commonly used among countries when conducting environmental risk/safety assessment, aiming to encourage information sharing and prevent duplication of effort among countries. Biosafety consensus documents are one of the major outputs of its work.

The OECD Biosafety consensus documents are intended to be a "snapshot" of current information on a specific host organism or trait, for use during regulatory assessments of organisms considered for their release in the environment. These publications are not designed to be a comprehensive source of information on everything that is known about

a specific host or trait, but they do address the key elements and core set of science-based issues that member countries believe are relevant to biosafety assessment. This information is said to be mutually acceptable among OECD Members and other economies associated with the work. The Biosafety consensus documents offer practical tools which compile science-based information useful for environmental risk/safety evaluation process. Because these documents are publicly available, they can also benefit other countries around the world wishing to use these tools along the same principles.

To date, a total of 57 consensus and guidance documents have been published by the WG-HROB. They mainly address the biology of crops, trees and micro-organisms, as well as specific traits introduced in engineered plants. Their scope is currently enlarging, in line with the new biotechnological developments and wider applications to new fields. The first document related to an animal species was published in 2017 on Atlantic salmon, a fish reared for food production but also occurring in the wild in undomesticated populations.

A further step has been taken in 2018 with this document on the mosquito *Aedes aegypti*, addressing for the first time the biology and ecology of an insect species. It is also the first OECD Biosafety consensus document to focus on an organism for which biotechnological applications are not aimed at an increase in production or the quality enhancement of the product (which are usual targets of crop variety improvement for instance) but are driven here by health considerations. In the case of mosquitoes, the objective of some current biotechnological developments is to fight against disease outbreaks by reducing the insect population or limiting its capacity to transmit diseases.

The mosquito *Ae. aegypti* is the main vector of viruses responsible for severe diseases such as yellow fever, dengue fever, Zika fever and chikungunya. This insect is currently subject to biotechnological research and applications (including genetic engineering), aiming to contribute to the control its population, reduce its capacity to spread diseases and thus limit its drastic impact on human health.

Over recent years, the epidemics brought by the mosquito have drastically spread in many tropical and sub-tropical regions of the world. The countries involved are developing strategic programmes which are specifically designed to control the *Ae. aegypti* population at local or regional level. These programmes often combine a range of chemical, biological and genetic control means, in addition to environmental management aiming to prevent the propagation of mosquito populations. The integrated vector management (IVM) is the approach promoted by the World Health Organization in support of these initiatives. More details on *Ae. aegypti* control is collated under Annex A to this document, while information regarding human and animal health affected by the mosquito is available in Annex B.

At the initiative of Central and South American countries involved in the OECD biosafety activities, the WG-HROB decided a few years ago to develop this document on the biology of the *Ae. aegypti* mosquito species. The objective was to provide a tool which could help authorities in charge of performing biosafety assessment relating to this insect. To cope with the new challenge, a team of regulators, assessors and scientists was established. The project was co-led by Mexico, Brazil and the ILSI Research Foundation, with additional expertise from Australia, France, India, Kenya, the United States and the industry sector. Other countries and observer organisations involved in the WG-HROB activities also contributed to the preparation of the document. In a workshop hosted by Mexico in May 2014 for launching the project, the experts elaborated the detailed outline of the document and agreed on an action plan. Successive drafts were prepared

through electronic exchange and reviewed by the whole WG-HROB at its annual meetings at each stage of the project development.

To conduct biosafety assessment of *Aedes aegypti*, a deep knowledge of the mosquito species is required to get a comprehensive view of its development, behaviour, and fully consider its potential interaction with the environment where it is to be released. This publication contains information relating to the mosquito taxonomy, morphology, reproductive biology, genetics, ecology and other aspects. Experts have summarised in this single document key elements from a vast range of solid scientific publications, selected for their potential interest during biosafety assessment, and carefully referenced at the end of the document. This information is intended to benefit-risk assessors that may need to consider potential effects on the environment when releasing engineered *Ae. aegypti* in the context of mosquito control programmes and therefore may contribute to facilitating the decision-making process.

The set of science-based information and data contained in this document, previously agreed by consensus and published by the OECD, constitute a solid reference recognised internationally, a tool for use during the biosafety assessment process. It is not intended to be a substitute for nationally-required information for risk/safety assessment, because they address only a part of the necessary elements. Nevertheless, they should make an important contribution to environmental risk/safety assessment. As such, this publication should be of value to applicants for commercial and public uses of engineered *Ae. aegypti* mosquitoes, to risk assessors and regulators from national authorities responsible for granting approvals to their release in the environment, as well as the wider scientific community.

The OECD Biosafety consensus documents are compiled in the successive volumes constituting the Series on Harmonisation of Regulatory Oversight in Biotechnology. The list shown at the end of the publication summarises the extent of the species covered, and indicates how they are grouped in their respective volumes. This Volume 8, however, contains a single document which differentiates by dealing with a novel topic (the biology of an insect species) and is large enough to constitute a specific publication.

Along with the previous seven volumes, Volume 8 includes the 'Introduction to the biosafety consensus documents' which explains in detail the purpose of these documents and how they are relevant to risk/safety assessment. It also describes the process by which the consensus documents are drafted, using a 'lead country(ies)' approach (two co-lead countries and one observer organisation in the case of the *Ae. aegypti* document).

The consensus documents published in the Volumes 1 to 8 of the Series are also available individually free of charge on the OECD BioTrack website www.oecd.org/biotrack.

In reading the OECD Biosafety consensus documents, it may be useful to also consult the "*Points to Consider for Consensus Documents on the Biology of Cultivated Plants*". Although this additional text is specifically for cultivated plants (crops and trees), it contains a structured checklist of "points to consider" relevant to risk/safety assessment that can also be considered by authors when drafting or reviewing a consensus document on the biology of animals used in agriculture or in health-related programmes.

Another document on the "*Environmental Considerations for Risk/Safety Assessment for the Release of Transgenic Plants*" is under preparation by the WG-HROB and will be published in the near future.

This biosafety work is complementary to the activities of the OECD programme on the safety of novel foods and feeds, in particular to the consensus documents developed on the composition of foods and feeds derived from transgenic organisms. These documents describe the key nutrients, anti-nutrients, toxicants and other constituents that can be used in a comparative approach. More information on this programme can be found in the introduction to this volume.

Another mosquito, *Anopheles gambiae*, is currently being considered by the WG-HROB for developing a similar biology document. This insect is causing a major public health concern at the global level, as the *A. gambiae* complex of species includes the most important vectors of malaria disease. A range of biotechnological solutions for its control is being explored. The future publication will constitute a useful complement to this publication by enlarging the range of insects covered by the OECD Biosafety consensus documents.

Acknowledgements

This book, containing the consensus document on the biology of mosquito *Aedes aegypti*, is the result of the common effort of the participants of the OECD's Working Group on Harmonisation of Regulatory Oversight in Biotechnology, with Mexico, Brazil and the ILSI Research Foundation having served as co-leads for the project and established their teams of specialists on the subject. In addition, experts from other delegations involved in the Ad hoc drafting group, namely Australia, France, India, Kenya, the United States and the industry sector, provided valuable contributions. During the preparation of the successive draft versions of the document, useful inputs and suggestions were also provided by a number of delegates and experts from the Working Group, whether from OECD member countries, non-member economies or observer organisations.

Once finalised and agreed on declassification in June 2018, the document was published by the OECD Environment, Health and Safety (EHS) Division in the Series on Harmonisation of Regulatory Oversight in Biotechnology. This volume was prepared by Ryudai Oshima and Bertrand Dagallier, with the editing contribution of Eleonore Morena, under the supervision of Peter Kearns at the EHS Division, OECD Environment Directorate.

The OECD is grateful to the scientists, safety assessors, regulators and authorities who participated in the development of this biosafety document on an innovative topic, and wishes to thank each of them.

Table of contents

Abbreviations and acronyms **13**

Executive summary **15**

Introduction to the biosafety consensus documents **17**

About the OECD's working group for biosafety 17
Regulatory harmonisation 17
The need for harmonisation activities at the OECD 18
Key background concepts and principles 18
A common approach to risk/safety assessment 18
The emergence of the concept of consensus documents 19
The purpose of consensus documents 19
The process through which consensus documents are initiated and brought to publication 20
Current and future trends in the Working Group on Harmonisation of Regulatory Oversight in Biotechnology 21
The OECD Working Group for the Safety of Novel Foods and Feeds 22
Notes 23
References 25
Annex to the Introduction: OECD biosafety principles and concepts developed prior to the Working Group on Harmonisation of Regulatory Oversight in Biotechnology (1986-94) 27

BIOLOGY OF MOSQUITO *AEDES AEGYPTI* **31**

Chapter 1. Taxonomy, description and distribution of the mosquito *Ae. aegypti* **33**

Classification and nomenclature of *Aedes aegypti* 34
Morphology 36
Origin and current geographic distribution 42
Notes 43
References 45

Chapter 2. Reproductive biology of the mosquito *Ae. aegypti* **49**

Life cycle 50
Reproduction 53
Life table analysis, under natural (and laboratory) conditions 59
Interspecific breeding 59
Effect of *Wolbachia* on reproduction 60
Notes 61
References 62

Chapter 3. Genetics of the mosquito *Ae. aegypti* **69**

Linkage map organisation of *Ae. aegypti* 70
Population genetics and phylogeography of *Ae. aegypti* 71
Genetics of insecticide susceptibility and development of insecticide resistance 73
Genetics of vector competence in *Ae. aegypti* 76
Notes 81
References 82

Chapter 4. Ecology of the mosquito *Ae. aegypti* **91**

Ecological niche/species distribution modelling of *Ae. aegypti* 92
Ae. aegypti niche and trophic interactions 92
Anthropic habitats 93
Abiotic requirements and tolerance 94
Biotic interactions in the landscape 95
Life history traits and fitness 96
Notes 99
References 100

Annex A. Control of the mosquito *Ae. aegypti* **107**

Current control strategies 108
Integrated control management 114
Prevention and management of insecticide resistance 115
References 116

Annex B. Human and animal health affected by mosquitoes **121**

Pathogens and diseases 122
Virus infection vectored by mosquitoes 122
Ae. Aegypti other characteristics 128
References 129

List of OECD consensus documents on environmental safety assessment, 1996-2018 ..... **131**

Tables

Table 1.1. Standardised taxonomic hierarchy and nomenclature for *Ae. aegypti* (Linnaeus, 1762) 34

Table A A.1. Summary of control tools/strategies available for *Ae. aegypti* 109
Table A B.1. Some important arbovirus infections of humans in geographic regions of the world ... 124

Figures

Figure 1.1. Dorsal view of *Ae. aegypti* larva 37
Figure 1.2. Comparison of the adults, eggs, larvae and pupae of mosquito genera *Anopheles*, *Aedes*, *Culex* and *Mansonia* 38
Figure 1.3. Comb scales of *Ae. aegypti* exhibiting a medial spine with stout, subapical spines and of *Ae. albopictus* without subapical spines 39

Figure 1.4. Anal segments of *Ae. aegypti* pupae - ventral view, showing dimorphism characters between females and males 40
Figure 1.5. Dorsal view of the female mosquito *Ae. aegypti* 40
Figure 1.6. Comparative dorsal view of thoracic scutum of *Ae. aegypti* and *Ae. albopictus* 42
Figure 1.7. Global map of the predicted distribution of *Ae. aegypti* 44
Figure 2.1. Life cycle of *Ae. aegypti* 50
Figure 2.2. *Ae. aegypti* gonotrophic cycle, taking into account the factors that can cause host-seeking behaviour to return after an initial blood meal 54
Figure 3.1. A physical map of the *Ae. aegypti* genome 71

Figure A B.1. Number of suspected or laboratory-confirmed dengue cases notified to WHO, 1990-2015 126

Abbreviations and acronyms

°C	Degree Celsius
µl	Microlitre
Ae.	*Aedes*
Bti	*Bacillus thuringiensis* var. *israelensis*
CDC	Centres for Disease Control and prevention
CHIKV	Chikungunya virus
CI	Cytoplasmic incompatibility
CRISPR	Clustered regularly interspaced short palindromic repeats
DDT	Dichlorodiphenyltrichloroethane
DENV	Dengue virus
DENV-1	Dengue 1 virus
DENV-2	Dengue 2 virus
DENV-3	Dengue 3 virus
DENV-4	Dengue 4 virus
DNA	Deoxyribonucleic acid
EFSA	European Food Safety Authority
EIP	Entrinsic incubation period
ELISA	Enzyme-linked immunosorbent assay
FISH	Fluorescence in-situ hybridisation
g/ha	Gramme per hectare
Gb	Gigabases (= 10^9 base pairs; genome size unit)
GE	Genetically engineered
HEGs	Homing endonuclease genes
ICTV	International Committee on Taxonomy of Viruses
IGR	Insect growth regulator
IIT	Incompatible insect technique
IRM	Insecticide resistance management
IRS	Indoor residual spraying
ISS	Indoor space-spraying
IVM	Integrated vector management
kdr	Knockdown resistance
km	Kilometre
LGT	Lateral gene transfer
LLIN	Long-lasting insecticidal netting
m	Metre
Mb	Megabases (= 10^6 base pairs; genome size unit)
MDT	Mean distance travelled
MEB	Midgut escape barrier
MG	Midgut

MIB	Midgut infection barrier
MITE	Miniature inverted repeat transposable element
ml	Millilitre
mm	Millimetre
NADH	Nicotinamide adenine dinucleotide
ND4	NADH dehydrogenase subunit 4
NVA	Neovolcanic axis
PAHO	Pan American Health Organization
PCR	Polymerase chain reaction
piRNA	Piwi-interacting RNA (ribonucleic acid)
QTL	Quantitative trait locus [plural form: Q. t. loci]
RAPD	Random amplified polymorphic DNA (deoxyribonucleic acid)
RFLP	Restriction fragment length polymorphism
RNAi	RNA (ribonucleic acid) interference
SIT	Sterile insect technique
SNP	Single nucleotide polymorphism
STS	Sequence-tagged site
TE	Transposable elements
UBH	Uriah Butler Highway (in Trinidad Island, Trinidad and Tobago)
VC	Vector competence
WHO	World Health Organization
WHOPES	World Health Organization Pesticide Evaluation Scheme
YFV	Yellow fever virus
ZIKV	Zika virus

Executive summary

This Volume 8 contains the "OECD consensus document on the biology of mosquito *Aedes aegypti*". It is published in the Series on Harmonisation of Regulatory Oversight in Biotechnology which relates to the environmental risk/safety assessment of transgenic organisms, also called "biosafety" assessment. This new publication provides a useful tool to national authorities and scientists involved in the evaluation of the safety of genetically-engineered mosquitoes when released in the environment.

The mosquito *Aedes aegypti* is of major public health concern, being the main vector of viruses responsible for diseases such as yellow fever, dengue fever, Zika fever and chikungunya. Its development in tropical and sub-tropical areas is intrinsically linked to human habitats and activities that offer the insect its adequate living conditions and the blood meal it needs for reproduction. This mosquito species is subject to biotechnological research and applications (including genetic engineering), aiming to contribute to the control of its population and thus limiting its drastic impact on human health.

Considering the rising spread of related epidemics in many parts of the world, together with the development of genetically-engineered mosquitoes contemplated for use in integrated control management, the OECD Working Group on Harmonisation of Regulatory Oversight in Biotechnology (WG-HROB) decided to develop this document on *Aedes aegypti* biology. The project, launched in 2014, was co-led by Mexico, Brazil and the ILSI Research Foundation, with additional expertise provided by Australia, France, India, Kenya, the United States and the industry sector. Other countries and observer organisations involved in the WG-HROB activities also participated in the preparation of the document.

Modern biotechnologies are applied to crop plants, as well as trees, animals and micro-organisms. The safety of the resulting transgenic organisms when released in the environment for their use in agriculture, forestry, the food and feed industry or for other applications represents a challenging issue. Genetically engineered products are rigorously assessed by their developers during their elaboration, and by governments when ready for release, to ensure high safety standards. This remains essential with new biotechnology developments using insects to fight against disease outbreaks: engineered mosquitoes need to be evaluated through a scientifically sound approach to risk/safety assessment that will inform biosafety regulators and support the decision concerning the release of these novel organisms in the environment.

The OECD offers a long-standing recognised expertise in biosafety and contributes to facilitating an harmonised approach. The environmental risk/safety assessment of transgenic organisms is usually based on the information collected on the characteristics of the host organism, the introduced traits, the environment into which the organism will be released, the interaction between these, and the intended use of the organism. The OECD Biosafety consensus documents elaborated by the WG-HROB identify parts of this information which could be commonly used in countries when conducting environmental risk/safety assessment, aiming to encourage

information sharing and prevent duplication of effort among countries. They offer practical tools which compile science-based information relevant for this purpose. They are not a substitute for national requirements and locally-available data should also be taken into account, but they can contribute to the risk/safety assessment process. These documents are publicly available and considered worldwide as sustainable references for use in biosafety evaluation.

To conduct biosafety assessment of *Aedes aegypti*, a deep knowledge of the mosquito species is required to fully consider its potential interaction with the environment of release. Useful information can go from accurate taxonomic nomenclature, the origin of the species and its current distribution in the world, up to the life cycle of the mosquito in its successive forms (eggs, larvae, pupae and male/female adults). The reproductive biology is also essential to understand its behaviour: what are its breeding sites and reproduction features (mating, physiological aspects, fecundity), and the potential effect of *Wolbachia* bacteria. The *Ae. aegypti* genetics is also of great value, including genetic linkage map, population genetics and phylogeography, susceptibility to insecticides and resistance mechanisms, as well as genetic variability in the mosquito competence to transmit virus infection. Then, it is crucial for biosafety assessors to acquire extensive knowledge of the ecology of this mosquito, i.e. its interactions with the other species in the environment: ecological niche it occupies; the climatic parameters influencing its development; its anthropic habitats in strong connection with human population; the abiotic requirements in terms of water and food availability; and the fitness to local conditions including its dispersal, population distribution and modelling.

To prepare this publication, experts have summarised in this single document key elements from a vast range of solid science-based publications, selected for their potential interest during biosafety assessment and carefully referenced. This information is intended to benefit-risk assessors that may need to consider potential effects on the environment when releasing engineered *Aedes aegypti* in the context of mosquito control programmes, and therefore may contribute in facilitating the decision-making process.

Opening Volume 8, the introduction to the Biosafety consensus documents provides additional information on the key background concepts, principles and common approach prevailing in risk/safety assessment of transgenic organisms. The purpose of the OECD consensus documents are described in detail, as well as the process by which they are developed. These publications address the biology of crops, trees and micro-organisms, as well as specific traits introduced in engineered plants, with a recent scope extension to animal species.

Another mosquito, *Anopheles gambiae*, is currently considered by the WG-HROB for developing a similar biology document. The *A. gambiae* complex of species includes the most important vectors of malaria disease, and biotechnological solutions for its control are being explored. The future document will constitute a useful complement to this publication by enlarging the scope of insects covered by the OECD Biosafety consensus documents.

Introduction to the biosafety consensus documents

About the OECD's working group for biosafety

The OECD's Working Group on Harmonisation of Regulatory Oversight in Biotechnology (the "WG-HROB") comprises delegates from the 35 member countries of the OECD and the European Commission. Typically, delegates are from those government ministries and agencies which have responsibility for the environmental risk/safety assessment of products of modern biotechnology. The WG-HROB also includes a number of observer delegations and invited experts who participate in its work, such as Argentina, the Russian Federation, the United Nations Environment Programme (UNEP), the Secretariat of the Convention on Biological Diversity (SCBD), the Food and Agriculture Organization of the United Nations (FAO), the United Nations Industrial Development Organisation (UNIDO), and the Business and Industry Advisory Committee to the OECD (BIAC).

In recent years, with the increasing use of biotech products in many regions of the world, together with the development of activities relating to tropical and subtropical species, participation was enlarged to other non-member economies including Bangladesh, Brazil, the People's Republic of China, Colombia, India, Indonesia, Kenya, Lithuania, Paraguay, the Philippines, South Africa and Viet Nam, as well as the African Biosafety Network of Expertise from the New Partnership for Africa's Development, a body of the African Union (AU-NEPAD-ABNE). From July 2011 to December 2014, a programme was jointly implemented by the World Bank, the ILSI Research Foundation – Center for Environmental Risk Assessment (ILSI-CERA) and the OECD in the framework of the "Partnership for Biosafety Risk Assessment and Regulation", which developed new links, enhanced collaboration and supported the participation of four non-member economies in the activities of the WG-HROB.

Regulatory harmonisation

The Working Group on Harmonisation of Regulatory Oversight in Biotechnology was established in 1995[1] at a time when the first commercial transgenic crops were being considered for regulatory approval in a number of OECD countries. From the beginning, one of the group's primary goals was to promote international regulatory harmonisation in biotechnology among members. Regulatory harmonisation is the attempt to ensure that the information used in risk/safety assessments, as well as the methods used to collect such information, is as similar as possible. This should lead to countries recognising or even accepting information from one another's assessments. The benefits of harmonisation are clear. It increases mutual understanding among countries, which avoids duplication, saves on scarce resources and increases the efficiency of the risk/safety assessment process. This, in turn, improves safety while reducing unnecessary barriers to trade (OECD, 2000).

The need for harmonisation activities at the OECD

The establishment of the WG-HROB and its programme of work followed a detailed analysis by member countries of whether there was a need to continue work on harmonisation in biotechnology at the OECD, and if so, what it should entail. This analysis was undertaken by the Ad Hoc Group for Environmental Aspects of Biotechnology (established by the Joint Meeting),[2] in 1994.

The Ad Hoc Group for Environmental Aspects of Biotechnology took into consideration and built upon, the earlier work at the OECD which had begun in the mid-1980s. Initially, these OECD activities focused on the environmental and agricultural implications of field trials of transgenic organisms, but this was soon followed by a consideration of their large-scale use and commercialisation (a summary of this extensive body of work can be found in the annex to this Introduction section.)

Key background concepts and principles

The Ad Hoc Group for Environmental Aspects of Biotechnology took into account previous work on risk analysis that is summarised in Safety Considerations for Biotechnology: Scale-up of Crop Plants (OECD, 1993a). The following quote gives the flavour: "Risk/safety analysis is based on the characteristics of the organism, the introduced trait, the environment into which the organism is introduced, the interaction between these, and the intended application". This body of work has formed the basis for environmental risk/safety assessment that is now globally accepted. In considering the possibilities for harmonisation, the Ad hoc group paid attention to these characteristics and the information used by risk/safety assessors to address them.

This was reinforced by the concept of familiarity, also elaborated in the above-mentioned document (OECD, 1993a). This concept "is based on the fact that most genetically engineered organisms are developed from organisms such as crop plants whose biology is well understood. Familiarity allows the risk assessor to draw on previous knowledge and experience with the introduction of plants and micro-organisms into the environment". For plants, familiarity takes account of a wide range of attributes including, for example, knowledge and experience with "the crop plant, including its flowering/reproductive characteristics, ecological requirements, and past breeding experiences" (OECD, 1993a; see also the annex for a more detailed description). This illustrates the role of information related to the biology of the host organism as a part of an environmental risk/safety assessment.

The Ad Hoc Group for Environmental Aspects of Biotechnology also considered the document Traditional Crop Breeding Practices: An Historical Review to Serve as a Baseline for Assessing the Role of Modern Biotechnology (OECD, 1993b), which focuses on host organisms. It presents information on an initial group of 17 different crop plants, which are used (or are likely to be used) in modern biotechnology. It includes sections on phytosanitary considerations in the movement of germplasm and on current uses of these crop plants. There is also a detailed section on current breeding practices.

A common approach to risk/safety assessment

An important aspect for the Ad Hoc Group for Environmental Aspects of Biotechnology was to identify the extent to which member countries address the same questions and issues during

risk/safety assessment. Big differences would mean difficulties in working towards harmonisation, while a high level of similarity would suggest it is more feasible.

This point was resolved by two studies considered by the Ad hoc group: one covered crop plants (OECD, 1995a; 1995b) while the other concerned micro-organisms (OECD, 1995c; 1995d). Both studies involved a survey with national authorities responsible for risk/safety assessment. The aim was to identify the questions they address during the assessment process (as outlined in national laws/regulations/guidance texts) in order to establish the extent of similarity among national authorities. The studies used the information provided in the OECD's "Blue Book" on Recombinant DNA Safety Considerations (OECD, 1986) as a reference point, in particular the sections covering: 1) general scientific considerations; 2) human health considerations; and 3) environmental and agricultural considerations (Appendices B, C and D). Both studies showed a remarkably high degree of similarity among countries in the questions/issues addressed in risk/safety assessment.

The emergence of the concept of consensus documents

The Working Group on Harmonisation of Regulatory Oversight in Biotechnology was therefore established with the knowledge that national authorities have much in common in terms of the questions/issues addressed when undertaking risk/safety assessment. It also took into account those characteristics identified as part of the assessment (i.e. the organism, the introduced trait and the environment) around which harmonisation activities could focus.

It was further recognised that much of the information used in risk/safety assessment relating to the biology of host organisms (crop plants, trees, animals or micro-organisms) would be similar or virtually the same in all assessments involving the same organism. In other words, the questions addressed during risk/safety assessment which relate to the biology of the organism, for example the potential for gene transfer within the crop plant species, and among related species, as well as the potential for weediness, remain the same for each application involving the same host species. This also applies to some extent to information related to introduced traits.

Consequently, the WG-HROB put forth the idea of compiling information common to the risk/safety assessment of a number of transgenic products and decided to focus on two specific categories: the biology of the host species and traits used in genetic modifications. The aim was to encourage information sharing and prevent duplication of effort among countries by avoiding the need to address the same common issues in applications involving the same organism or trait. It was recognised that biology and trait consensus documents could be agreed upon relatively quickly by member countries (within a few years). This compilation process was quickly formalised in the drafting of consensus documents.

The purpose of consensus documents

The consensus documents are not intended to be a substitute for a risk/safety assessment, because they address only a part of the necessary information. Nevertheless, they should make an important contribution to environmental risk/safety assessment.

Consensus documents are intended to be a "snapshot" of current information, for use during the regulatory assessment of products of biotechnology. They are not intended to be a comprehensive source of information covering the full knowledge about a specific

host organism or trait; but they address – on a consensus basis – the key or core set of issues that countries believe to be relevant to risk/safety assessment.

The aim of the documents is to share information on these key components of an environmental safety review in order to prevent duplication of effort among countries. The documents are envisaged to be used: 1) by applicants as information to be given in applications to regulatory authorities; 2) by regulators as a general guide and reference source in their reviews; and 3) by governments for information sharing, research reference and public information.

Originally, it was said that the information in the consensus documents is intended to be mutually recognised or mutually acceptable among OECD member countries, though the precise meaning of these terms is still open for discussion. During the period of the Ad Hoc Group for Environmental Aspects of Biotechnology and the early days of the WG-HROB (1993-95), the phrase "mutual acceptance of data" was discussed. This concept, borrowed from OECD's Chemicals Programme, involves OECD Council decisions that have legally binding implications for member countries. In the case of the consensus documents, there has never been a legally binding commitment to use the information they contain, though the WG-HROB is interested in enhancing the commitment of countries to make use of the documents. Participation in the development of documents, and the intention by countries to use the information, is done in "good faith". It is expected, therefore, that reference will be made to relevant consensus documents during risk/safety assessments. As these documents are publicly available, they can be of interest for any country wishing to use them in national assessments.

The process through which consensus documents are initiated and brought to publication

There are a number of steps in the drafting of a specific consensus document. The first occurs when a delegation, in a formal meeting of the Working Group on Harmonisation of Regulatory Oversight in Biotechnology, makes a proposal to draft a document on a new topic, typically a crop species or a trait. If the WG-HROB agrees to the proposal, a provisional draft is prepared by either a single country or two or more countries working together ("lead country approach"). Typically, the lead country(ies) has had experience with the concerned crop, animal or trait and is able to draw on experts to prepare a provisional draft. Where relevant an Ad hoc group is constituted with experts from several interested countries and observer organisations, bringing the range of current knowledge on the specific topic, in order to contribute at best to the drafting exercise.

The provisional draft is first reviewed by the Bureau of the WG-HROB[3] to ensure that it addresses the range of issues normally covered by consensus documents and is of sufficiently high quality to merit consideration by the WG-HROB as a whole.

Based on the comments of the Bureau, a first draft is prepared for consideration by the full WG-HROB. This is the opportunity for each delegation to review the text and provide comments based on their national experiences. Inputs are incorporated in a second draft, which is again circulated to the WG-HROB. At this point, the WG-HROB may decide to recommend that the document should be declassified. Such a recommendation is only forthcoming when all delegations have come to a consensus that the document is complete and ready for publication. Sometimes, however, the text may need a third round of discussions or even more within the WG-HROB before a declassification can be contemplated.

Once the WG-HROB has agreed for a final document to be ready for publication, it is forwarded to the supervisory committee, the Joint Meeting, recommending declassification. Following the agreement of the Joint Meeting, the document is then published.

It is important to note that the review of consensus documents is not limited to formal meetings of the WG-HROB. The Ad hoc expert groups might also exchange in face-to-face meetings or workshops, where feasible. And much discussion occurs through electronic means during the whole process, especially via the protected website dedicated to the WG-HROB. This enables a range of experts to have input into drafts.

For a number of documents, it has also been necessary to include information from non-member countries. This wider share of expertise has become increasingly important in recent years with the development of activities relating to tropical and subtropical species. This has been particularly true in the case of crop plants where the centre of origin and diversity occurs in a non-member country(ies). In these cases, UNEP, UNIDO, the FAO and other organisations have assisted in the preparation of documents by identifying experts from relevant countries, including international agricultural research centres as appropriate.

The full series of consensus documents developed by the WG-HROB is also published in compendium documents, as it is the case for this volume. Volume 7 was issued in 2017 (covering 2016-17), Volumes 5 and 6 in 2016 (covering 2011-15), Volumes 3 and 4 in 2010 (covering 2007-10), while Volumes 1 and 2 were published in 2006 (covering 1996-2006) (OECD, 2006a, 2006b, 2010a, 2010b, 2016a, 2016b, 2016c, 2017).

Current and future trends in the Working Group on Harmonisation of Regulatory Oversight in Biotechnology

The WG-HROB continues its work on the preparation of specific consensus documents, and on the efficiency of the process by which they are developed. An increasingly large number of crops and other host species (trees, animals, micro-organisms) are being modified, for an increasing number of traits, and the WG-HROB aims to fulfil the current needs whilst preparing for emerging topics.

At the OECD Workshop on Consensus Documents and Future Work in Harmonisation held in Washington, DC in October 2003, the WG-HROB considered how to set priorities for drafting future consensus documents among a large number of possibilities. It was also recognised that published consensus documents may be in need of review and updating from time to time, to ensure that they include the most up-to-date information. The WG-HROB considers these aspects on a regular basis when planning future work. For the preparation of future documents, the workshop identified the usefulness of developing a standardised structure of consensus documents. Thus, the WG-HROB has developed, first, a guidance document on “points to consider” for consensus documents on the biology of cultivated plants that was published in 2006, and then that of the trait documents. The “points to consider” document, included in Volumes 3 and 4 of the compendia series, is currently being updated by the WG-HROB.

Among the important activities of the WG-HROB, a new document is being developed on the “environmental considerations for the risk/safety assessment for the release of transgenic plants”. Focused on the core of the biosafety work that is applied to crops and trees and taking into account the most recent views from countries of all regions of

the world, this document will constitute a key guidance tool for developers, assessors and regulatory authorities. It is expected to be published in 2019.

An important step was taken in 2017 with the publication of the first consensus biology document dedicated to an animal species, the Atlantic salmon (*Salmo salar*). It was followed one year after by the publication on the mosquito *Aedes aegypti* (included in the present volume), which constitutes a key development for the WG-HROB by enlarging further the range of organisms potentially covered, and directly contributing to human health issues for the first time. Some genetically engineered strains of *Ae. aegypti* have been used since 2014 in limited areas, aiming to control the virus-vector insect population in the fight against tropical diseases (yellow fever, dengue and others) that have been dramatically extending in many regions of the world over the last decade.

The WG-HROB is also considering projects on micro-organisms, therefore opening up to new areas, for instance, bioenergy, with the preparation of a document on eukaryotic micro-algae having started recently. The photosynthetic cyanobacteria are potential providers of renewable energy and are of special interest as they can be cultivated year-round on non-arable areas, alleviating the pressure on farmland and freshwater resources that would be exerted by crops grown for biofuel purposes, as stated in the proceedings of the OECD Conference on Biosafety and the Environmental Uses of Micro-Organisms set up by the WG-HROB in 2012 (OECD, 2015a). Other biotechnology developments applied to micro-organisms might be considered to prepare future documents: an updated review of biofertiliser organisms living in symbiosis in crop roots and optimising the nitrogen fixation, or biocontrol agents acting as plant protection products to control disease and attack by insects and other herbivores. Other exploratory fields may comprise bioremediation by using living organisms for removing contaminants from the environment such as polluted land, or the development of detergents containing micro-organisms.

In recent years, the WG-HROB has started to exchange knowledge and promote discussion on the new plant breeding techniques and their potential impact on biosafety assessment. An OECD workshop was organised on these matters in 2014; the key message from its report at the time was that "experience to date indicates that current guidance and tools for environmental risk/safety assessment of transgenic plants are applicable to plants developed using [new plant breeding techniques]", where such assessment may be required (OECD, 2016c). Specific events on new plant breeding techniques are regularly organised at the OECD for increasing awareness and sharing information, including a conference on genome editing applications in 2018. The subject will be kept under review.

The OECD Working Group for the Safety of Novel Foods and Feeds

The OECD Task Force for the Safety of Novel Foods and Feeds, established in 1999, addresses aspects of the assessment of human food and animal feed derived from genetically engineered crops. This body was renamed the Working Group for the Safety of Novel Foods and Feeds (WG-SNFF) from 1 January 2017. As with the WG-HROB, the main focus of the WG-SNFF work is to ensure that the types of information used in risk/safety assessment, as well as the methods to collect such information, are as similar as possible amongst countries. The approach is to compare transgenic crops and derived products with similar conventional ones that are already known and considered safe because of their history of safe use. Harmonised methods and the sharing of information are facilitated through the WG-SNFF's activities.

In a similar approach to the biosafety programme, the main outcome of the foods and feeds programme is the set of consensus documents on compositional considerations of new varieties of specific crops. The WG-SNFF documents compile a common base of scientific information on the major components of crop plants, such as key nutrients, anti-nutrients, toxicants, allergens and other constituents. These documents constitute practical tools for regulators and risk/safety assessors dealing with these new varieties, with respect to foods and feeds. To date, 28 consensus documents have been published on major crops and on general considerations for facilitating harmonisation, including regular updates of the oldest issues. They constitute the Series on the Safety of Novel Foods and Feeds which is also available on the OECD's website (www.oecd.org/biotrack).

The full series of consensus documents developed by the Task Force was published in 2015 in two compendium documents, Volume 1 covering 2002-08 and Volume 2 covering 2009-14 (OECD, 2015b, 2015c). Volume 3 is under preparation.

The two bodies (WG-HROB and WG-SNFF) are implementing closely related and complementary programmes, focused on environmental aspects for the first and on food and feed aspects for the second. Their co-operation on issues of common interest resulted in a document developed jointly by the two bodies, the "Consensus document on molecular characterisation of plants derived from modern biotechnology", published in 2010 (included in Volume 3 of the current series). The two bodies also refer to the same "Unique Identifiers" assigned to transgenic products approved for cultivation and/or for food and feed use, and they wish to keep this system defined by OECD (described in Volume 3 of the current series) always relevant and adapted to new types of products-new species.

Notes

[1] The original title of the Working Group was the "Expert Group for the Harmonisation of Regulatory Oversight in Biotechnology". It became an OECD working group in 1998.

[2] The Joint Meeting was the supervisory body of the Ad Hoc Group for Environmental Aspects of Biotechnology and, as a result of its findings, established the working group as a subsidiary body. Today, its full title is the Joint Meeting of the Chemicals Committee and the Working Party on Chemical, Pesticides and Biotechnology.

[3] The Bureau comprises the Chair and Vice-Chairs of the working group. The Bureau is elected by the working group once per year. At the time of preparing this publication, the Chair is from the United States, and the Vice-Chairs from Australia, Belgium, Canada, Finland and Japan.

References

OECD (2017), *Safety Assessment of Transgenic Organisms in the Environment, Volume 7: OECD Consensus Documents*, Harmonisation of Regulatory Oversight in Biotechnology, OECD Publishing, Paris, http://dx.doi.org/10.1787/9789264279728-en

OECD (2016a), *Safety Assessment of Transgenic Organisms in the Environment, Volume 6: OECD Consensus Documents*, Harmonisation of Regulatory Oversight in Biotechnology, OECD Publishing, Paris, http://dx.doi.org/10.1787/9789264253421-en

OECD (2016b), *Safety Assessment of Transgenic Organisms in the Environment, Volume 5: OECD Consensus Documents*, Harmonisation of Regulatory Oversight in Biotechnology, OECD Publishing, Paris, http://dx.doi.org/10.1787/9789264253018-en

OECD (2016c), "Report of the OECD Workshop on Environmental Risk Assessment of Products Derived from New Plant Breeding Techniques (February 2014)", *Series on Harmonisation of Regulatory Oversight in Biotechnology No. 61*, OECD, Paris, www.oecd.org/officialdocuments/publicdisplaydocumentpdf/?cote=env/jm/mono(2016)5&doclanguage=en.

OECD (2015a), *Biosafety and the Environmental Uses of Micro-Organisms: Conference Proceedings*, OECD Publishing, Paris, http://dx.doi.org/10.1787/9789264213562-en.

OECD (2015b), *Safety Assessment of Foods and Feeds Derived from Transgenic Crops, Volume 2*, Novel Food and Feed Safety, OECD Publishing, Paris, http://dx.doi.org/10.1787/9789264180147-en.

OECD (2015c), *Safety Assessment of Foods and Feeds Derived from Transgenic Crops, Volume 1*, Novel Food and Feed Safety, OECD Publishing, Paris, http://dx.doi.org/10.1787/9789264180338-en.

OECD (2010a), *Safety Assessment of Transgenic Organisms: OECD Consensus Documents: Volume 4*, Harmonisation of Regulatory Oversight in Biotechnology, OECD Publishing, Paris, http://dx.doi.org/10.1787/9789264096158-en.

OECD (2010b), *Safety Assessment of Transgenic Organisms: OECD Consensus Documents: Volume 3*, Harmonisation of Regulatory Oversight in Biotechnology, OECD Publishing, Paris, http://dx.doi.org/10.1787/9789264095434-en.

OECD (2006a), *Safety Assessment of Transgenic Organisms: OECD Consensus Documents: Volume 2*, Harmonisation of Regulatory Oversight in Biotechnology, OECD Publishing, Paris, http://dx.doi.org/10.1787/9789264095403-en.

OECD (2006b), *Safety Assessment of Transgenic Organisms: OECD Consensus Documents: Volume 1*, Harmonisation of Regulatory Oversight in Biotechnology, OECD Publishing, Paris, http://dx.doi.org/10.1787/9789264095380-en.

OECD (2000), "Report of the Working Group on Harmonisation of Regulatory Oversight in Biotechnology", prepared for the G8 Summit held in Okinawa, Japan on 21-23 July 2000, C(2000)86/ADD2, OECD, Paris, www.oecd.org/chemicalsafety/biotrack/Report-of-the-Working-Group-on-Harmonisation-of-Regulatory.pdf.

OECD (1995a), "Commercialisation of agricultural products derived through modern biotechnology: Survey results", *OECD Environment Monograph: Series No. 99*, OECD, Paris, www.oecd.org/science/biotrack/1876950.pdf.

OECD (1995b), "Report of the OECD Workshop on the Commercialisation of Agricultural Products Derived through Modern Biotechnology", *OECD Environment Monograph: Series No. 107*, OECD, Paris, www.oecd.org/officialdocuments/publicdisplaydocumentpdf/?cote=OCDE/GD(95)72&docLanguage=En.

OECD (1995c), "Analysis of information elements used in the assessment of certain products of modern biotechnology", *OECD Environment Monograph: Series No. 100*, OECD, Paris, www.oecd.org/officialdocuments/publicdisplaydocumentpdf/?cote=OCDE/GD(95)11&docLanguage=En.

OECD (1995d), *Safety Considerations for Biotechnology: Scale-up of Micro-organisms as Biofertilizers*, OECD, Paris, www.oecd.org/env/ehs/biotrack/Safety-considerations-scale-up-of-micro-organisms-as-biofertilizers.pdf.

OECD (1993a), *Safety Considerations for Biotechnology: Scale-up of Crop Plants*, OECD, Paris, www.oecd.org/env/ehs/biotrack/1958527.pdf.

OECD (1993b), *Traditional Crop Breeding Practices: An Historical Review to Serve as a Baseline for Assessing the Role of Modern Biotechnology*, OECD, Paris, www.oecd.org/env/ehs/biotrack/1946204.pdf.

OECD (1986), *Recombinant DNA Safety Considerations. Safety Considerations for Industrial, Agricultural and Environmental Applications of Organisms Derived by Recombinant DNA Techniques* ("The Blue Book"), OECD, Paris, www.oecd.org/env/ehs/biotrack/Recombinant-DNA-Safety-Considerations.pdf.

Annex to the Introduction: OECD biosafety principles and concepts developed prior to the Working Group on Harmonisation of Regulatory Oversight in Biotechnology (1986-94)

Since the mid-1980s the OECD has been developing harmonised approaches to the risk/safety assessment of products of modern biotechnology. Prior to the establishment of the Working Group on Harmonisation of Regulatory Oversight in Biotechnology, the OECD published a number of reports on safety considerations, concepts and principles for risk/safety assessment as well as information on field releases of transgenic crops, and a consideration of traditional crop breeding practices. This annex notes some of the highlights of these achievements that were background considerations in the establishment of the working group and its development of consensus documents.

Underlying scientific principles

In 1986, the OECD published its first safety considerations for genetically engineered organisms (OECD, 1986). These included the issues relevant to human health, the environment and agriculture that might be considered in a risk/safety assessment. In its recommendations for agricultural and environmental applications, it suggested that risk/safety assessors:

- "Use the considerable data on the environmental and human health effects of living organisms to guide risk assessments.
- Ensure that recombinant DNA organisms are evaluated for potential risk, prior to application in agriculture and the environment by means of an independent review of potential risks on a case-by-case basis.
- Conduct the development of recombinant DNA organisms for agricultural and environmental applications in a stepwise fashion, moving, where appropriate, from the laboratory to the growth chamber and greenhouse, to limited field testing and finally to large-scale field testing.
- Encourage further research to improve the prediction, evaluation, and monitoring of the outcome of applications of recombinant DNA organisms."

The role of confinement in small-scale testing

In 1992, OECD published its *Good Developmental Principles* (OECD, 1992) for the design of small-scale field research involving transgenic plants and micro-organisms. It describes the use of confinement in field tests. Confinement includes measures to avoid the dissemination or establishment of organisms from a field trial, for example, the use of physical, temporal or biological isolation (such as the use of sterility).

Scale-up of crop-plants – "risk/safety analysis"

By 1993, the focus of attention had switched to the scale-up of crop plants as plant breeders began to move to larger scale production and commercialisation of transgenic plants. The OECD published general principles for scale-up, which reaffirmed that:

> *…safety in biotechnology is achieved by the appropriate application of risk/safety analysis and risk management. Risk/safety analysis comprises hazard identification and, if a hazard has been identified, risk assessment. Risk/safety analysis is based on the characteristics of the organism, the introduced trait, the environment into which the organism is introduced, the interaction between these and the intended*

> *application. Risk/safety analysis is conducted prior to an intended action and is typically a routine component of research, development and testing of new organisms, whether performed in a laboratory or a field setting. Risk/safety analysis is a scientific procedure which does not imply or exclude regulatory oversight or imply that every case will necessarily be reviewed by a national or other authority.* (OECD, 1993)

The role of familiarity in risk/safety assessment

The issue of scale-up also led to an important concept – familiarity – which is one key approach that has been used subsequently to address the environmental safety of transgenic plants.

The concept of familiarity is based on the fact that most genetically engineered organisms are developed from organisms such as crop plants, whose biology is well understood. It is not a risk/safety assessment in itself (US-NAS, 1989). However, the concept facilitates risk/safety assessments, because to be familiar means having enough information to be able to make a judgement of safety or risk (US-NAS, 1989). Familiarity can also be used to indicate appropriate management practices, including whether standard agricultural practices are adequate or whether other management practices are needed to manage the risk (OECD, 1993). Familiarity allows the risk assessor to draw on previous knowledge and experience with the introduction of plants and micro-organisms into the environment and this indicates appropriate management practices. As familiarity depends also on the knowledge about the environment and its interaction with introduced organisms, the risk/safety assessment in one country may not be applicable in another country. However, as field tests are performed, information will accumulate about the organisms involved, and their interactions with a number of environments.

Familiarity comes from the knowledge and experience available for conducting a risk/safety analysis prior to scale-up of any new plant line or crop cultivar in a particular environment. For plants, for example, familiarity takes account of, but need not be restricted to, knowledge and experience with the following:

- "The crop plant, including its flowering/reproductive characteristics, ecological requirements, and past breeding experiences.
- The agricultural and surrounding environment of the trial site.
- Specific trait(s) transferred to the plant line(s).
- Results from previous basic research including greenhouse/glasshouse and small-scale field research with the new plant line or with other plant lines having the same trait.
- The scale-up of lines of the plant crop varieties developed by more traditional techniques of plant breeding.
- The scale-up of other plant lines developed by the same technique.
- The presence of related (and sexually compatible) plants in the surrounding natural environment, and knowledge of the potential for gene transfer between crop plant and the relative.
- Interactions between/among the crop plant, environment and trait". (OECD, 1993)

Risk/safety assessment and risk management

Risk/safety assessment involves the identification of potential environmental adverse effects or hazards, and when a hazard is identified, determining the probability of

it occurring. If a potential hazard or adverse effect is identified, measures may be taken to minimise or mitigate it. This is risk management. Absolute certainty, or "zero risk", in a safety assessment is not achievable, so uncertainty is an inescapable aspect of all risk assessment and risk management (OECD, 1993). For example, there is uncertainty in extrapolating the results of testing in one species to identify potential effects in another. Risk assessors and risk managers thus spend considerable effort to address uncertainty. Many of the activities in intergovernmental organisations, such as the OECD, address ways to handle uncertainty (OECD, 2000).

References

OECD (2000), "Report of the Working Group on Harmonisation of Regulatory Oversight in Biotechnology", prepared for the G8 Summit held in Okinawa, Japan on 21-23 July 2000, (2000)86/ADD2, OECD, Paris, www.oecd.org/chemicalsafety/biotrack/Report-of-the-Working-Group-on-Harmonisation-of-Regulatory.pdf.

OECD (1993), *Safety Considerations for Biotechnology: Scale-up of Crop Plants*, OECD, Paris, www.oecd.org/env/ehs/biotrack/1958527.pdf.

OECD (1992), *Safety Considerations for Biotechnology – Part Two: Good Developmental Principles (GDP)*, OECD, Paris, www.oecd.org/sti/biotech/2375496.pdf.

OECD (1986), *Recombinant DNA Safety Considerations. Safety Considerations for Industrial, Agricultural and Environmental Applications of Organisms Derived by Recombinant DNA Techniques* ("The Blue Book"), OECD, Paris, www.oecd.org/env/ehs/biotrack/Recombinant-DNA-Safety-Considerations.pdf.

US-NAS (1989), *Field Testing of Genetically Modified Organisms: Framework for Decisions*, National Research Council, Committee on Scientific Evaluation of the Introduction of Genetically Modified Microorganisms and Plants into the Environment, National Academy Press, Washington, DC, www.nap.edu/catalog/1431/field-testing-genetically-modified-organisms-framework-for-decisions.

BIOLOGY OF MOSQUITO *AEDES AEGYPTI*

Chapter 1. **Taxonomy, description and distribution of the mosquito *Ae. aegypti***

This chapter presents the taxonomic classification, nomenclature and systematics of the mosquito species Aedes aegypti *and its two sub-species. Then the morphologic features of* Ae. aegypti *are described at successive stages: Eggs, Larvae (including differences with other mosquito genera), Pupae (showing sexual dismorphism), and Adults that present distinct characteristics of head, thorax and abdomen between male and female individuals. Elements on the origin of mosquito* Ae. aegypti, *and its current geographic distribution in tropical and subtropical regions of the world, are also provided.*

Classification and nomenclature of *Aedes aegypti*

Classification (Taxonomy)

The family Culicidae is divided into three subfamilies: Toxorhynchitinae, Anophelinae and Culicinae, within which only subfamilies Anophelinae and Culicinae have medically-important mosquito species. The subfamily Culicinae includes over 3 050 species, belonging to 109 genera, of which the most important regarding health issues are the genera *Aedes*, *Culex*, *Mansonia*, *Haemagogus*, *Sabethes*, and *Psorophora* (Service, 2012; Tyagi, Munirathinam and Venkatesh, 2015).

The systematic classification of *Aedes aegypti* is presented in Table 1.1 and localises this species within the order Diptera, family Culicidae, subfamily Culicinae, tribe Aedini, genus *Aedes*, subgenus *Stegomyia*, and species *Aedes aegypti* (ITIS, 2014; WRBU, 2014).

Table 1.1. Standardised taxonomic hierarchy and nomenclature for *Ae. aegypti* (Linnaeus, 1762)

TAXON	NOMENCLATURE (Authority)
Kingdom	Animalia (Margulis and Schwartz, 1998)
Subkingdom	Bilateria (Hatschek, 1888)
Infrakingdom	Protostomia (Grobben, 1908)
Superphylum	Ecdysozoa (Aguinaldo et al., 1997)
Phylum	Arthropoda (Latreille, 1829)
Subphylum	Hexapoda (Latreille, 1825)
Class	Insecta (Linnaeus, 1758)
Subclass	Pterygota (Lang, 1888)
Infraclass	Neoptera (Martynov, 1923)
Superorder	Endopterygota (Sharp, 1898)
Order	Diptera (Linnaeus, 1758)
Suborder	Nematocera (Berthold, 1827)
Infraorder	Culicomorpha (Wood and Borkent, 1989)
Family	Culicidae (Stephens, 1829)
Subfamily	Culicinae (Meigen, 1818)
Tribe	Aedini (Neveu-Lemaire, 1902)
Genus	*Aedes* (Meigen, 1818)
Subgenus	*Stegomyia* (Theobald, 1901)
Species	*Aedes aegypti* (Linnaeus, 1762)

Source: ITIS (2014), *Aedes aegypti*, Integrated Taxonomic Information System (database), www.itis.gov/servlet/SingleRpt/SingleRpt?search_topic=TSN&search_value=126240; WRBU (2014), Mosquito Classification Comparison, 2013, The Walter Reed Biosystematics Unit.

Subspecies. Human population increase and extension to wild habitats, in addition to the evolution of vector behaviour, are important phenomena that greatly influence the "domestication" and the constitution of subpopulations of many mosquitoes (Powell and Tabachnick, 2013). *Ae. aegypti* presents two subspecies or subpopulations:

- The first subspecies, *Ae. aegypti formosus*, is the ancestor of the domestic form of *Ae. aegypti* and still lives in forests and vegetated ecotones in sub-Saharan Africa (Lounibos, 1981). In addition to its attraction to tree holes for breeding habitats and egg laying, it has a preference for non-human blood as sources of blood meals (required by females for egg production) and feeds on wild animals. Morphologically, this form is much darker than the form adapted to human habitats (McClelland, 1974).
- The second subspecies, *Ae. aegypti aegypti* (often designated by the shorter name *Ae. aegypti*), is found globally in tropical and subtropical regions, typically in association with humans, but is absent from the interior of Africa south of the Sahara (Moore et al., 2013; Powell and Tabachnick, 2013). In contrast to the first subspecies, *Ae. aegypti aegypti* predominantly breeds in artificial containers provided by humans, also breeds indoors, and has a preference for feeding on human blood (Moore et al., 2013).

A third subspecies was previously thought to exist, *Ae. aegypti queenslandensis*, described as a light-coloured form found in the Mediterranean Basin (Mattingly, 1967). However, recent analysis suggests that *Ae. aegypti queenslandensis* is genomically identical to the second subspecies *Ae. aegypti aegypti* (Rašić et al., 2016).

Nomenclature

Common names. The usual common name for *Ae. aegypti* is the "yellow fever mosquito", as it is a principal vector for yellow fever. The closely-related species *Ae. albopictus* is often referred to as "Asian tiger mosquito". In colloquial language, "tiger mosquito" is sometimes used for naming both species indistinctly, drawn from the observation of their striped-colour abdomen.

Synonyms. If two or more names are found to apply to the same species, they are considered synonyms. The name *Ae. aegypti* (Linnaeus, 1762) is now in general use and has been for more than five decades. However, this species has appeared under many other names in the past, among the most cited are (ITIS, 2014; WRBU, 2014):

- *Culex aegypti* (Linnaeus, 1762)
- *Culex excitans* (Walker, 1848) and
- *Culex taeniatus* (Weidemann, 1828).

Recent studies have resulted in a number of generic and subgeneric changes to the classification of the tribe Aedini in Europe and other regions of the world. Among other changes, the subgenus *Stegomyia* was elevated to the category of genus for the species *Ae. aegypti* and *Ae. albopictus* (*Stegomyia aegypti* and *St. albopicta*, respectively) (Reinert and Harbach, 2005). In practice, it is rarely called *St. aegypti* and is still commonly referred to as *Ae. aegypti*.

Systematics

Ae. aegypti and *Ae. albopictus* populations seem to have different evolutionary histories, the former originated from Africa and the latter from South-East Asia. For *Ae. aegypti*, the general structure of the phylogenetic trees based on mitochondrial genes showed that most populations from South America were found to be genetically similar to populations from South-East Asia (Thailand and Viet Nam), except for one sample from Boa Vista (northern Amazonia), which was more closely related to samples from Africa (Côte d'Ivoire and Guinea). This suggests that African populations of *Ae. aegypti* introduced during the slave trade have persisted in Boa Vista, resisting eradication campaigns (Mousson et al., 2005).

Over the past 50 years, many population genetic studies of *Ae. aegypti* have documented large genetic differences among worldwide populations. Phylogenetic analyses, including through studies involving population genetics of *Ae. aegypti* s.1. using mitochondrial DNA markers, have shown that global collections fell into two clades (Tabachnick and Powell, 1979; Powell, Tabachnick and Arnold, 1980; Tabachnick, 1982, 1991; Lorenz et al., 1984; Wallis, Tabachnick and Powell, 1984; Tabachnick et al., 1985; Muñoz et al., 2013; Moore et al., 2013). One clade contained *Ae. aegypti* from East Africa, South America and the Caribbean, suggesting that these New World populations were derived directly from East African populations. The other clade contained Asian and south-eastern United States *Ae. aegypti*, along with a basal branch containing subspecies *Ae. aegypti formosus* from both East and West Africa, suggesting an independent introduction of *Ae. aegypti* to Asia (Moore et al., 2013; Powell and Tabachnick, 2013). Further support for the existence of two principal clades worldwide is provided from studies in Africa (Brown et al., 2011; Delatte et al., 2011) as well as the New World (Bracco et al., 2007; Scarpassa, Cardoza and Cardoso Junior, 2008).

Morphology

Morphologic features have been used in many studies to describe variations among populations of the same species. Morphological characteristics of *Ae. aegypti* life stages are described in greater detail in the following sub-sections.

Eggs

Eggs of *Ae. aegypti* are long, smooth, more or less ovoid shaped, and approximately 1 mm long. They are white in colour when freshly laid but turn black as a result of melanisation about two hours after oviposition (this colour change is not exclusive to *Aedes* mosquito species) (Nelson, 1986; Service, 2012).

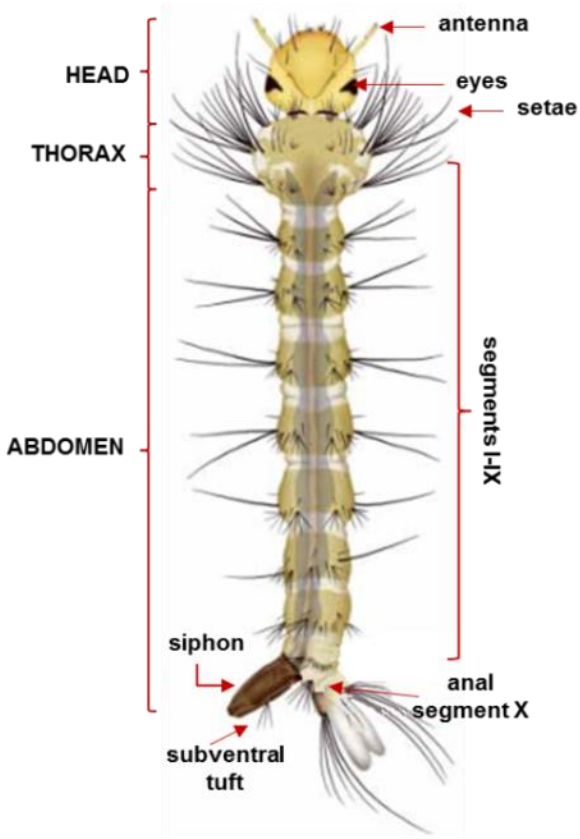
Aedes females lay individual eggs in artificial collections of water, often placed at varying distances from the water line. In addition, a female will preferably not lay the entire clutch at a single site, but rather spread the eggs over two or more sites in a practice known as "skip oviposition". Thus, the eggs stand a better chance of survival (Mogi and Mokry, 1980; Chadee, 1997; Harrington and Edmann, 2001; Foster and Walker, 2002). It was observed that eggs may be laid on successive occasions on the same site (Gillet, 1962) or in different sites (Fay and Perry, 1965; Chadee and Corbet, 1987). The practice of skip oviposition indicates the tendency of a female to avoid laying on surfaces that already bear her own eggs or those of conspecifics (Chadee, Corbet and Greenwood, 1990).

Ae. aegypti eggs can dry, survive desiccation, remain intact for several months and hatch when submerged with water. More details relating to their survival under different temperature and humidity conditions are given under the "Life cycle" section in Chapter 2.

Larvae

Ae. aegypti larvae resemble other mosquito larvae in their morphology; in general, they have an ovoid head, thorax, and abdomen of nine segments. The posterior segment (anal) has four lobed gills for osmotic regulation and a short barrel-shaped siphon bearing a single pair of subventral tufts for breathing at the water surface (Figure 1.1) (Nelson, 1986; Clements, 2000; Service, 2012). Additional morphologic characteristics include at least three pairs of setae in the ventral brush, antennae that are not greatly flattened, and a lack of enormous setae on the thorax. These characteristics are sufficient in distinguishing *Aedes* larvae from most others belonging to family Culicidae and subfamily Culicinae (Service, 2012).

Figure 1.1. Dorsal view of *Ae. aegypti* larva

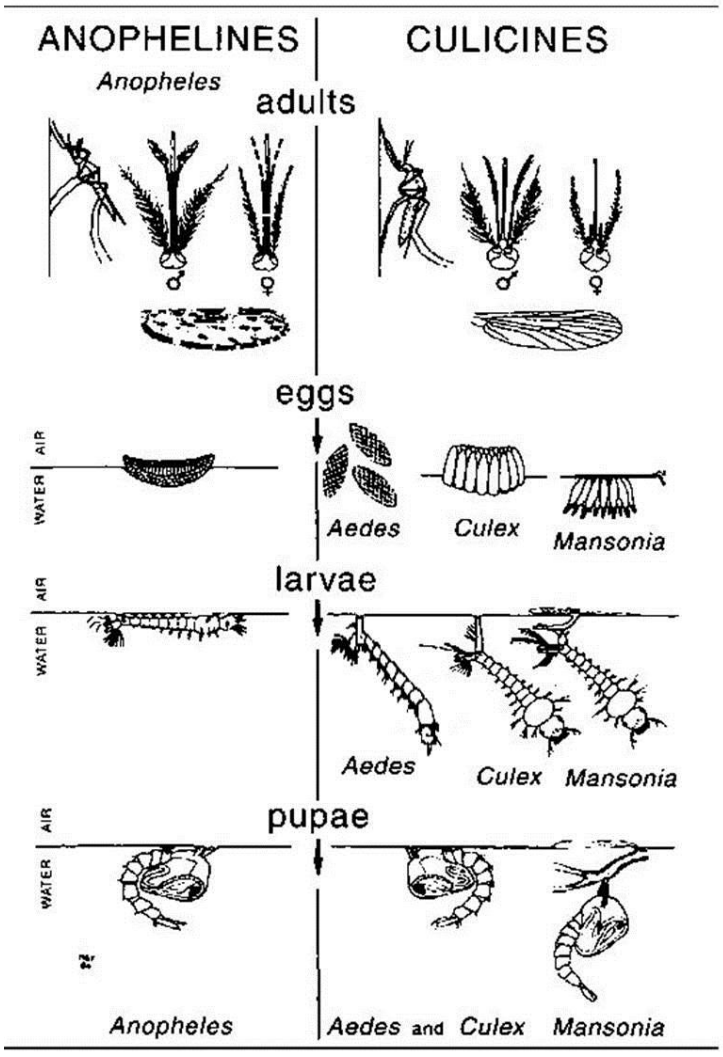


Source: Modified from Rueda, L. (2004), "Pictorial keys for the identification of mosquitoes (Diptera: Culicidae) associated with dengue virus transmission", in *ZOOTAXA 589*, Magnolia Press, Auckland, pp. 60.

The resting position at the water surface is also different among the various mosquito species: *Anopheles* larvae lay parallel to the water surface, *Culex* larvae rest at an angle and *Aedes* larvae hang almost vertically (Figure 1.2). The larvae pass through four instars (I, II, III, and IV respectively) with growth and changes in form and size occurring during

their development. The first instar *Ae. aegypti* larva is only about 1 mm in length, whereas in the fourth instar stage it reaches a length of approximately 8 mm (Schaper and Hernandez-Chavarria, 2006; Bar and Andrew, 2013a). Growth and development of larval instars is temperature dependent, however, complex interactions with other factors such as resource availability and intraspecific density also contribute to variation in development rate (Couret and Benedict, 2014). At cool environmental temperatures (around 15°C), *Ae. aegypti* larvae can remain in a particular instar for months, so long as the water supply is sufficient (Foster and Walker, 2002; Bar and Andrew, 2013a; Brady et al., 2013).

Figure 1.2. Comparison of the adults, eggs, larvae and pupae of mosquito genera *Anopheles, Aedes, Culex* and *Mansonia*

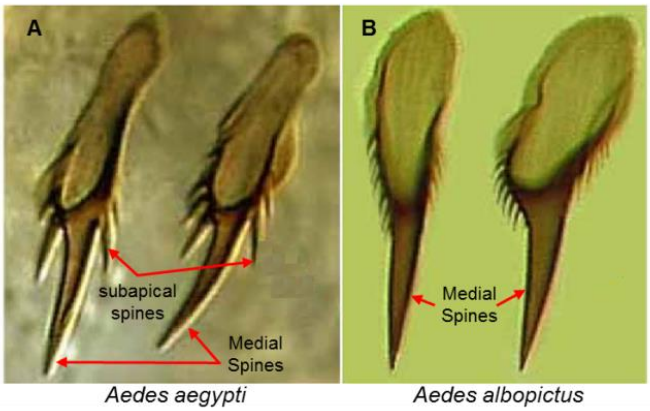


Source: Modified from Warrell, D.A. and H.M. Gilles (eds.) (2002), *Essential Malariology, 4th Ed.*, Hodder Arnold, London, pp. 350.

The most distinguishing characteristics facilitating the differentiation of *Ae. aegypti* larvae from many other species of the *Aedes* genus are the 2 lateral spines on each side of

the thorax and the straight row of 7 to 12 comb scales on the 8th abdominal segment. *Ae. aegypti* exhibits a medial spine with stout, subapical spines (Figure 1.3, panel A) which are absent in *Ae. albopictus* (Figure 1.3, panel B) (Nelson, 1986).

Figure 1.3. Comb scales of *Ae. aegypti* exhibiting a medial spine with stout, subapical spines and of *Ae. albopictus* without subapical spines



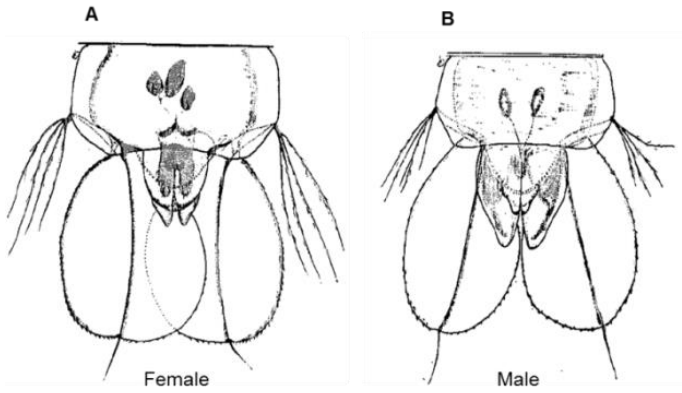
Source: Modified from Rueda, L. (2004), "Pictorial keys for the identification of mosquitoes (Diptera: Culicidae) associated with dengue virus transmission", in *ZOOTAXA 589*, Magnolia Press, Auckland, pp. 60.

Pupae (sexual dimorphism)

The pupa is the stage of the life cycle of mosquitoes that follows the last larval instar and precedes the adult stage. Pupae are comma-shaped, composed of two main sections, cephalothorax (head and thorax fused) and abdomen (Nelson, 1986; Service, 2012). At the base of the cephalothorax of the pupa is a pair of breathing tubes or "trumpets" that pierce the water surface to allow breathing (Nelson, 1986). At the tip of the abdomen there is a pair of oars or paddles used for swimming, which in the female (Figure 1.4, panel A) are wider and overlap, but in the male (Figure 1.4, panel B) are narrow and separated (Vargas, 1968).

Another morphologic difference between female and male pupae is their overall size, with the female usually being larger than the male (Figure 1.4). Since the range in body size between female and male pupae overlaps considerably and can be affected by both biotic and abiotic, including environmental factors such as diet, temperature, rearing conditions, overcrowding, it is deemed necessary to select additional sexually dimorphic characteristics such as the differences in paddles in order to determine the sex of pupae (Vargas, 1968).

Figure 1.4. Anal segments of *Ae. aegypti* pupae - ventral view, showing dimorphism characters between females and males

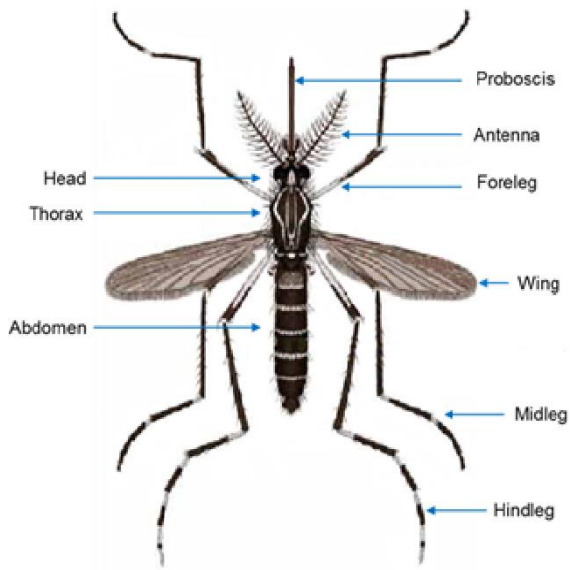


Source: Modified from Vargas, V.M. (1968), "Sexual dimorphism of larvae and pupae of *Ae. aegypti* (Linn.)", *Mosquito News*, Vol. 28, pp. 374-379.

Adults (male and female)

The body of an adult *Ae. aegypti* mosquito is composed of head, thorax, and abdomen (Figure 1.5). *Ae. aegypti* males and females are similar in appearance except for the differences in size and form of the antennae (males have plumose antennae), maxillary palps (females have shorter palps), abdomen, claws and in scale markings (Bar and Andrew, 2013b). These differences are described in detail below.

Figure 1.5. Dorsal view of the female mosquito *Ae. aegypti*



Source: Modified from Rueda, L. (2004), "Pictorial keys for the identification of mosquitoes (Diptera: Culicidae) associated with dengue virus transmission", in *ZOOTAXA 589*, Magnolia Press, Auckland, pp. 60.

Head

In both male and female *Ae. aegypti*, dorsally the head is globular in shape and laterally convex with a vertex that has silvery-white flat scales. The female clypeus has two silvery white dots, whereas the male has no dots. Females have a larger head capsule (0.55 ± 0.09 mm) than males (0.53 ± 0.06 mm) (Bar and Andrew, 2013b). The head bears several structures critical to the mosquito's ability to feed as well as to act as a vector of human diseases.

Mouthparts. The mouthparts in these mosquitoes include a pair of maxillary palps, which have five white scale bands and are longer (0.77 ± 0.06 mm) and more developed in males than in females (0.53 ± 0.06 mm) (Bar and Andrew, 2013b). The proboscis is longer in males (0.76 ± 0.04 mm) than in females (0.66 ± 0.03 mm) (Bar and Andrew, 2013b). However, only in females is this structure adapted for skin penetration to enable blood feeding, even though they may survive in nature by sucking plant juices. The male proboscis is adapted to feed on nectar and plant juices rich in carbohydrates (Clements, 1992).

Antenna. Each antenna of *Ae. aegypti* arises from a globular pedicel, has 13 flagellar segments and a greatly reduced scape. Males have longer antennae (0.57 ± 0.03 mm) than females (0.52 ± 0.07 mm). The antennal hairs are bushy and plumose in males whereas in females they are smaller and less dense (Nelson, 1986; Bar and Andrew, 2013b).

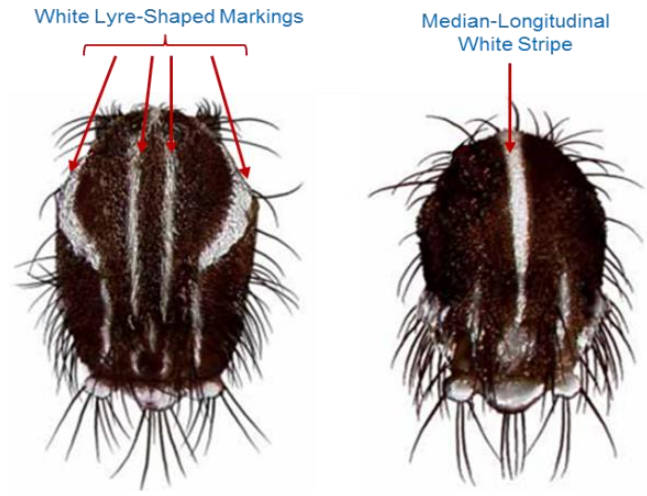
Thorax

Females of *Ae. aegypti* have a larger thorax measuring 0.50 ± 0.08 mm in length and 0.35 ± 0.07 mm in width while the shorter male thorax is 0.41 ± 0.06 mm in length and 0.29 ± 0.02 mm in width. The thorax of *Ae. aegypti* is black or dark brown coloured and consists of the pro-, meso-, and metathoracic segments, which together bear the wings (one pair), legs (three pairs), and halteres (one pair) (Bar and Andrew, 2013b).

Many, but not all, *Aedes* adults have conspicuous patterns on the thorax formed by white or silver coloured scales (Service, 2012), and these patterns vary between species. An example of the difference across species is the case of *Ae. aegypti* with its typical, white, lyre-shaped markings (Figure 1.6, panel A), compared to *Ae. albopictus* with its median-longitudinal white stripe (Figure 1.6, panel B) (Nelson, 1986). The scutellum in *Ae. aegypti* is three-lobed with each lobe having silvery white scale patches, and a few dark scales at the apex of the midlobe (Bar and Andrew, 2013b).

Figure 1.6. Comparative dorsal view of thoracic scutum of *Ae. aegypti* and *Ae. albopictus*

Figure 1.6A: *Ae. aegypti* Figure 1.6B: *Ae. albopictus*



Source: Modified from Rueda, L. (2004), "Pictorial keys for the identification of mosquitoes (Diptera: Culicidae) associated with dengue virus transmission", in *ZOOTAXA 589*, Magnolia Press, Auckland, pp. 60.

At the same time, adults of *Aedes* and other Culicinae may be distinguished from adult *Anopheles* mosquitoes by their shorter palps and their resting position which is more horizontal or parallel to the resting surface (Nelson, 1986; Service, 2012).

Abdomen

The abdomen consists of eight segments covered with black and white scales forming distinctive patterns in both males and females. In females, the eighth segment is greatly reduced. The tergites (dorsal portion of each abdominal segment) are dark brown in colour and the first abdominal segment has a patch of pale, median scales. The dorsal side of abdominal segments II through VII has transverse white bands. The size of abdomen in males is larger (length 3.03 ± 0.18 mm and width 0.51 ± 0.07 mm) than in females (length 2.94 ± 0.20 mm and width 0.41 ± 0.06 mm) (Bar and Andrew, 2013b).

The posterior tip of the abdomen is narrow in males while in females it has a broad round shape. *Ae. aegypti* can be differentiated from most of the other Culicinae by their pointed abdomen and the absence of spiracular bristles (Service, 2012).

With age, the lyre-shaped markings on the thorax may disappear, but the distinctive white scales on the pedicel, clypeus, and tip of the palps, and the pattern of white scales on abdominal sternites (ventral plate on each abdominal segment) III-V, usually remain. These characteristics are essential for the identification of *Ae. aegypti* females with damaged morphological structures and to differentiate them from *Ae. albopictus* females (Nelson, 1986; Savage and Smith, 1995).

Origin and current geographic distribution

The likely origin of *Ae. aegypti* is the Ethiopian region of the tropical belt in Africa, from which it has spread to tropical and subtropical regions throughout the world in association with humans (Nelson, 1986; Powell and Tabachnick, 2013). *Ae. aegypti* was probably carried to other continents via trading and transport ships that resupplied in African ports

during the 15th century through to the end of the 17th (Christophers, 1960; Reiter, 1998). These ships carried freshwater reservoirs on board and could maintain breeding colonies of *Ae. aegypti* (Christophers, 1960), so it is probable that the species was introduced to the rest of the world via this means (Tabachnick, 1991).

To date, *Ae. aegypti* is an invasive tropical species worldwide with a cosmopolitan habitat from 40° N to 40° S latitude (a range extending across all or most of the world in appropriate habitats).

Ae. aegypti is usually tolerant to temperatures ranging from 14°C to 30°C (Hemme et al., 2010; Brady et al., 2013, 2014). Under optimal conditions of temperature and humidity, the embryo needs two to three days for full development from oviposition to the next stage of the life cycle. The definition of physiological embryonic parameters within this temperature range correlates with the presence of *Ae. aegypti* in tropical and subtropical regions of the world (Farnesi et al., 2009). Larval development in *Ae. aegypti* is a function of temperature, and these effects have been well studied. Temperature also impacts on adult size, dry weight, and ovariole number, all of which decrease as the temperature increases (Christophers, 1960; Rueda et al., 1990). High extreme temperatures alone (> 40°C) are unlikely to limit the species, but low temperatures are a limiting factor. Below 15°C, adult *Ae. aegypti* mosquitoes become torpid, unable to fly, and can move their limbs only slowly (Christophers, 1960; Rowley and Graham, 1968; Yang et al., 2009). Lower temperatures can slow development to such a degree (where egg-to-adult cycles are longer than 45 days) that the species is prevented from establishing itself in the environment, although human habitations may afford some seasonal protection.

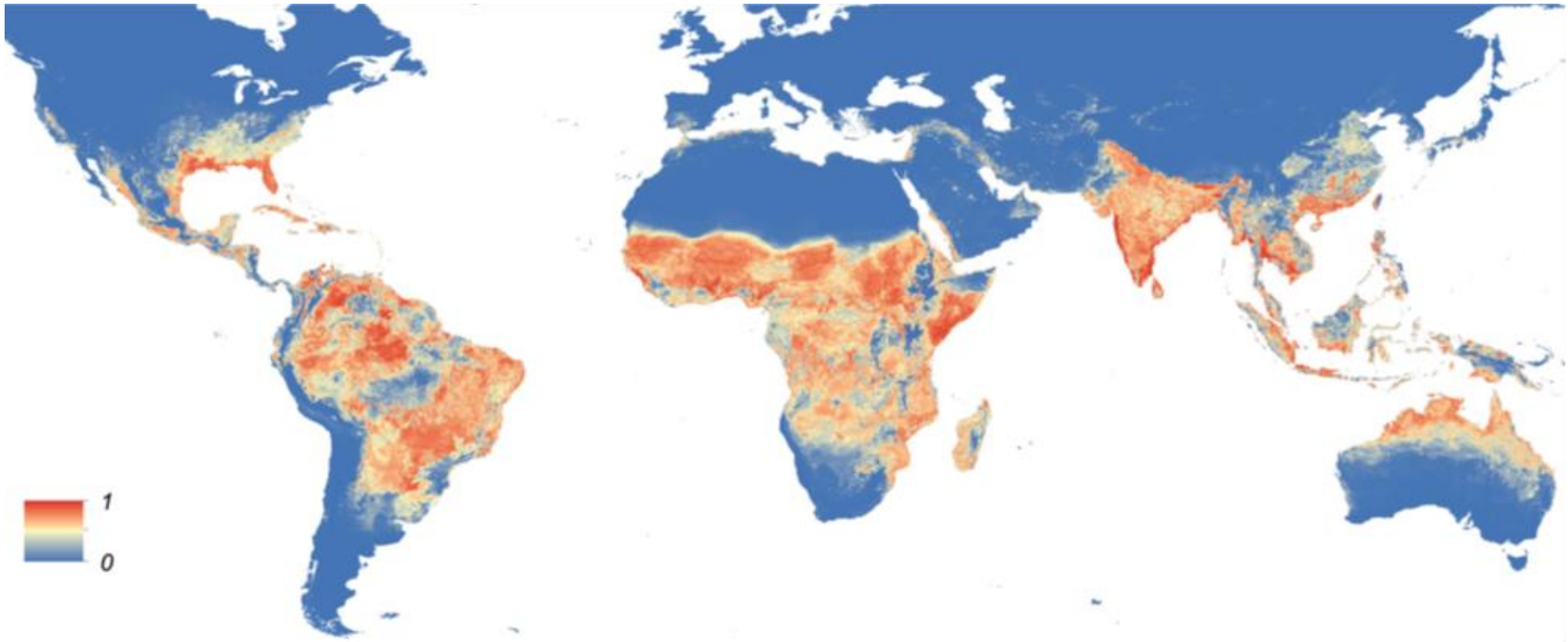
Rain quantity and frequency (precipitation level) is another factor which, combined with temperature, affects the sustainable establishment of the species in a given area.

Global historical collections and laboratory experiments on this well-studied vector have suggested its distribution is limited by the 10°C winter isotherm[1] (Christophers, 1960), while a more recent and complex stochastic population dynamics model analysis suggests the temperature's limiting value to be more towards the 15°C yearly isotherm (Otero, Solari and Schweigmann, 2006). Scholte et al. (2010) indicated that *Ae. aegypti* could not survive winter temperatures in Northern Europe. The predicted global distribution of *Ae. aegypti*, based on occurrence data as well as environmental and land-cover variables, is shown in Figure 1.7 (Kraemer et al., 2015).

Notes

[1] An isotherm is a line on a map or chart of the earth's surface connecting points having the same temperature at a given time or the same mean temperature for a given period.

Figure 1.7. Global map of the predicted distribution of *Ae. aegypti*



Note: The map depicts the probability of occurrence.
• Blue (dark grey) = 0
• Red (light grey) = 1

Source: Kraemer, M.U.G. et al. (2015), "The global distribution of the arbovirus vectors *Aedes aegypti* and *Ae. albopictus*", *eLife*, Vol. 4: e08347.

References

Bar, A. and J. Andrew (2013a), "Morphology and morphometry of *Aedes aegypti* larvae", *Annual Review and Research in Biology*, Vol. 3, pp. 1-21.

Bar, A. and J. Andrew (2013b), "Morphology and morphometry of *Aedes aegypti* adult mosquito", *Annual Review and Research in Biology*, Vol. 3, pp. 52-69.

Bracco, J.E. et al. (2007), "Genetic variability of *Aedes aegypti* in the Americas using a mitochondrial gene: Evidence of multiple introductions", *Memórias do Instituto Oswaldo Cruz* , Vol. 102, No. 5, pp. 573-580.

Brady, O.J. et al. (2014), "Global temperature constraints on *Aedes aegypti* and *Ae. albopictus* persistence and competence for dengue virus transmission", *Parasites and Vectors*, Vol. 7, pp. 338.

Brady, O.J. et al. (2013), "Modelling adult *Aedes aegypti* and *Aedes albopictus* survival at different temperatures in laboratory and field settings", *Parasit Vectors*, Vol. 6, pp. 351.

Brown, J.E. et al. (2011), "Worldwide patterns of genetic differentiation imply multiple 'domestications' of *Aedes aegypti*, a major vector of human diseases", *Proceedings of the Royal Society B: Biological Sciences*, Vol. 278, pp. 2446–2454.

Chadee, D.D. (1997), "Effects of forced egg-retention on the oviposition patterns of female *Aedes aegypti* (Diptera: Culicidae)", *Bulletin of Entomological Research*, Vol. 87, pp. 649-651.

Chadee, D.D. and P.S. Corbet (1987), "Seasonal incidence and diel patterns of oviposition in the field of the mosquito, *Aedes aegypti* (L.) (Diptera: Culicidae) in Trinidad, West Indies: A preliminary study", *Annals of Tropical Medicine and Parasitology*, Vol. 81, pp. 151-161.

Chadee, D.D., P.S. Corbet and J.J.D. Greenwood (1990), "Egg-laying Yellow Fever Mosquitoes avoid sites containing eggs laid by themselves or by conspecifics", *Entomology Experimental Applied*, Vol. 57, pp. 295-298.

Christophers, S.R. (1960), Aedes aegypti *(L.) The Yellow Fever Mosquito. Its Life History, Bionomics and Structure*, Cambridge University Press, Cambridge.

Clements, A.N. (2000), *The Biology of Mosquitoes, Volume I: "Development, Nutrition and Reproduction" Second Edition*, CABI Publishing, Oxford.

Clements, A.N. (1992), *The Biology of Mosquitoes, Volume I "Development, Nutrition and Reproduction"*, Chapman and Hall, London.

Couret, J. and M.Q. Benedict (2014). "A meta-analysis of the factors influencing development rate variation in *Aedes aegypti* (Diptera: Culicidae)", *BioMed Central Ecology*, Vol. 14, No. 3, pp 1-15.

Delatte, H. et al. (2011), "The invaders: Phylogeography of dengue and chikungunya viruses *Aedes* vectors, on the South West islands of the Indian Ocean*", Infection, Genetics and Evolution*, Vol. 11, No. 7, pp. 1769-1781.

Farnesi, L.C. et al. (2009), "Embryonic development of *Aedes aegypti* (Diptera: Culicidae): Influence of different constant temperatures", *Memórias do Instituto Oswaldo Cruz*, Vol. 104, No. 1, pp. 124-126.

Fay, R.W. and A.S. Perry (1965), "Laboratory studies of ovipositional preferences of *Aedes aegypti*", *Mosquito News*, Vol. 25, pp. 276-281.

Foster, W.A. and E.D. Walker (2002), "Mosquitoes (Culicidae)", in G. Mullen and L. Durden (eds.), *Medical and Veterinary Entomology*, Academic Press, San Diego, pp. 203-262.

Gillett, J.D. (1962), "Contributions to the oviposition cycle by individual mosquitoes in a population", *Journal of Insect Physiology*, Vol. 8, pp. 665-681.

Harrington, L. and J.D. Edmann (2001), "Indirect evidence against delayed "Skip-Oviposition" behavior by *Aedes aegypti* (Diptera: Culicidae) in Thailand", *Journal of Medical Entomology*, Vol. 38, pp. 641-645.

Hemme, R.R. et al. (2010), "Influence of urban landscapes on population dynamics in a short-distance migrant mosquito: Evidence for the dengue vector *Aedes aegypti*", *PLoS Neglected Tropical Diseases*, Vol. 4, No. 3: e634.

ITIS (2014), *Aedes aegypti*, Integrated Taxonomic Information System (database), www.itis.gov/servlet/SingleRpt/SingleRpt?search_topic=TSN&search_value=126240.

Kraemer, M.U.G. et al. (2015), "The global distribution of the arbovirus vectors *Aedes aegypti* and *Ae. albopictus*", *eLife*, Vol. 4: e08347.

Lorenz, L. et al. (1984), "The effect of colonization upon *Aedes aegypti* - Susceptibility to oral infection with yellow-fever virus", *American Journal of Tropical Medicine and Hygiene*, Vol. 33, pp. 690-694.

Lounibos, L.P. (1981), "Habitat segregation among African treehole mosquitoes", *Ecological Entomology*, Vol. 6, pp. 129-154.

Mattingly, P.F. (1967), "Taxonomy of *Aedes aegypti* and related species", *Bulletin of the World Health Organization*, Vol. 36, No. 4, pp. 552-554.

McClelland, G.A.H. (1974), "A worldwide survey of variation in scale pattern of the abdominal tergum of *Aedes aegypti* (L.) (Diptera: Culicidae)", *Transaction Royal Entomological Society London*, Vol. 126, pp. 239-259.

Mogi, M. and J. Mokry (1980), "Distribution of *Wyeomyia smithii* (Diptera: Culicidae) eggs in pitcher plants in Newfoundland, Canada", *Tropical Medicine*, Vol. 22, pp. 1-12.

Moore, M. et al. (2013), "Dual African origins of global *Aedes aegypti* s.l. populations revealed by mitochondrial DNA", *PLoS Neglected Tropical Diseases*, Vol. 7, No. 4: e2175.

Mousson, L. et al. (2005), "Phylogeography of *Aedes (Stegomyia) aegypti* (L.) and *Aedes (Stegomyia) albopictus* (Skuse) (Diptera: Culicidae) based on mitochondrial DNA variations", *Genetic. Research (Camb.)*, Vol. 86, pp. 1–11.

Muñoz, M.deL. et al. (2013), "Gene flow pattern among *Aedes aegypti* populations in Mexico", *Journal of the American Mosquito Control Association*, Vol. 29, No. 1, pp. 1-18.

Nelson, M.J. (1986), Aedes aegypti*: Biology and Ecology*, Pan American Health Organization, Washington, DC, PNSP/86-63, pp. 50.

Otero, M., H.G. Solari and N. Schweigmann (2006), "A stochastic population dynamics model for *Aedes aegypti*: Formulation and application to a city with temperate climate", *Bulletin of Mathematical Biology*, Vol. 68, No. 8, pp. 1945-1974.

Powell, J.R. and W.J. Tabachnick (2013), "History of domestication and spread of *Aedes aegypti* – A review", *Memórias do Instituto Oswaldo Cruz*, Vol. 108, pp. 11-17.

Powell, J.R., W.J. Tabachnick and J. Arnold (1980), "Genetics and the origin of a vector population – *Aedes aegypti*, a case-study", *Science*, Vol. 208, pp. 1385–1387.

Rašić, G. et al. (2016), "The queenslandensis and the type form of the dengue fever mosquito (*Aedes aegypti* L.) are genomically indistinguishable", *PLoS Neglected Tropical Diseases*, Vol. 10, No. 11: e0005096.

Reinert, J.F. and R.E. Harbach (2005), "Generic changes affecting European aedine mosquitoes (Diptera: Culicidae: Aedini) with a checklist of species", *Journal of the European Mosquito Control Association, European Mosquito Bulletin*, Vol. 19, pp. 1-4.

Reiter, P. (1998), "*Aedes albopictus* and the world trade in used tires, 1988 - 1995: The shape of things to come?", *Journal of the American Mosquito Control Association*, Vol. 14, pp. 83-94.

Rowley, W.A. and C.L. Graham (1968), "The effect of temperature and relative humidity on the flight performance of female *Aedes aegypti*", *Journal of Insect Physiology*, Vol. 14, No. 9, pp. 1251-1257.

Rueda, L. (2004), "Pictorial keys for the identification of mosquitoes (Diptera: Culicidae) associated with dengue virus transmission", in *ZOOTAXA 589*, Magnolia Press, Auckland, pp. 60.

Rueda, L.M. et al. (1990), "Temperature-dependent development and survival rates of Culex quinquefasciatus and *Aedes aegypti* (Diptera. Culicidae)", *Journal of Medical Entomology*, Vol. 27, No. 5, pp. 892-898.

Savage, H.M. and G.C. Smith (1995), "*Aedes albopictus* y *Aedes aegypti* en las Américas: Implicaciones para la transmisión de arbovirus e identificación de hembras adultas dañadas" [*Aedes albopictus* and *Aedes aegypti* in the Americas: Implications for the transmission of arboviruses and identification of damaged adult females], *Boletin de la Oficina Sanitaria Panamericana*, Vol. 118, pp. 473-487.

Scarpassa, V.M., T.B. Cardoza and R.P. Cardoso Junior (2008), "Population genetics and phylogeography of *Aedes aegypti* (Diptera: Culicidae) from Brazil", *The American Journal of Tropical Medicine and Hygiene*, Vol. 78, No. 6, pp. 895-903.

Schaper, S. and F. Hernandez-Charvarria (2006), "Scanning electron microscopy of the four larval instars of the Dengue fever vector *Aedes aegypti* (Diptera: Culicidae)", *Revista de Biología Tropical*, Vol. 54, pp. 847-852.

Scholte, E. et al. (2010), "Introduction and control of three invasive mosquito species in the Netherlands, July-October 2010", *Euro Surveillance*, Vol. 15, No. 45, pii: 19710.

Service, M. (2012), *Medical Entomology for Students, 5th Ed*., Cambridge University Press, New York, pp. 303.

Tabachnick, W.J. (1991), "Evolutionary genetics and arthropod-borne disease: The yellow fever mosquito", *AmeriThe Canadian Entomologistogist*, Vol. 37, pp. 14-24.

Tabachnick, W.J. (1982), "Geographic and temporal patterns of genetic variation of *Aedes aegypti* in New Orleans", *American Journal of Tropical Medicine and Hygiene*, Vol. 31, pp. 849–853.

Tabachnick, W.J. and J.R. Powell (1979), "A world-wide survey of genetic-variation in the yellow fever mosquito, *Aedes aegypti*", *Genetical Research*, Vol. 34, No. 3, pp. 215-229.

Tabachnick, W.J. et al. (1985), "Oral infection of *Aedes aegypti* with yellow-fever virus – Geographic variation and genetic considerations", *American Journal of Tropical Medicine and Hygiene*, Vol. 34, No. 6, pp. 1219-1224.

Tyagi, B.K., A. Munirathinam and A. Venkatesh (2015), "A catalogue of Indian mosquitoes", *International Journal of Mosquito Research*, Vol. 2, No. 2, pp. 50-97.

Vargas, V.M. (1968), "Sexual dimorphism of larvae and pupae of *Ae. aegypti* (Linn.)", *Mosquito News*, Vol. 28, pp. 374-379.

Wallis, G.P., W.J. Tabachnick and J.R. Powell (1984), "Genetic-heterogeneity among Caribbean populations of *Aedes aegypti*", *American Journal of Tropical Medicine and Hygiene*, Vol. 33, pp. 492-498.

Warrell, D.A. and H.M. Gilles (eds.) (2002), *Essential Malariology, 4th Ed*., Hodder Arnold, London, pp. 350.

WRBU (2014), *Mosquito Classification Comparison, 2013*, The Walter Reed Biosystematics Unit.

Yang, H.M. et al. (2009), "Assessing the effects of temperature on the population of *Aedes aegypti*, the vector of dengue", *Epidemiology and Infection*, Vol. 137, No. 8, pp. 1188-1202.

Chapter 2. **Reproductive biology of the mosquito *Ae. aegypti***

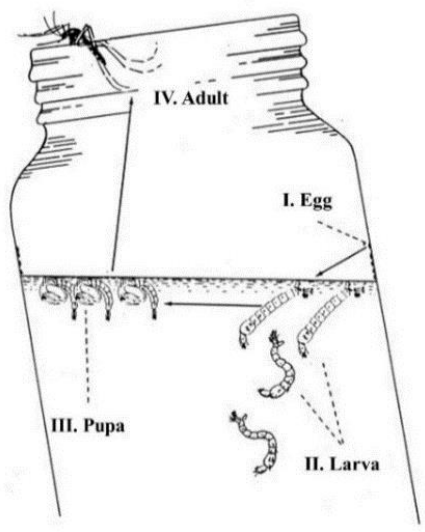
This chapter details the four life stages of mosquito Aedes aegypti *in their reproductive biology aspects. The breeding sites that can be either natural sites or artificial containers provided by human habitats. The eggs can survive dry conditions, their hatching and embryonic development depending on humidity and temperature. The larval and pupal stages are strictly aquatic, the whole development phase in water comprising four successive larval instars followed by the mobile pupae. The adult stage occurs in open-air and constitutes the reproductive and dispersal phase. The mosquito characteristics regarding mating, physiology and behaviour of reproduction, fecundity and fertility are also described. Then elements are given on* Ae. aegypti *life table analysis, interspecific breeding, and the effect of the bacteria* Wolbachia *on the mosquito reproduction.*

Life cycle

Four life stages

The life cycle of all species of mosquitoes, including *Ae. aegypti*, corresponds to the holometabolous type (Gordh, 2001) which is basically characterised by complete metamorphosis and four distinct life stages: egg; larva; pupa; and adult (Figure 2.1). The development cycle depends directly on the presence of water and ambient temperature. In warm days with temperatures averaging 25°C, development of eggs into adults is completed in a little more than 1 week. In the case of cool days, development may occur over a period of months (Foster and Walker, 2002). The stages are described below.

Figure 2.1. Life cycle of *Ae. aegypti*



Source: NCDENR (n.d.), *Mosquitoes... Some Facts: Information Pamphlet*, www.alamance-nc.com/envhealth/wp-content/uploads/sites/9/2013/10/Mosquitoes_Facts.pdf.

Breeding sites

The females lay individual eggs above the water level within fresh water held in natural breeding sites including holes in trees, bamboo trunks, hollow rocks, plant axilla, coconut shells, and leaves. Females will also oviposite on the inner wall of various artificial containers such as tanks, vases, jars, tires, drums, buckets, pots, cans, scrap metal and gutters (Nelson, 1986; Ulloa et al., 2010; Pilger et al., 2011), distributed inside houses or in their yards (Kampen and Schaffner, 2008).

The variability in the preference of the different types of containers as sites for oviposition by female *Ae. aegypti* depends on the availability of artificial containers, the degree of urbanisation and the season (Mogi and Mokry, 1980; García-Rejón et al., 2011; Rubio, Cardo and Vezzani, 2011).

Egg stage and embryonic development

Eggs can survive dry conditions for months and hatch once submerged in water, thus enhancing dissemination during the rainy periods. This survival ability of *Ae. aegypti* populations to dry seasons, combined with their intensive spread during rainy seasons, makes the control of *Ae. aegypti* very difficult (Nelson, 1986; Service, 2012).

There has been some research on the correlated effects of temperature and humidity on the eggs of *Ae. aegypti*. Experimental studies indicate that 20% of eggs remain viable after 6 months in 98% humidity (Luz et al., 2008). In Japan, Sota and Mogi (1992) measured survival times of eggs from several *Aedes* species including *Ae. aegypti* and *Ae. albopictus* under 3 different humidity conditions (42%, 68% and 88% of relative humidity) at 25°C, showing that *Ae. aegypti* survived longer than *Ae. albopictus* at all humidity conditions. Sota and Mogi (1992) attributed this to egg volume, with *Ae. aegypti* having the greatest egg volume and thus the greatest ability to resist desiccation. Juliano et al. (2002) also found the effects of temperature and humidity on egg mortality significantly different between the two species, with *Ae. albopictus* experiencing much higher mortality at all combinations except at the highest humidity. The maximum temperature limit for embryogenesis is 35°C and the minimum 12°C and below; and for egg viability optimal temperature ranges between 16-31°C, and with relative humidity above 80% (Farnesi et al., 2009). In a recent study, Thomas et al. (2012) found that eggs of a tropical strain of *Ae. aegypti* could survive at a threshold of 2°C for 24 hours only before hatching ceased. Egg survival at temperatures below freezing is therefore extremely unlikely.

The first 48 hours of embryonic development are critical and microclimatic factors are crucial for embryo survival (Thirión, 2003; Farnesi et al., 2009). The eggs of aedine mosquitoes usually enter a diapause-like state (suspension of development or quiescence) in unfavourable weather conditions (such as low temperature and humidity). They will hatch asynchronously several weeks or even months after being deposited with the return of more favourable conditions (Gillett, Roman and Phillips, 1977; Jeffery et al., 2012). In a natural setting, flooding from rainfall induces a physicochemical stimulus that results in egg hatching. Similarly, eggs are stimulated to hatch when submerged as the water level rises in water storage containers which are in everyday use (Koenraadt and Harrington, 2008). Additionally, other types of stimuli have been associated with hatching, for example, the low concentration of oxygen dissolved in water (Judson, 1960) and the presence of some water-soluble compounds or organisms in the water as a result of microbial activity (Gillett, Roman and Phillips, 1977; Ponnusamy et al., 2011).

Larval and pupal stages

The larval and pupal stages are strictly aquatic. Larval development begins with the first of four instars, each larger than the last. Passing from one larval stage to the next is accomplished by the moulting of chitinous skin that is shed, allowing growth and development of the next instar. Complete larval development typically lasts five to seven days and ends when the fourth instar larva develops and reaches the pupal form (Thirión, 2003). Larvae are omnivorous and spend most of their time feeding with the help of oral silks arranged in a fan which is used to filter particles of suspended organic matter and microorganisms in the water. They also graze organic matter on the bottom and sides of the flooded container (Colvard, 1978). The larvae feed in the water on protozoa, bacteria, yeasts and algae, both at the bottom of the habitat as well as in the water column (Ponce, 1999).

The duration of the aquatic phase of *Ae. aegypti* from first instar larvae to adult emergence, in the laboratory with water temperature at 24-27°C and no interspecific competition, is 8.42 days on average, with a range of 7.9-9.0 days. However, for both *Ae. aegypti* (Hancock et al., 2016) and *Ae. albopictus* (Sánchez-Hernández, 2011), the development time of larvae is significantly increased by competition for the limited amount of food in containers where the time to pupation can extend up to eight weeks.

Larval development is also favoured by the high prevalence of bacteria such as *Aeromonas hydrophila/caviae*, *Klebsiella oxytoca*, *Pseudomonas* sp., and *Enterobacter cloacae* in artificial breeding sites (tires, tanks, others) (Ulloa, 1996). These bacteria are potential food sources for larvae of *Ae. aegypti*. This study also revealed that discarded tires were the most important in terms of persistence in mosquito density and production of larvae. In this regard, Manrique-Saide et al. (1998) reported that the average time for immature stages of *Ae. aegypti* to develop in used tires was 11.15 to 12.95 days. Temperature, diet, density and their two-way interactions are all significant factors in explaining development rate variation of the larval stages of *Ae. aegypti* mosquitoes (Couret and Benedict, 2014).

The pupa is the last aquatic developmental stage, usually lasting between 2.0 and 3.6 days under optimal conditions (Focks et al., 1981; Nelson, 1986; Manrique-Saide et al., 1998). This stage is mobile (although non-feeding), and swims actively within the container in response to external stimuli such as vibrations and changes in light intensity.

Arrivillaga and Barrera (2004) determined the duration of the whole aquatic development phase (from first larval instar to adult) of *Ae. aegypti* in the laboratory associated with different levels of starvation for the immature stages. Development times varied between 8.5 days and 18.5 days with faster growth associated with increased food, highest water levels, and reduced density of larvae. Moreover, a comparative study between an *Ae. aegypti* wild type strain and a genetically engineered (GE) line[1] showed a shorter time of pupation for the GE line (one day on average) as compared to the wild type strain, with this difference being more pronounced for females (1.4 days) than for males (0.9 day) (Bargielowski et al., 2011).

Adult stage

The adult or imago of the genus *Aedes*, like other groups of mosquitoes, is the reproductive and dispersal stage. Emergence of adult *Ae. aegypti* is usually crepuscular with adults released from the pupal exuviae performing an initial flight to a dry, resting place. The initial 24-hour period post-emergence is the teneral period, a physiological state during which the exoskeleton hardens and sexual maturation occurs (Clements, 2000). The teneral phase results in a fully mature aerial adult capable of flight and mating. Males are the first to emerge and a balanced sex ratio is produced, although sex ratios can be skewed by the presence of other competing species (Sánchez-Hernández, 2011).

The adult life expectancy varies from 10-35 days for female mosquitoes (Goindin et al., 2015) and 3-6 days for male mosquitoes (Clements, 2000) although this is highly dependent on temperature, being shorter in tropical regions and longer in more temperate climates, etc.

The dispersal range of adults is variable and is influenced by a variety of factors including the sex of the mosquito, density of human hosts, availability of breeding sites, abundance of plants in houses, as well as composition and configuration of ecological landscape

(Reiter et al., 1995; Martinez-Ibarra et al., 1997; Rubio, Cardo and Vezzani, 2011). More information is given under Dispersal sub-section in Chapter 4.

Reproduction

Mating

Mosquitoes utilise sexual reproduction to produce new generations. Within 2-3 days after emergence, both sexes mate, and females can take a blood meal which is required for egg development (Lehane, 1991). These two activities often occur simultaneously because males are attracted to both the vertebrate host and the females, thus facilitating mating (Nelson, 1986).

The sound emitted by the flight frequency of females is used by males to locate and copulate with them (Brogdon, 1994). A source of attraction of a male to a female is the sound made by the beating of her wings during flight (Cator et al., 2009; Cator and Harrington, 2011). However, mating after engorgement of the females is rare because once the female has taken a blood meal, she must beat her wings more rapidly to carry her increased weight and the wing-beat frequency is no longer attractive to the male (Nelson, 1986; Cator et al., 2009).

During mating, the male clasps the tip of the female abdomen with his terminalia and inserts his aedeagus into the genital chamber. The female bursa copulatrix becomes filled with male sperm that passes within two minutes to the spermatheacae where they are stored prior to fertilisation of the eggs (Nelson, 1986).

Ae. aegypti females generally mate only once, since a single insemination event allows sufficient sperm to be stored within the spermatheacae to fertilise all the eggs that a female will develop during her lifetime. In addition, the seminal fluid proteins transferred from the male during mating render females unreceptive and refractory to further copulation (Sirot et al., 2008; Avila et al., 2011; Helinski et al., 2012). Thus, once mated, *Ae. aegypti* females are generally not responsive to additional matings for the duration of one or more egg-laying cycles (Cator et al., 2009). They may remate, however, if the spermatheacae is not adequately filled. Results from laboratory studies have revealed that 14% of females are involved in multiple matings (polyandry) within a 48-hour period (Helinski et al., 2012). Polyandry in a natural population of *Ae. aegypti* is low (6.25%), but also likely an underestimate and is within the range of polyandry estimates in other mosquito species (Richardson et al., 2015).

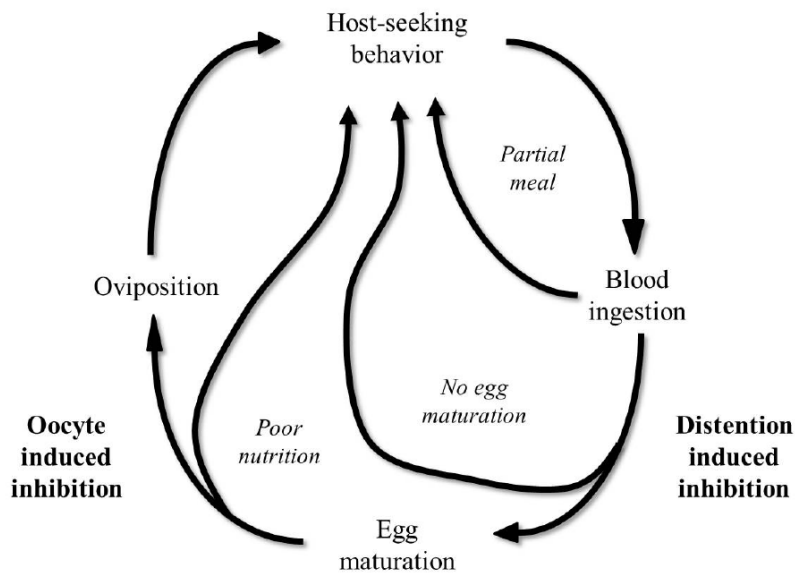
Laboratory studies have determined that the body size of male *Ae. aegypti* is a major predictor of total spermatozoa number, with significantly greater sperm numbers in larger males (2.27 mm wing length) versus smaller males (1.85 mm wing length) within the same age group (Ponlawat and Harrington, 2007). Other studies have shown that under field conditions, larger males inseminated females with more sperm than smaller ones and that older males transferred the greatest number of sperm to females (1 152 sperm by 1-day-old males compared to 1 892 sperm by 10-day-old males). At the same time, larger females successfully mated with males more often than smaller females, especially with older males (> 25-day-old) (Ponlawat and Harrington, 2009).

Physiology of reproduction

From the biological point of view, the physiological condition and the time required for females to carry out the digestion of a blood meal, maturation of the follicles, and

subsequent oviposition constitutes a strategy of reproductive competition; a strategy for competition between females for resources required for reproduction (Wheeler, 1996). The gonotrophic cycle includes the search for the host, the ingestion of a blood meal, the digestion of the blood, the maturation of ovaries. It is completed with the laying of eggs once females have found an appropriate oviposition site (Beklemishev, 1940). Figure 2.2 graphically describes the integration of the physiological processes (summarised as host-seeking, ingestion and digestion of blood, egg maturation and oviposition) associated with feeding and reproduction of *Ae. aegypti* as suggested by Klowden (1994).

Figure 2.2. *Ae. aegypti* gonotrophic cycle, taking into account the factors that can cause host-seeking behaviour to return after an initial blood meal



Source: Klowden, M.J. (1994), "Endogenous regulation of the attraction of *Aedes aegypti* mosquitoes", *Journal of the American Mosquito Control Association*, Vol. 10, pp. 326-332.

The host-seeking behaviour of *Ae. aegypti* is closely associated with anthropogenic environments, in and around homes and other places that people frequent. During host-seeking behaviour in mosquitoes, visual, thermal and olfactory stimuli all contribute to host location, but olfaction is probably the dominant sensory mode used for this purpose (Bowen, 1996).

The visual capacity of *Aedes* mosquitoes to distinguish between various optical stimuli such as luminous reflectance, vertical contrast, and movement (Muir, Kay and Thorne, 1992; Hoel, Kline and Allan, 2009), as well as their preference of resting on black, stationary objects and non-reflective surfaces such as clothing, are characteristics that have served in the development of various entomological sampling devices and traps, e.g. the BG Sentinel trap, ovitraps, gravid Aedes Traps and BDV tent trap (Fay and Prince, 1970; Muir, Kay and Thorne, 1992; Edman et al., 1997; Kroeckel et al., 2006; Silver, 2007; Casas Martínez et al., 2013; Eiras, Buhagiar and Ritchie, 2014).

With regard to the role of olfaction in host-seeking behaviour, carbon dioxide (CO_2) is involved in both short-range and long-range attraction. Olfactory cues that are primarily involved in long-range attraction include skin emanations, exhaled air and urine.

Each of these is attractive to all mosquito species. Attraction is caused by a mixture of several host emanated compounds (Takken, 1991). Lactic acid in the presence of CO_2 is attractive, and lactic acid-sensitive neurosensilla are present on the antennae of *Ae. aegypti*. Other host-produced chemicals are also attractive. The plant-derived odorant linalool oxide, in combination with CO_2 is also an effective long-range attractant (Nyasembe et al., 2015).

The amount of blood ingested by a female *Ae. aegypti* mosquito (> 2.5 µl on average) can affect its host-seeking response. The suspension of host-seeking behaviour is caused by abdominal distension due to the ingested blood, or due to hormonal inhibition (Klowden and Lea, 1978).

To meet the adult female's energy and reproductive needs this species has adopted a strategy that includes reduced consumption of plant carbohydrates, highly focused blood feeding on humans, and frequently engaging in multiple blood feedings (Scott and Takken, 2012). *Ae. aegypti* almost exclusively fed on humans (99%) as a single host species, and 97% of multiple-host blood meals included at least one human host. A low frequency of other hosts, including bovine, swine, cat, rat and chicken were detected, but they represented less than 1% of blood meals (Ponlawat and Harrington, 2005). Both males and females can feed on plant juices (nectar), damaged fruits, damaged and intact vegetative tissue, and homopterans (aphids) which act as an energy source for their physiological maintenance and locomotion (Clements, 2000). Carbohydrate consumption rates (fructose) ranged from 1% to 27% for females and 9% to 65% for males (males are not hematophagous) (Van Handel et al., 1994; Martínez-Ibarra et al., 1997). Sugar feeding in *Ae. aegypti* is believed to be facultative because studies indicate that in the absence of human hosts, females showed higher fructose feeding rates, up to 74% (Van Handel et al., 1994).

The usual gonotrophic cycle of *Ae. aegypti* is described above. However, lack of association between blood feeding and ovogenesis, a term known as gonotrophic discordance, is fairly common in *Ae. aegypti*. This concept is defined as the need for multiple blood meals during a single gonotrophic cycle. The occurrence of multiple partial meals for a gonotrophic cycle (Feinson and Spielman, 1980; Clements, 1992) and reduced feeding success may be due to host defensive behaviour, body size of females and local female *Ae. aegypti* mosquito abundance (Klowden and Lea, 1978; Clements, 1992). The habit of feeding on blood twice during one gonotrophic cycle depends greatly on the size and hence stored energy reserves of the teneral female (Takken et al., 1998).

Some field studies with *Ae. aegypti* females demonstrated that 88% of all detectable meals were identified as being from a single host (human) and only 7% of all the females had taken multiple meals (Scott et al., 1993). Engorged females in Thailand revealed that half to one-third imbibed two or more blood meals in a 36-hour time period. On average, the human biting rate was high, with 0.63-0.76 blood meals per day (Scott et al., 2000). Multiple blood meals were also recorded using histological examination.

Protein obtained from the blood meal supplies the amino acids needed for vitellogenin synthesis, which is a protein critical for egg production in the female *Ae. aegypti* mosquito. In general, the post-ingestion digestion of blood takes about 38-48 hours in the midgut (MG) of *Ae. aegypti* (O'Gower, 1955; Gaio et al., 2011) and is dependent upon temperature and, to a lesser extent, humidity (Shlenova, 1938; West and Eligh, 1952).

Many bacteria live and multiply in the MG of *Ae. aegypti*, contributing to digestion, nutrition, and development of their host. The reduction in these symbiotic MG bacteria (primarily *Enterobacter* sp. and *Serratia* sp.) can affect the lysis of red blood cells, subsequently retarding protein digestion, depriving the mosquito of essential nutrients and eventually affecting oocyte maturation resulting in the production of fewer viable eggs (Gaio et al., 2011).

The gonotrophic cycle duration is operationally defined as the average number of days that gravid mosquitoes took to oviposit after taking a blood meal. From a human health perspective, the gonotrophic cycle is one of the most important physiological processes in the life of mosquitoes vectoring dengue and represents an essential epidemiological component in the model of vectorial capacity. It is a significant and determining biological aspect in the population dynamics of *Ae. aegypti* and *Ae. albopictus*, both of which can coexist in urban, suburban, and rural regions with endemic dengue and other arboviral diseases. Bacon (1916) in West Africa found that the first meal was taken one to two days after emergence and subsequent meals taken after each oviposition at about three-day intervals. The development of follicles from stage I to V (Christophers, 1911), takes 1.67 days during the first gonotrophic cycle of *Ae. aegypti* females when fed with a blood supply to repletion, and maintained at an average temperature of 28.9°C. The maturation of eggs can extend up to 2.7 days with an average temperature of 26.2°C (Tamayo-Domínguez, 2011). Additionally, female *Ae. aegypti* took 2.8 days to complete the first gonotrophic cycle when the average temperature was 26.2°C (Tamayo-Domínguez, 2011).

Because the processes of feeding and reproduction are closely related in most anautogenous (requiring a blood meal) anthropophilic mosquitoes like *Ae. aegypti*, therefore larval nutritional regimen, body size of newly-emerged adults, and the quantity and quality of blood ingested by females are key considerations (Macdonald, 1956). In mosquitoes, egg production is a cyclic process; therefore, with each successive reproductive or gonotrophic cycle a batch of oocytes matures and a new set of follicles forms within the germaria, separates and starts development. In *Ae. aegypti*, secondary follicles appear when the primary follicles enter the previtellogenic resting stage (Clements, 2000).

Once ovogenesis, which is asynchronous (Clements, 1992), is complete (or reaches Christophers' stage V), the priority of a female *Ae. aegypti* is to search for an oviposition site. Typically, eggs are deposited in naturally occurring collections of fresh water (such as coconut shells, leaves and axils of plants, tree holes, hollows of rocks) and various artificial containers made of plastic, glass, ceramic or metal, while holding temporal (e.g. tires, vases, bottles, kitchenware, scrap metal) and/or permanent water sources (pools, drums, tanks, etc.) that provide both habitat and food for immature life stages (Thavara et al., 2001; Vezzani and Schweigmann, 2002; García-Rejón et al., 2011). Oviposition sites may be located inside and outside human habitations, as well as in non-residential places such as cemeteries, workshops, junkyards, tire repair facilities and vacant plots. There have been reports of *Ae. aegypti* larvae being found in the surface clear water layer of septic tanks (Burke et al., 2010), but this is not frequent and usually occurs where the lid is cracked or broken, providing the female access; nonetheless, septic tanks can be prolific producers (Barrera et al., 2008). Breeding sites also can include those that might contain brackish water such as boats or man-made containers at coastal edges or on beaches (Ramasamy et al., 2011). Waste material containers that are situated in areas with overhanging vegetation provide more favourable habitats as the breeding site is both shaded from intense sunshine and the build-up of heat and

provides a ready source of detritus and bacteria for larval consumption. These containers are usually breeding sites for mosquitoes only during the rainy season in countries with wet and dry seasons, but the eggs are resistant to desiccation and can remain in suitable containers until rains of the following season. These desiccated eggs form what is known as the egg bank.

The choice of an egg-laying site by *Ae. aegypti* is influenced by the presence of conspecific larvae and pupae, the container fill method, container size, lid and sun exposure (Wong et al., 2011). Surprisingly, egg-laying females were most attracted to sites containing other immature *Ae. aegypti*, rather than to sites containing the most food. Physical attributes of oviposition sites, such as size, light-dark contrasts and specular reflectance from water surfaces, also play a significant role in oviposition site selection (Harrington et al., 2008). Characteristics of oviposition sites can vary according to the geographic and sociocultural context such as region, country and location. The degree of landscape modification (urban-rural) is also a factor (Kittayapong and Strickman, 1993; Honório et al., 2009), as well as intra- and interspecific competition (Chadee, Corbet and Greenwood, 1990; Braks et al., 2004; Sánchez-Hernández, 2011).

Behaviour of reproduction

Ae. aegypti is recognised as a highly anthropophilic, endophilic, endophagic, and day-biting species (Scott and Takken, 2012; Brown et al., 2014; McBride et al., 2014). These designations are based on activity patterns exhibited by this mosquito around the world. An important aspect of the bionomics of *Ae. aegypti* that contributes to its efficiency as an epidemiological vector is the close association with domestic habitats (Scott et al., 2000). Adult mosquitoes frequently reside indoors in human dwellings, most commonly in bedrooms (60.3% to 63.5%) followed by living/dining rooms (9.3% to 18.4%), kitchens (7.5% to 9.7%) and bathrooms (6.6% to 11.5%) (García-Rejón et al., 2008; Casas-Martínez, 2013). Immature forms develop primarily in artificial containers such as cans, jars, tires and buckets (Winch et al., 1992; García-Rejón et al., 2011). In Chennai, one of the major metropolitan areas in India, intradomiciliary cement tubs containing water for multi-purpose works were mostly preferred by the *Ae. aegypti* immature forms for development (Arunachalam et al., 2010). More details are given under Chapter 4.

Mosquitoes are exposed to daily changes in environmental light-dark cycles along with variations in humidity and temperature. Adaptation to these changes is seen in the form of specific behaviours, which are in turn linked to the expression of specific endogenously-controlled genes. *Ae. aegypti* is a major vector of arbovirus in many countries,[2] therefore the ethological study of this mosquito is crucial to better understand their behaviour, the dynamics of transmission of the viruses, as well as to optimise the entomological surveillance and increase the efficiency of vector control (Lima-Camara, 2010; Sivagnaname and Gunasekaran, 2012).

There are two significant copulation peaks in indoor housing, an early-morning peak between 6h00 and 8h00 (25% of events) and a pre-sunset peak from 16h00 to 18h00 (24% of events). The outdoors copulation periodicity presents almost the same pattern in the timing, with 30% of events during the early morning peak and 25% of events during the pre-sunset peak. Observations in insectary have shown similar copulation patterns. Studies indicated that 38.6% of copulating females collected in and around breeding sites were nulliparous and not inseminated, whereas over 85% of the copulating females found indoors were parous, suggesting that successful insemination encounters occur at

alternative sites (such as around the human host). Furthermore, males may not be able to detect the difference between virgin and mature, parous female mosquitoes (Chadee and Gilles, 2013).

Oviposition is also diurnal and bimodal, both indoors and outdoors, with consistent peaks at 6h00-8h00 and 16h00-18h00 (Chadee and Corbet, 1989, 1990; Corbet and Chadee, 1989). The oviposition activity intensifies during the rainy season due to increased availability of water filled containers and mosquito population abundance.

Visual observations of the mating behaviour of *Ae. aegypti* have shown that males swarm around the feet and lower legs of a sitting/standing human host, flying in a horizontal figure of eight pattern. Mating was usually initiated in flight at a height of not more than one metre from the ground. Copulating pairs have been observed in flight, on human bodies, on their trousers and on the ground (Hartberg, 1971). Mating also occurs near adult oviposition sites and resting sites. Tests carried out by Jong and Knols (1996), demonstrated that *Ae. aegypti* prefers to bite the head and upper part of the trunk of persons lying in prone or supine position but will often bite on the lower legs beneath tables and when the host is seated.

The host-seeking behaviour and biting activity of *Ae. aegypti* are closely related, therefore, both events describe overlapping biorhythms. Many authors have documented that males and females show a bimodal flying and landing activity and that the periodicity is the same for nulliparous, parous, inseminated or uninseminated females, all activity being predominantly diurnal, with sharp peaks at post-sunrise and pre-sunset (as reported above) in intra-, peri- and extradomiciliary sites (Trpis et al., 1973; Corbet and Smith, 1974; Casas-Martínez et al., 2013). Landing activity patterns of *Ae. aegypti* are influenced by environmental factors (for example electrical lighting in and around houses), both indoor and outdoor, and in urban and rural areas (Chadee and Martinez, 2000).

Fecundity and fertility

Some mosquito strains or species are able to lay eggs without taking a blood meal, a trait named autogeny. This may allow populations to persist through times or places where vertebrate hosts are scarce. Environmental and genetic factors determine whether the mosquito *Ae. aegypti* lays eggs without a blood meal (Ariani et al., 2015). Autogeny is increased by growth at a temperature of 28°C (compared with 22°C), good nutrition of larval stages and feeding on higher concentrations of sugar solution during the adult stage. There appears to be a genetic component to autogeny which allows adult females from some strains to utilise amino acids from fat stores from the larva stage instead of obtaining these nutrients from a blood meal. Genetic differences associated with autogeny also affect fecundity in autogenous *Ae. aegypti* strains as shown by blood feeding behaviour (Christophers, 1960), quantity and quality of blood acquired (Klowden and Lea, 1978; Clements, 1992) and insemination status (Lavoipierre, 1958).

Mosquito body size has been linked to longevity, the number of eggs per batch and vector competence, and it is therefore an important measure of mosquito fitness (Siegel et al., 1992). The average body size corresponding to a wing length of 2.6 mm was associated with 61.23 ± 29.15 eggs per batch (Tamayo-Domínguez, 2011).

Life table analysis, under natural (and laboratory) conditions

A life table of aquatic phase *Ae. aegypti* grown under favourable laboratory conditions (temperature maintained at 28 ± 1°C, humidity 70 ± 10%, app. 50 larvae in a 6-inch x 8-inch tray, yeast + dog biscuit powder, or an alternative, as food on alternate days) suggests that natural mortality during development of larval stages is initially low (1%, stage I) and then increases (9%, stage II; 34%, stage III; 34%, stage IV). Mortality then decreases during the pupal stage (6%). When grown in the laboratory, approximately 48% of eggs survive to adults (Sánchez-Hernández, 2011).

Observed patterns of coexistence/exclusion of *Ae. albopictus* and *Ae. aegypti* in the field (Murrell and Steven, 2008) may be due to variation in detritus type. Experimental trials confirm competitive asymmetry in favour of *Ae. albopictus* with oak, pine, rubber (Tyagi et al., 2006) or insect detritus. Certain detritus types may eliminate interspecific competition among the larvae of these species (Murrel and Steven, 2008), thereby allowing for stable coexistence. Desiccation and thermal tolerance of eggs are also factors affecting co-existence (Juliano et al., 2002). More details on the biotic interactions in the landscape are given in the related section of Chapter 4.

In general, the effects of microclimatic factors (temperature, humidity and rainfall) and other environmental variables (food source, breeding sites and shelters) in the life cycle of the mosquito *Ae. aegypti* and generation time have been well documented, in either natural or controlled conditions in an insectarium or laboratory. Environmental changes affect all life stages of the mosquito, influence their survival and thus their ability to transmit pathogens. Low humidity, for instance, can negatively affect adult survival and may decrease the vector population. Frequency and host type of blood meal influence fecundity and female survival (Christophers, 1960; Nelson, 1986; Rueda et al., 1990; Day, Edman and Scott, 1994; Carrington et al., 2013).

Interspecific breeding

Harper and Paulson (1994) examined the dynamics of interspecific and intraspecific mating between Florida strains of *Ae. aegypti* and *Ae. albopictus*. In non-choice experiments where conspecific males were not available, dissection of the spermatheacae showed that interspecific insemination was an infrequent event. Few eggs were produced from interspecific crosses and all were non-viable. The frequency of interspecific mating was not increased when the hind tarsi of females were removed, eliminating a significant mechanism for fending off unwanted courtship. When held with males of both species, females mated with conspecifics and oviposited without regard to the presence of other species. In low-density experiments in which a single female of either species is caged with an excess of males of the other species, the conspecific male always located and inseminated the female. However, the presence of females of the other species has some negative influence on the intraspecific mating success in male *Ae. aegypti*, most likely due to misdirected courting or mating efforts (Bargielowski, Blosser and Lounibos, 2015).

Additional studies further suggest that matings of *Ae. aegypti* with *Ae. albopictus* do not produce viable offspring in the laboratory (Harper and Paulson, 1994; Nazni et al., 2009). Forced matings in the laboratory between wild-type *Ae. aegypti* and *Ae. albopictus* yielded eggs but they were not viable, and when bleached were shown to have no embryos (Nazni et al., 2009). More recently a study showed that there is cross-species insemination in the field between *Ae. aegypti* and *Ae. albopictus* (Tripet et al., 2011), but these interspecific matings encounter many barriers and occur at low frequencies

(a single *Ae. albopictus* was found to have *Ae. aegypti* sperm in this study, and three *Ae. aegypti* females were inseminated by *Ae. albopictus*), resulting in no viable progeny.

These results indicate that significant reproductive isolation exists between *Ae. aegypti* and *Ae. albopictus*. This occurs at both the prezygotic level (very low mating frequency) and the postzygotic level (non-viable progeny).

In rare cases, viable hybrids resulting from cross-mating between *Ae. aegypti* females and *Ae. albopictus* males have occurred in laboratories (Martínez-López et al., 2014). Eggs obtained from this cross-mating were viable, and the larvae and pupae showed development in seven days. Therefore, it is possible that viable hybrids can be produced experimentally, but this is rare and may be restricted to matings of only particular strains of each species. As reported in the previous section, there are important reproductive barriers existing between these two species living in sympatry in natural environments (Harper and Paulson, 1994; Nazni et al., 2009) and the possibility of hybridisation is unlikely, given the probable sterility of F1 hybrids (Haldane's rule).

Effect of *Wolbachia* on reproduction

Wolbachia pipientis is a monophyletic group of maternally inherited, gram-negative, endosymbiotic bacteria, related to the *Ehrlichia*, *Anaplasma* and *Neorickettsia* genera, all being members of Alphaproteobacteria (O'Neill et al., 1992; Lo et al., 2007). In recent years, evidence has been accumulated that shows *Wolbachia* infections affect several aspects of host biology, physiology, immunity, ecology, evolution and reproduction (Bourtzis, Braig and Karr, 2003; Bourtzis and Robinson, 2006; Werren, Baldo and Clark, 2008; Saridaki and Bourtzis, 2010). This bacterial group is widespread and abundant among insect species and has been associated with the induction of a number of reproductive outcomes including the death of males (Hurst et al., 2000), feminisation (Rousset et al., 1992), parthenogenesis (Stouthamer, Breeuwer and Hurst, 1999) and, most commonly, cytoplasmic incompatibility (Yen and Barr, 1973; O'Neill et al., 1997; Nirgianaki et al., 2003).

The cytoplasmic incompatibility (CI) results in the generation of unviable offspring when an uninfected female mates with a *Wolbachia*-infected male (McGraw et al., 2001). In contrast, *Wolbachia*-infected females can produce viable progeny when they mate with both infected and uninfected males, resulting in a selective reproductive advantage over uninfected females (Hoffmann and Turelli, 1997). This CI phenotype is induced by *Wolbachia* in mosquito species and allows the maternally-transmitted *Wolbachia* to efficiently invade host populations without being infectious or moving horizontally between individuals (Hoffmann and Turelli, 1997).

The ability of *Wolbachia* to manipulate diverse functional systems of its hosts (Bourtzis et al., 2014), particularly reproduction, has led to the proposal and the development of promising symbiont-based strategies aimed at the control of insect pests and disease vectors including mosquito species. Different *Wolbachia* species/strains can be naturally found in *Aedes* mosquitoes, for example, *Ae. albopictus*, *Ae. polynesiensis* and *Ae. scutellaris*, but not in *Ae. aegypti*. Thus, the use of *Wolbachia* for *Ae. aegypti* control via CI has required its transinfection from naturally infected insect species (Ye et al., 2013; Joubert et al., 2016) and currently includes the wAlbB strain (from *Ae. albopictus*) and the wMelPop-CLA (cell-line-adapted) and wMel strains (from *Drosophila melanogaster*). More information on the use of *Wolbachia* as a biological control for virus transmission is given under Annex A.

Notes

[1] This GE line was carrying a tetracycline repressible, lethal positive feedback system.

[2] See Annex B. Human and animal health affected by mosquitoes, Table A B.1 on arbovirus definition and important infections.

References

Ariani, C.V. et al. (2015), "Environmental and genetic factors determine whether the mosquito *Aedes aegypti* lays eggs without a blood meal", *The American Journal of Tropical Medicine and Hygiene*, Vol. 92, No. 4, pp. 715-721.

Arrivillaga, J. and R. Barrera (2004), "Food as a limiting factor for *Aedes aegypt*i in water-storage containers", *Journal of Vector Ecology*, Vol. 29, No. 1, pp. 11-20.

Arunachalam, N. et al. (2010), "Eco-bio-social determinants of dengue vector breeding: A multi-country study in urban and periurban Asia", *Bulletin of the World Health Organization*, Vol. 88, No. 3, pp. 173–184.

Avila, F.W. et al. (2011), "Insect seminal fluid proteins: Identification and function", *Annual Review of Entomology*. *2011*, Vol. 56, pp. 21–40.

Bacon AW. (1916), Investigation Report of Yellow Fever Commission West Africa", in *Report of the entomological investigation undertaken for the Yellow Fever (West Africa) Commission for the year August 1914, to July 1915* (Quoted by Edwards, 1941).

Bargielowski, I., E. Blosser and L.P. Lounibos (2015), "The effects of interspecific courtship on the mating success of *Aedes aegypti* and *Aedes albopictus* (Diptera: Culicidae) males", *Annals of the Entomological Society of America*, Vol. 108, No. 4, pp. 513–518.

Bargielowski, I. et al. (2011), "Comparison of life history characteristics of the genetically modified OX513A line and a wild type strain of *Aedes aegypti*", *PLoS One*, Vol. 6, No. 6: e20699.

Barrera, R. et al. (2008), "Unusual productivity of *Aedes aegypti* in septic tanks and its implications for dengue control", *Medical and Veterinary Entomology*, Vol. 22, No. 1, pp. 62-69.

Beklemishev, W.N. (1940), "Gonotrophic rhythm as a basic principle of the biology of *Anopheles*", *Vopr Fiziol Ekol Malar Komara*, Vol. 1, pp. 3-22.

Bourtzis, K., H.R. Braig and T.L. Karr (2003), "Cytoplasmic incompatibility", in K. Bourtzis and T.A. Miller (eds.), *Insect Symbiosis, Vol. 1*, CRC Press, pp. 217-246.

Bourtzis, K. and A.S. Robinson (2006), "Insect pest control using *Wolbachia* and/or radiation", in K. Bourtzis and T.A. Miller (eds.), *Insect Symbiosis, Vol. 2*, CRC Press, pp. 225-246.

Bourtzis, K. et al. (2014), "Harnessing mosquito-*Wolbachia* symbiosis for vector and disease control", *Acta Tropica*, Vol. 132 (Suppl.), S150-S163.

Bowen, M.F. (1996), "Sensory aspects of host location in mosquitoes", in *CIBA Foundation Symposium 200*, pp. 197-201 and 226-232.

Braks, M.A. et al. (2004), "Interspecific competition between two invasive species of container mosquitoes, *Aedes aegypti* and *Aedes albopictus (Diptera: Culicidae)*, in Brazil", *Annals of Entomological Society of America*, Vol. 97, No. 1, pp. 130-139.

Brogdon, W.G. (1994), "Measurement of flight tone differences between female *Aedes aegypti* and *A. albopictus* (Diptera: Culicidae)", *Journal of Medical Entomology*, Vol. 31(5), pp.700-703.

Brown, J.E. et al. (2014), "Human impacts have shaped historical and recent evolution in *Aedes aegypti*, the dengue and yellow fever mosquito", *Evolution*, Vol. 68, No. 2, pp. 514-525.

Burke, R.L. et al. (2010), "Examination of a miniaturized funnel trap for *Aedes aegypti* (Diptera: Culicidae) larval sampling", *Journal of Medical Entomology*, Vol. 47, No. 6, pp. 1231–1234.

Carrington, L.B. et al. (2013), "Effects of fluctuating daily temperatures at critical thermal extremes on *Aedes aegypti* life-history traits", *PLoS ONE*, Vol. 8, No. 3: e58824.

Casas-Martínez, M. (2013), "Bionomía comparativa de *Aedes aegypti* y *Aedes albopictus* y sus implicaciones en la transmisión del dengue en el sur de México" [Comparative bionomy of *Aedes aegypti* and *Aedes albopictus* and its implications for the transmission of dengue fever in southern Mexico], Doctoral thesis, Tapachula.

Casas-Martínez, M. et al. (2013), "A new tent trap for monitoring the daily activity of *Aedes aegypti* and *Aedes albopictus*", *Journal of Vector Ecology*, Vol. 38, No. 2, pp.277-288.

Cator, L.J. and L.C. Harrington (2011), "Harmonic convergence and sexy sons: Indirect benefits associated with acoustic signals in the dengue vector", *Animal Behaviour*, Vol. 82, No. 4, pp. 627-633.

Cator, L.J. et al. (2009), "Harmonic convergence in the love songs of the dengue vector mosquito", *Science*, Vol. 323, No. 5917, pp. 1077-1079.

Chadee, D.D. and J.R.L. Gilles (2013), "The diel copulation periodicity of the mosquito, *Aedes aegypti* (L.) (Diptera: Culicidae) at indoor and outdoor sites in Trinidad, West Indies", *Acta Tropica*, Vol. 132S, pp. 91-95.

Chadee, D.D. and P.S. Corbet (1990), "A night-time role of the oviposition site of the mosquito, *Aedes aegypti* (L.) (Diptera: Culicidae)", *Annals of Tropical Medicine and Parasitology*, Vol. 84, No. 5, pp. 429-433.

Chadee, D.D. and P.S. Corbet (1989), "Diel patterns of oviposition of the mosquito, *Aedes aegypti* (L.) (Diptera: Culicidae) in Trinidad, W. I.: A preliminary study", *Annals of Tropical Medicine and Parasitology*, Vol. 84, No. 1, pp. 79-84.

Chadee, D.D., P.S. Corbet and J.J.D. Greenwood (1990), "Egg-laying Yellow Fever Mosquitoes avoid sites containing eggs laid by themselves or by conspecifics", *Entomology Experimental Applied*, Vol. 57, pp. 295-298.

Chadee, D.D. and R. Martinez (2000), "Landing periodicity of *Aedes aegypti* with implications for dengue transmission in Trinidad, West Indies", *Journal of Vector Ecology*, Vol. 25, No. 2, pp. 158-163.

Christophers, S.R. (1960), Aedes aegypti *(L.) The Yellow Fever Mosquito. Its Life History, Bionomics and Structure*, Cambridge University Press, Cambridge.

Christophers, S.R. (1911), "The development of the egg follicle in Anophelines", *Paludism*, Vol. 2, pp. 73-78.

Clements, A.N. (2000), *The Biology of Mosquitoes, Volume I: "Development, Nutrition and Reproduction" Second Edition*, CABI Publishing, Oxford.

Clements, A.N. (1992), *The Biology of Mosquitoes, Volume I "Development, Nutrition and Reproduction"*, Chapman and Hall, London.

Colvard, J. (1978), "El comportamiento alimentario de los mosquitos", *Investigación y Ciencia, Spanish edition of Scientific American*, Vol. 23, pp. 86-93.

Corbet, P.S. and D.D. Chadee (1989), "Incidence and diel pattern of oviposition outdoors of the mosquito, *Aedes aegypti* (L.) (Diptera: Culicidae) in Trinidad, W. I. in relation to solar aspect", *Annals of Tropical Medicine and Parasitology*, Vol. 84, No. 1, pp. 63-78.

Corbet, P.S. and S.M. Smith (1974), "Diel periodicities of landing of nulliparous and parous *Aedes aegypti* (L.) at Dar es Salaam, Tanzania (Diptera: Culicidae)", *Bulletin of Entomological Research*, Vol. 64, pp. 111-121.

Couret, J. and M.Q. Benedict (2014). "A meta-analysis of the factors influencing development rate variation in *Aedes aegypti* (Diptera: Culicidae)", *BioMed Central Ecology*, Vol. 14, No. 3, pp 1-15.

Day, J.F., J.D. Edman and T.W. Scott (1994), "Reproductive fitness and survivorship of *Aedes aegypti* (Diptera: Culicidae) maintained on blood, with field observations from Thailand", *Journal of Medical Entomology*, Vol. 31, No. 4, pp. 611-617.

Edman, J.D. et al. (1998), "*Aedes aegypti* (Diptera: Culicidae) movement influenced by availability of oviposition sites", *Journal of Medical Entomology*, Vol. 35, pp. 578-583.

Eiras, A.E., T.S. Buhagiar and S.A. Ritchie (2014), "Development of the Gravid *Aedes* Trap for the capture of adult female container-exploiting mosquitoes (Diptera: Culicidae)", *Journal of Medical Entomology*, Vol. 51, pp. 200-209.

Farnesi, L.C. et al. (2009), "Embryonic development of *Aedes aegypti* (Diptera: Culicidae): Influence of different constant temperatures", *Memórias do Instituto Oswaldo Cruz*, Vol. 104, No. 1, pp. 124-126.

Fay, R.W. and W.H. Prince (1970), "A modified visual trap for *Aedes aegypti*", *Mosquito News*, Vol. 28, pp. 1-7.

Feinson, F.M. and A. Spielman (1980), "Nutrient mediated juvenile hormone secretion in mosquitoes", *Journal of Insect Physiology*, Vol. 26, pp. 113-117.

Focks, D.A. et al. (1981), "Observations on container-breeding mosquitoes in New Orleans, Louisiana with an estimate of the population density of *Aedes aegypti* (L)", A*merican Journal of Tropical Medicine and Hygiene*, Vol. 30, pp. 1329-1335.

Foster, W.A. and E.D. Walker (2002), "Mosquitoes (Culicidae)", in G. Mullen and L. Durden (eds.), *Medical and Veterinary Entomology*, Academic Press, San Diego, pp. 203-262.

Gaio, A. de O. et al. (2011), "Contribution of midgut bacteria to blood digestion and egg production in *Aedes aegypti* (Diptera: Culicidae) (L.)", *Parasites and Vectors*, Vol. 4, pp. 105.

García-Rejón, J.E. et al. (2011), "Productive container types for *Aedes aegypti* immatures in Mérida, México", *Journal of Medical Entomology*, Vol. 48, No. 3, pp. 644-650.

Gillett, J.D., E.A. Roman and V. Phillips (1977), "Erratic hatching in *Aedes* eggs: A new interpretation", *Proceedings of the Royal Society of London. Series B, Biological Sciences*, Vol. 196, pp. 223-232.

Goindin, D. et al. (2015), "Parity and longevity of *Aedes aegypti* according to temperatures in controlled conditions and consequences on dengue transmission risks", *PLoS ONE*, Vol. 10, No. 8: e0135489.

Gordh, G. (2001), *A Dictionary of Entomology*, compiled by G. Gordh with assistance by D. Headrick, CABI Publishing, Wallingford (United Kingdom) and Cambridge (Massachusetts).

Hancock, P.A. et al. (2016), "Density dependent population dynamics in *Aedes aegypti* slow the spread of wMel *Wolbachia*", *Journal of Applied Ecology*, Vol. 53, No. 3, pp. 785-793.

Harper, J.P. and S. Paulson (1994), "Reproductive isolation between Florida strains of *Aedes aegypti* and *Aedes albopictus*", *Journal of the American Mosquito Control Association*, Vol. 10, pp. 88-92.

Harrington, L.C. et al. (2008), "Influence of container size, location, and time of day on oviposition patterns of the dengue vector, *Aedes aegypti*, in Thailand", *Vector-Borne and Zoonotic Diseases*, Vol. 8, No. 3, pp. 415–423.

Hartberg, W.K. (1971), *Observation on the Mating Behavior of* Aedes aegypti *in Nature*, World Health Organization, pp. 847-850.

Helinski, M.E. et al. (2012), "Evidence of polyandry for *Aedes aegypti* in semifield enclosures", *American Journal of Tropical Medicine and Hygiene*, Vol. 86, No. 4, pp. 635-641.

Hoel, D., D. Kline and S. Allan (2009), "Evaluation of six mosquito traps for collection of *Aedes albopictus* (Skuse) and associated mosquito species in a suburban setting in north central Florida", *Journal of the American Mosquito Control Association*, Vol. 25, pp. 47-57.

Hoffman, A.A. and M. Turelli (1997), "Cytoplasmic incompatibility in insects", in S.L. O'Neill, A.A. Hoffman and J.H. Werren (eds), *Influential Passengers*, Oxford University Press, New York, pp. 42–80.

Honório, N.A. et al. (2009), "The spatial distribution of *Aedes aegypti* and *Aedes albopictus* in a transition zone, Rio de Janeiro, Brazil", *Cad Saude Publica*, Vol. 25, No. 6, pp. 1203-1214.

Hurst, G.D. et al. (2000), "Male-killing *Wolbachia* in Drosophila: A temperature-sensitive trait with a threshold bacterial density", *Genetics*, Vol. 156, No. 2, pp. 699-709.

Jeffery, J.A.L. et al. (2012), "Water level flux in household containers in Vietnam - A key determinant of *Aedes aegypti* population dynamics", *PLoS ONE*, Vol. 7, No. 7: e39067.

Jong, R. and B.G. Knols (1996), "Selection of biting sites by mosquitoes", in G.R. Bock and G. Cardew, *Olfaction in Mosquito-Host Interactions*, Ciba Foundation, England, pp. 1-331.

Joubert, D.A. et al. (2016), "Establishment of a Wolbachia superinfection in *Aedes aegypti* mosquitoes as a potential approach for future resistance management", *PLoS Pathos*, Vol. 12, No. 2: e1005434.

Judson, C.L. (1960), "The physiology of hatching of aedine mosquito eggs: Hatching stimulus", *Annals of the Entomological Society of America*, Vol. 53, pp. 688-691.

Juliano, S.A. et al. (2002), "Desiccation and thermal tolerance of eggs and the coexistence of competing mosquitoes", *Oecologia*, Vol. 130, pp. 458-469.

Kampen, H. and F. Schaffner (2008), "11. Mosquitoes", in X. Bonnefoy, H. Kampen and K. Sweeney (eds.), *Public Health Significance of Urban Pests*, World Health Organization, WHO Regional Office for Europe, Denmark, pp. 347-386.

Kittayapong, P. and D. Strickman (1993), "Distribution of container-inhabiting *Aedes* larvae (Diptera: Culicidae) at a dengue focus in Thailand", *Journal of Medical Entomology*, Vol. 30, No. 3, pp. 601-606.

Klowden, M.J. (1994), "Endogenous regulation of the attraction of *Aedes aegypti* mosquitoes", *Journal of the American Mosquito Control Association*, Vol. 10, pp. 326-332.

Klowden, M.J. and A.O. Lea (1978), "Blood meal size as a factor affecting continued host-seeking by *Aedes aegypti* (L.)", *American Journal of Tropical Medicine and Hygiene*, Vol. 27, pp. 827-831.

Koenraadt, C.J.M. and L.C. Harrington (2008), "Flushing effect of rain on container-inhabiting mosquitoes *Aedes aegypti* and *Culex pipiens* (Diptera: Culicidae)", *Journal of Medical Entomology*, Vol. 45, No. 1, pp. 28-35.

Kroeckel, U. et al. (2006), "New tools for surveillance of adult yellow fever mosquitoes: Comparison of trap catches with human landing rates in an urban environment", *Journal of the American Mosquito Control Association*, Vol. 22, No. 2, pp. 229-238.

Lavoipierre, M.M. (1958), "Biting behavior of mated and unmated females of an African strain of *Aedes aegypti*", *Nature*, Vol. 181, pp. 1781-1782.

Lehane, M.J. (1991), *Biology of Blood-Sucking Insects. First Edition*, Chapman and Hall, London, pp. 288.

Lima-Camara, T.N. (2010), "Activity patterns of *Aedes aegypti* and *Aedes albopictus* (Diptera: Culicidae) under natural and artificial conditions", *Oecologia Aust*ralis Vol. 14, pp. 737-744.

Lo, N. et al. (2007), "Taxonomic status of the intracellular bacterium *Wolbachia pipientis*", *International Journal of Systematics and Evolutionary Microbiology*, Vol. 57, pp. 654-657.

Luz, C. et al. (2008), "Impact of moisture on survival of *Aedes aegypti* eggs and ovicidal activity of Metarhizium anisopliae under laboratory conditions", *Memórias do Instituto Oswaldo Cruz*, Vol.103, No.2, pp. 214-215.

Macdonald, W.W. (1956), "*Aedes aegypti* in Malaya: II larval and adult biology", *Annals of Tropical Medicine and Parasitology*, Vol. 50, pp. 399-414.

Manrique-Saide, P. et al. (1998), "*Mesocyclops longisetus* effects on survivorship of *Aedes aegypti* immature stages in car tyres", *Medical and Veterinary Entomology.*, Vol. 12, No. 4, pp. 386-390.

Martínez-Ibarra, J.A. et al. (1997), "Influence of plant abundance on nectar feeding by *Aedes aegypti* (Diptera: Culicidae) in southern Mexico", *Journal of Medical Entomology*, Vol. 34, No. 6, pp. 589-593.

Martínez-López, Y. et al. (2014), "Cruzamiento interespecífico entre *Aedes aegypti* y *Aedes albopictus* en el laboratorio" [Interspecific breeding between *Aedes aegypti* and *Aedes albopictus* in laboratory], *Revista Cubana de Medicina Tropical* [*Cuban Journal of Tropical Medicine*], Vol. 66, No. 1, pp. 148-151.

McBride, C.S. et al. (2014), "Evolution of mosquito preference for humans linked to an odorant receptor", *Nature*, Vol. 515, pp. 222–227.

McGraw, E.A. et al. (2001), "*Wolbachia*-mediated sperm modification is dependent on the host genotype in Drosophila", *Proceedings of the Royal Society B: Biological Sciences*, Vol. 268, No. 1485, pp. 2565-2570.

Mogi, M. and J. Mokry (1980), "Distribution of *Wyeomyia smithii* (Diptera: Culicidae) eggs in pitcher plants in Newfoundland, Canada", *Tropical Medicine*, Vol. 22, pp. 1-12.

Muir, L.E., B.H. Kay and M.J. Thorne (1992), "*Aedes aegypti* (Diptera: Culicidae) vision: Response to stimuli from the optical environment", *Journal of Medical Entomology*, Vol. 29, pp. 445-450.

Murrell, E.G. and A.J. Steven (2008), "Detritus type alters the outcome of interspecific competition between *Aedes aegypti* and *Aedes albopictus* (Diptera: Culicidae)", *Journal of Medical Entomology*, Vol. 45, No. 3, pp. 375-385.

Nazni, W.A. et al. (2009), "Cross-mating between Malaysian strains of *Aedes aegypti* and *Aedes albopictus* in the laboratory", *The Southeast Asian Journal of Tropical Medicine and Public Health*, Vol. 40, No. 1, pp. 40-46.

NCDENR (n.d.), *Mosquitoes... Some Facts: Information Pamphlet*, NC Dep. of Environment and Natural Resources, Div. of Environmental Health, Public Health Pest Management Section, Burlington North Carolina, United States, www.alamance-nc.com/envhealth/wp-content/uploads/sites/9/2013/10/Mosquitoes_Facts.pdf.

Nelson, M.J. (1986), Aedes aegypti*: Biology and Ecology*, Pan American Health Organization, Washington, DC, PNSP/86-63, pp. 50.

Nirgianaki, A. et al. (2003), "Wolbachia infections of the whitefly *Bemisia tabaci*", *Current Microbiology*, Vol. 47, No. 2, pp. 93-101.

Nyasembe, V.O. et al. (2015), "Linalool oxide: Generalist plant-based lure for mosquito disease vectors", *Parasites and Vectors*, Vol. 8, No. 581, pp. 1-8.

O'Gower, A.K. (1955), "The rate of digestion of human blood by certain species of mosquitoes", *Australian Journal of Biological Sciences*, Vol. 9, No.1, pp. 125-129.

O'Neill, S.L. et al. (1997), "In vitro cultivation of *Wolbachia pipientis* in an *Aedes albopictus* cell line", *Insect Molecular Biology*, Vol. 6, pp. 33-39.

O'Neill, S.L. et al. (1992), "16S rRNA phylogenetic analysis of the bacterial endosymbionts associated with cytoplasmic incompatibility in insects", *PNAS*, Vol. 89, pp. 2699-2702.

Pilger, D. et al. (2011), "Is routine dengue vector surveillance in central Brazil able to accurately monitor the *Aedes aegypti* population? Results from a pupal productivity survey", *Tropical Medicine & International Health*, Vol. 16, No. 9, pp. 1143-1150.

Ponce, G.G. (1999), "Efecto de concentraciones subletales de *Bacillus thuringiensis israelensis* H-14 Vectobac® AS en parámetros biológicos de *Aedes aegypti*" [Effect of sublethal concentrations of *Bacillus thuringiensis israelensis* H-14 Vectobac® AS on biological parameters of *Aedes aegypti*], Masters thesis, Universidad Autonoma de Nuevo León, Facultad de Ciencias Biologicas.

Ponlawat, A. and L. Harrington (2009), "Factors associated with male mating success of the dengue vector mosquito, *Aedes aegypti*", *American Journal of Tropical Medicine and Hygiene*, Vol. 80, No. 3, pp. 395-400.

Ponlawat, A. and L. Harrington (2007), "Age and body size influence male sperm capacity of the dengue vector *Aedes aegypti* (Diptera: Culicidae)", *Journal of Medical Entomology*, Vol. 44, No. 3, pp. 422-426.

Ponlawat, A. and L.C. Harrington (2005), "Blood feeding patterns of *Aedes aegypti* and *Aedes albopictus* in Thailand", *Journal of Medical Entomology*, Vol. 42, No. 5, pp. 844-849.

Ponnusamy, L. et al. (2011), "Bacteria stimulate hatching of yellow fever mosquito eggs", *PLoS ONE*, Vol. 6, No. 9: e24409.

Ramasamy, R. et al. (2011), "Larval development of *Aedes aegypti* and *Aedes albopictus* in peri-urban brackish water and its implications for transmission of arboviral diseases", *PLoS Neglected Tropical Diseases*, Vol. 5, No. 11: e1369.

Reiter, P. et al. (1995), "Dispersal of *Aedes aegypti* in an urban area after blood feeding as demonstrated by rubidium-marked eggs", *The American Journal of Tropical Medicine and Hygiene*, Vol. 52, pp. 177-179.

Richardson, J.B. et al. (2015), "Evidence of limited polyandry in a natural population of *Aedes aegypti*", *The American Journal of Tropical Medicine and Hygiene*, Vol. 93, No. 1, pp. 189–193.

Rousset, F. et al. (1992), "*Wolbachia* endosymbionts responsible for various alterations of sexuality in arthropods", *Proceedings of the Royal Society B: Biological Sciences*, Vol. 250, No. 1328, pp. 91-98.

Rubio, A., M.V. Cardo and D. Vezzani (2011), "Tire-breeding mosquitoes of public health importance along an urbanization gradient in Buenos Aires, Argentina", *Memórias do Instituto Oswaldo Cruz*, Vol. 106, No. 6, pp. 678-684.

Rueda, L.M. et al. (1990), "Temperature-dependent development and survival rates of Culex quinquefasciatus and *Aedes aegypti* (Diptera. Culicidae)", *Journal of Medical Entomology*, Vol. 27, No. 5, pp. 892-898.

Sánchez-Hernández, C. (2011), "Competencia larvaria interespecífica de *Aedes aegypti* y *Aedes albopictus* en condiciones de insectario en Tapachula", *Tesis de Licenciatura*, Universidad Autónoma de Chiapas, Tapachula.

Saridaki, A. and K. Bourtzis (2010), "*Wolbachia*: More than just a bug in insects genitals", *Current Opinion in Microbiology*, Vol. 13, pp. 67-72.

Scott, T.W. and W. Takken (2012), "Feeding strategies of anthropophilic mosquitoes result in increased risk of pathogen transmission", *Trends in Parasitology*, Vol. 28, No. 3, pp. 114-121.

Scott, T.W. et al. (2000), "Longitudinal studies of *Aedes aegypti* (Diptera: Culicidae) in Thailand and Puerto Rico: Population dynamics", *Journal of Medical Entomology*, Vol. 37, No. 1, pp. 77-88.

Scott, T.W. et al. (1993), "Blood-feeding patterns of *Aedes aegypti* (Diptera: Culicidae) collected in a rural Thai village", *Journal of Medical Entomology*, Vol. 30, No. 5, pp. 922-927.

Service, M. (2012), *Medical Entomology for Students, 5th Ed*., Cambridge University Press, New York, pp. 303.

Shlenova, M.F. (1938), "[The speed of blood digestion in female *A. maculipennis messeae* at stable effective temperatures - Vitesse de la digestion du sang par la femelle de l'*Anopheles maculipennis messeae* aux temperatures effectives constantes] (in Russian)", *Med. Parazit. (Moscow)*, Vol. 7, pp. 716-735.

Siegel, J.P. et al. (1992), "Statistical appraisal of the weight-wing length relationship of mosquitoes", *Journal of Medical Entomology*, Vol. 29, No. 4, pp. 711-714.

Silver, J.B. (2007), *Mosquito Ecology: Field Sampling Methods*, Springer Science and Business Media.

Sirot, L.K. et al. (2008), "Identity and transfer of male reproductive gland proteins of the dengue vector mosquito, *Aedes aegypti*: Potential tools for control of female feeding and reproduction", *Insect Biochemistry and Molecular Biology*, Vol. 38, No. 2, pp. 176-189.

Sivagnaname, N. and K. Gunasekaran (2012), "Need for an efficient adult trap for the surveillance of dengue vectors", *Indian Journal of Medical Research*, No. 136, No. 5, pp. 739-749.

Sota, T. and M. Mogi (1992), "Interspecific variation in desiccation survival time of *Aedes* (*Stegomyia*) mosquito eggs is correlated with habitat and egg size", *Oecologia*, Vol. 90, pp. 354-358.

Stouthamer, R., J.A. Breeuwer and G.D. Hurst (1999), "*Wolbachia pipientis*: Microbial manipulator of arthropod reproduction", *Annual Review of Microbiology*, Vol. 53, pp. 71-102.

Takken, W. (1991), "The role of olfaction in host-seeking of mosquitoes: A review", *Insect Science Applied*, Vol. 12, No. 1-2-3, pp. 287-295.

Takken, W. et al. (1998), "Effect of body size on host seeking and bloodmeal utilization in Anopheles gambiae sensu stricto (Diptera: Culicidae): The disadvantage of being small", *Journal of Medical Entomology*, Vol. 35, pp. 639-645.

Tamayo-Domínguez, R. (2011), "Comparación del ciclo gonotrófico de hembras de *Aedes aegypti* Linnaeus, 1762 y *Aedes albopictus* (Skuse, 1894) (Diptera: Culicidae) en condiciones de insectario en el Municipio de Tapachula, Chiapas" [Comparison of the gonotrophic cycle of females of *Aedes aegypti* Linnaeus, 1762 and *Aedes albopictus* (Skuse, 1894) (Diptera: Culicidae) under insectary conditions in the Municipality of Tapachula, Chiapas], *Degree thesis*, Universidad Autónoma de Chiapas, Tapachula.

Thavara, U. et al. (2001), "Larval occurrence, oviposition behavior and biting activity of potential mosquito vector of dengue in Samui Island, Thailand", *Journal of Vector Ecology*, Vol. 26, No. 2, pp. 172-180.

Thirión, I.J. (2003), *El Mosquito* Aedes aegypti *y el Dengue en México* [*The* Aedes aegypti *Mosquito and Dengue in Mexico*], Bayer Environmental Science, Bayer de México, S.A. de C. V., pp. 151.

Thomas, S.M. et al. (2012), "Low-temperature threshold for egg survival of a post-diapause and non-diapause European aedine strain, *Aedes albopictus* (Diptera: Culicidae)", *Parasites and Vectors*, Vol. 5, No. 100.

Tripet, F. et al. (2011), "Competitive reduction by satyrization? Evidence for interspecific mating in nature and asymmetric reproductive competition between invasive mosquito vectors", *The American Journal of Tropical Medicine and Hygiene*., Vol. 85, No. 2, pp. 265-270.

Trpis, M. et al. (1973), "Diel periodicity in the landing of *Aedes aegypti* on man", *Bulletin of the World Health Organization*, Vol. 48, pp. 623-629.

Tyagi, B.K. et al. (2006), "Dengue in Kerala: A critical review", ICMR Bulletin, Vol. 36, No. 4-5, pp. 13-22.

Ulloa, A. et al. (2010), "Cement tank: An artificial water container for *Aedes aegypti*", *Journal of the American Mosquito Control Association*, Vol. 26, No. 3, pp. 307.

Ulloa, G.A. (1996), "Abundancia larvaria y fuentes alimenticias de *Aedes aegypti* (L) (Diptera: Culicidae) en algunos recipientes artificiales en el sur de Chiapas, México" [Larval abundance and food sources of *Aedes aegypti* (L) (Diptera: Culicidae) in some artificial containers in southern Chiapas, Mexico], *Science Masters Thesis in Medical Entomology*, Universidad Autónoma de Nuevo León, San Nicolás de los Garza.

Van Handel, E. et al. (1994), "Plant-sugar, glycogen, and lipid assay of Aedes aegypti collected in urban Puerto Rico and rural Florida", *Journal of the American Mosquito Control Association*, Vol. 10, pp. 149-153.

Vezzani, D. and N. Schweigmann (2002), "Suitability of containers from different sources as breeding sites of *Aedes aegypti* (L.) in a cemetery of Buenos Aires city, Argentina", *Memórias do Instituto Oswaldo Cruz*, Vol. 97, No. 6, pp. 789-792.

Werren, J.H., L. Baldo and M.E. Clark (2008), "*Wolbachia*: Master manipulators of invertebrate biology", *Nature Reviews Microbiology*, Vol. 6, pp. 741-751.

West, A.S. and G.S. Eligh (1952), "The rate of digestion of blood in mosquitoes. Precipitin test studies", *Canadian Journal of Zoology*, Vol. 30, pp. 267-272.

Wheeler, D. (1996), "The role of nourishment in oogenesis", *Annual Review of Entomology*, Vol. 41, pp. 407-431.

Winch, P.J. et al. (1992), "Variation in *Aedes aegypti* larval indices over a one-year period in a neighborhood of Merida, Yucatan, Mexico", *Journal of the American Mosquito Control Association*, Vol. 8, No. 2, pp. 193-195.

Wong, J. et al. (2011), "Oviposition site selection by the dengue vector *Aedes aegypti* and its implications for dengue control", *PLoS Neglected Tropical Diseases*, Vol. 5, No. 4: e1015.

Ye, Y.H. et al. (2013), "*Wolbachia*-associated bacterial protection in the mosquito *Aedes aegypti*", *PLoS Neglected Tropical Diseases*, Vol. 7, No. 8: e2362.

Yen, J.H. and A.R. Barr (1973), "New hypothesis of the cause of cytoplasmic incompatibility in Culex pipiens L", *Nature*, Vol. 232, pp. 657-658.

Chapter 3. **Genetics of the mosquito *Ae. aegypti***

This chapter deals with the genetics of Aedes aegypti*, including genetic linkage to the physical map of the mosquito genome, its population genetics and its geographic distribution (phylogeography). Information is then provided on the genetics of insecticide susceptibility, the diverse modes of action and mechanisms of resistance to insecticides (metabolic, target-site, reduced penetration, behavioural avoidance). The last section focuses on the genetics of vector competence in* Ae. aegypti*, comprising the susceptibility of the mosquito populations to viruses and dengue virus in particular, the anatomic barriers to infection (midgut cells), the climatic factors affecting the infection susceptibility, the genetic variability and geographical variations in* Ae. aegypti *vector competence.*

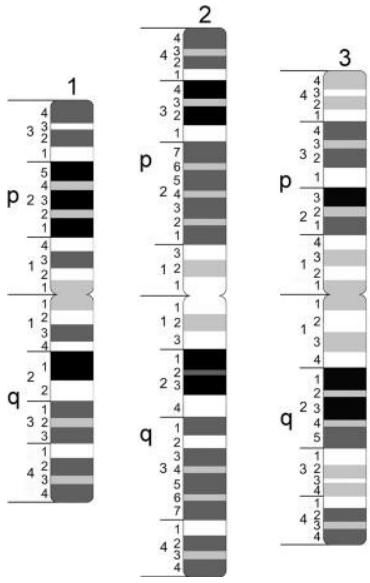
Linkage map organisation of *Ae. aegypti*

Ae. aegypti was the first mosquito species for which a detailed genetic linkage map was constructed and linked to the physical map (Craig and Hickey, 1967; Munstermann and Craig, 1979). Sequence-tagged site (STS) markers were developed for two different strategies, both based on physical maps using fluorescence in-situ hybridisation (FISH). The first mapping strategy used cosmids (8 RFLP markers) and the second strategy used cDNAs (21) (Brown et al., 2001). Recently, a band-based approach was used to perform a physical mapping of the *Ae. aegypti* genome to its mitotic chromosomes (Sharakhova et al., 2011; Timoshevskiy et al., 2013). The mitotic chromosome complement of *Ae. aegypti* consists of three pairs of metacentric chromosomes (Rai, 1963) that are numbered 1 (smallest), 2 (largest) and 3 (intermediate) (McDonald and Rai, 1970). *Ae. aegypti* sex determination alleles have been linked to the smallest homomorphic autosome 1 (McClelland, 1962). The three chromosomes of *Ae. aegypti* have been subdivided into a total of 23 regions and 94 subdivisions based upon staining of early metaphase chromosomes using YOYO-1 iodide (Timoshevskiy et al., 2014) (Figure 3.1). In addition to 100 genetic markers and 183Mb of genomic sequence, a marker linked with sex determination (Severson et al., 2002) and 12 quantitative trait loci (QTL) associated with pathogen transmission (Severson et al., 1995; Bosio, Fulton and Salasek, 2000; Gomez-Machorro, Bennett and Muñoz, 2004; Zhong et al., 2006) have been also anchored to the chromosomes (Timoshevskiy et al., 2014).

Although whole-genome sequencing has been undertaken (genome size of *Ae. aegypti* = 1.376 Gb), sequence compilation is still in progress due to the abundance of transposable elements (TEs) that cover approximately 50% of the *Ae. aegypti* genome (Nene et al., 2007; Severson and Behura, 2012; Timoshevskiy et al., 2014). TEs are extremely important to genome function and evolution (Arensburger et al., 2011) and may be key factors in mosquito genome plasticity. Low levels of polyteny resulting in poor quality of chromosome preparations add to the difficulty of using polytene chromosomes for physical mapping purposes in *Ae. aegypti* (Sharma et al., 1978; Campos, Andrade and Recco-Pimentel, 2003). Furthermore, the abundance of TEs complicates FISH experiments which require the use of unlabelled repetitive DNA fractions to block non-specific hybridisation. A physical map for *Ae. aegypti* corresponding to 13.3% of the genome was developed using FISH markers on mitotic chromosomes (Severson et al., 1993), genetic linked map (RFLP cDNA-based map) (Brown et al., 1995, 2001; Sharakhova et al., 2011), and QTL (Severson et al., 1994, 1995; Bosio, Fulton and Salasek, 2000; Gomez-Machorro, Bennett and Muñoz, 2004; Zhong et al., 2006). More recently, a more detailed physical map of the mosquito was constructed and a total of 624 Mb covered approximately 45% of the *Ae. aegypti* genome (Timoshevskiy et al., 2014).

Though this mosquito has a high TE load including miniature inverted-repeat transposable elements (MITEs) and piRNA biogenesis genes, its genome has a low proportion of transposon-specific piRNAs (Biryukova and Ye, 2015). This is important in preserving overall genome stability because the small RNA pathway controls TE mobilisation and movement (Saito and Siomi, 2010; Senti and Brennecke, 2010). Unregulated movement of active elements or non-autonomous sequences can lead to insertional mutagenesis through the genome resulting in a decrease in genetic fitness. Arensburger (2011) also stated that the stability of the transposons in *Ae. aegypti* is the result of a low proportion of transposon-specific piRNAs.

Figure 3.1. A physical map of the *Ae. aegypti* genome



Note: Chromosome regions and subdivisions are indicated on the left side of the idiograms.
Source: Timoshevskiy, V.A. et al. (2013), "An integrated linkage, chromosome, and genome map for the yellow fever mosquito *Aedes aegypti*", *PLOS Neglected Tropical Diseases*, Vol. 7, No. 2, pp. 2052; Timoshevskiy, V.A. et al. (2014), "Genomic composition and evolution of *Aedes aegypti* chromosomes revealed by the analysis of physically mapped supercontigs", *BMC Biology*, Vol. 12, pp. 27.

RNA interference (RNAi) is an important anti-viral defence mechanism. Although the *Ae. aegypti* genome encodes RNAi component orthologs, however, most populations of this mosquito are readily infected by, and subsequently transmit, arboviruses (Nene et al., 2007; Arensburger et al., 2011).

Population genetics and phylogeography of *Ae. aegypti*

The two *Ae. aegypti* subspecies: *Ae. aegypti formosus*, a wild mosquito apparently limited to sub-Saharan Africa, and *Ae. aegypti aegypti*, found globally in tropical and subtropical regions typically in association with humans (Moore et al., 2013), are described with their characteristics under the Classification (Taxonomy) section in Chapter 1. In addition, the Systematics section reports on the existence of two principal clades of *Ae. aegypti* collections worldwide.

In Mexico, local patterns of gene flow among *Ae. aegypti* populations were assessed using random amplified polymorphic DNA (RAPD) markers. Large genetic distances were observed, suggesting reduced gene flow among the mosquitoes (García-Franco et al., 2002; Gorrochotegui-Escalante et al., 2002; Muñoz et al., 2013a). The populations are panmictic along the Pacific coast, isolated by distance in northeast Mexico, and exhibit moderate gene flow across the Yucatan peninsula (Muñoz et al., 2013a). In the southern Pacific coast region reduced gene flow may result from sampling at altitudes greater than 1 500 m, which is close to the altitudinal limit for *Ae. aegypti* in Mexico (Lozano-Fuentes et al., 2009; Navarro et al., 2010), the mosquito being unable to survive at altitudes greater than 2 000 m (Lozano-Fuentes et al., 2012).

Sequence variation in the mitochondrial NADH dehydrogenase subunit 4 (ND4) gene, has been used to describe patterns of gene flow among *Ae. aegypti* s.l. collections within and among countries outside Africa (Gorrochotegui-Escalante et al., 2000, 2002; Bosio et al., 2005; Costa-da-Silva, 2005; Herrera et al., 2006; Bracco et al., 2007; Ribeiro et al., 2007; Paduan and Ribolla, 2008; Paupy et al., 2008; Urdaneta-Marquez et al., 2008; Dueñas et al., 2009; Hlaing et al., 2009; Lima and Scarpassa, 2009; Lozano-Fuentes et al., 2009; Paupy et al., 2012; Muñoz, 2013a; Moore et al., 2013). In Mexico, novel ND4 haplotypes were discovered and used to assess the amount of gene flow among breeding sites and to possibly predict the degree to which dengue virus (DENV) is transferred among sites (Gorrochotegui-Escalante et al., 2000, 2002; García-Franco et al., 2002).

To date 96 novel ND4 haplotypes have been discovered and three phylogenetic patterns have been consistently noted: either mtDNA haplotypes were distributed as two well-supported clades (Gorrochotegui-Escalante et al., 2000; Bosio et al., 2005; Lima and Scarpassa, 2009), or as a basal group similar to outgroup subspecies from which a second derived clade arises (Gorrochotegui-Escalante et al., 2002; Bracco et al., 2007; Paduan and Ribolla, 2008; Dueñas et al., 2009; Hlaing et al., 2009; Lozano-Fuentes et al., 2009; Paupy et al., 2012; Muñoz et al., 2013a). The broad distribution of specific haplotypes in Venezuela (Urdaneta-Marquez et al., 2008), Brazil (Bracco et al., 2007; Paduan and Ribolla, 2008; Lima and Scarpassa, 2009), Guatemala (Bracco et al., 2007), and Peru (Costa-da-Silva, 2005) demonstrates efficient mosquito dispersion in Central and South America.

Control practices are implicated as a major cause of genetic drift in *Ae. aegypti*. This was the conclusion of a study investigating 19 *Ae. aegypti* collections in Thailand, from Chiang Mai in the north to Songkhla province in the south (Bosio et al., 2005). That study found seven mitochondrial ND4 haplotypes, no evidence of isolation by distance, and low gene flow estimates among collections. They also concluded that these patterns are consistent with genetic drift arising from vector control efforts. Furthermore, polymorphisms were examined at 10 isoenzyme loci among 15 *Ae. aegypti* collections from Chiang Mai (Mousson et al., 2002). Low gene flow was also detected among these collections. These authors also concluded that this pattern was related to insecticide treatments. Additional studies further demonstrate the contribution of insecticide exposure to genetic drift in Martinique (Marcombe et al., 2009, 2012, 2013), Phnom Penh (Paupy et al., 2004) and French Guiana (Failloux et al., 2002). More on natural factors and human activities affecting gene flow or distribution is given under Chapter 4.

In summary, because *Ae. aegypti* is the primary global vector of severe viral diseases to humans (see Annexes A and B), it is crucial to study its population genetics in order to develop strategies to control the dispersion of the mosquito. Studies over the past 50 years have shown large differences among global populations of this species. Past studies based on morphological polymorphisms and allozymes were recently completed by the use of molecular genetic markers (including microsatellites and mitochondrial markers). Phylogenetic analyses consistently resolved two clades. In addition, phylogenetic analyses showed that populations of *Ae. aegypti* outside Africa consist of mosquitoes arising from two ancestral clades; one is basal and primarily associated with West Africa while the second arises from the first and contains primarily mosquitoes from East Africa. Across these many studies on population genetics, and those based on the distribution of the mosquito haplotypes around the world, it can be suggested that mosquito dispersion is very efficient, most likely due to commercial transportation and human movements.

Genetics of insecticide susceptibility and development of insecticide resistance

Resistance to insecticides

According to the World Health Organization (WHO, 2012a, 2012b), insecticide resistance is defined as the ability of an insect to withstand the effects of an insecticide by becoming resistant to its toxic effects by means of mutation and natural selection. Appropriate tools (biological, biochemical and/or molecular) are needed to identify the mechanisms involved in developing resistance and to conduct surveillance at individual and/or population levels.

Such resistance has been observed in more than 500 insect species worldwide, including more than 20 *Aedes* species (Diptera: Culicidae). Over 400 scientific reports worldwide document insecticide resistance in *Ae. aegypti*.

The large use of insecticides, and the resultant selection pressure on insect populations, has led to widespread resistance to all classes of insecticides among many invertebrate pests, making control difficult. Frequent applications of the same insecticide will select for those individuals in a population that are able to survive the recommended rates of the compounds owing to a genetically-fixed difference. Over time, this selection pressure will lead to a resistant population becoming established. In such cases, other compounds within the same class of chemistry are most often also affected; for instance, resistance to one pyrethroid type usually confers resistance across the whole group of pyrethroids, a phenomenon known as cross-resistance. Sometimes, depending on the nature of the resistance mechanism, multi-resistance can occur between different chemical classes, for example organophosphates and carbamates. The frequent treatments of crops with similar synthetic insecticides may also indirectly affect the susceptibility of insects of public health importance, with insect vectors additionally exposed when in the vicinity of agricultural sprays (Brogdon and McAllister, 1998; Hemingway and Ranson, 2000; Liu et al., 2006).

The portfolio of insecticides available for management of arthropod vectors (PAHO, 1994; WHOPES, 2005) is very limited and unlikely to increase dramatically in the near future. Development of resistance to commonly-used insecticides is therefore a serious threat to human ability to combat mosquito-borne diseases. Insecticide susceptibility must be viewed as a valuable "natural resource" at risk for being depleted. This underscores the critical importance of monitoring insecticide resistance through development and implementation of relevant management schemes. More information is contained in the section on "Prevention and management of insecticide resistance" of Annex A.

Insecticides, mode of action and resistance mechanisms

Vector control programmes include activities to control both immature and adult stages of *Ae. aegypti*. Chemical or biological larviciding and physical source reduction of container habitats are intended to control larvae, but house-to-house larval control is too laborious for sustainable implementation by vector control programmes or community participation (Reiter and Gubler, 1997; WHO, 2009; Horstick et al., 2010). During dengue outbreaks, outdoor and indoor spraying of insecticides is used to kill adults (WHO, 2009; Esu et al., 2010). Control measures are explored with more details in Annex A.

The four chemical classes of insecticides (organochlorides, organophosphates, carbamates, pyrethroids) used for larvae and adult mosquito control have their biochemical target sites in the insect central nervous system, which makes them fast-

acting killing agents. They act on only two different molecular target sites in the central nervous system, leading the insect to over-excitation and death. Organophosphates and carbamates both inhibit acetylcholinesterase (AChE), an enzyme of crucial importance in terminating nerve impulses by cleaving the natural neurotransmitter acetylcholine (Eto, 1974). In contrast, synthetic pyrethroids (and DDT, representing the organochlorides) modulate voltage-gated sodium channels, resulting in rapid knockdown properties (Khambay, 2002). It is important to note that these four chemical classes address only two different modes of action, so there is much less target-site diversity involved in the control of adult mosquitoes compared with the agricultural sector, which can rely on many more modes of action to date (Nauen and Bretschneider, 2002; Nauen, 2006). Insect growth regulator (IGR), pyriproxyfen is a juvenile hormone analogue that can be considered as an alternative to conventional insecticides because of its specific activity against immature insects, low persistence in the environment and virtually non-toxic to mammals (Madhu and Vijayan, 2009).

The various mechanisms that enable insects to resist the action of insecticides can be grouped into four distinct categories as follows: metabolic resistance, target-site resistance, reduced penetration, and behavioural avoidance.

Metabolic resistance

Metabolic resistance is the most common resistance mechanism that occurs in insects. This mechanism is based on the enzyme systems, which all insects possess to help them to detoxify naturally-occurring xenobiotics and insecticides. It is commonly accepted that insect detoxification systems derived from the plant-insect evolutionary arms race, and several insect detoxification enzymes have been associated with the detoxification of plant toxins and all types of chemicals, including insecticides (Despres, David and Gallet, 2007). Over-expression of enzymes capable of detoxifying insecticides or amino acid substitutions within these enzymes, which alter the affinity of the enzyme for the insecticide, can result in high levels of insecticide resistance (Hemingway et al., 2004; Flores et al., 2005, 2006). Increased expression of the genes encoding the major xenobiotic metabolising enzymes is the most common cause of insecticide resistance in mosquitoes. Over-expression of detoxifying enzymes can occur as the result of gene amplification (e.g. duplication) or due to changes in either transacting regulator elements or the promoter region of the gene (Guillemaud et al., 1997; Hemingway and Ranson, 2000; Hawkes and Hemingway, 2002). The consequence is a significant increase in enzyme production in resistant insects that enables them to metabolise or degrade insecticides before they are able to exert a toxic effect.

Three enzyme families (with a variable number of gene members), the cytochrome P450 monooxygenases (P450s), glutathione transferases (GST), and carboxyl/cholinesterases (CCE) are implicated in insecticide metabolism. Each of these catalyses a wide range of detoxification reactions. They are the primary enzymatic defence against xenobiotics, are responsible for the removal of many by-products of metabolism, play essential roles in multiple biosynthetic pathways and are involved in chemical communication (Feyereisen, 2005; Oakeshott et al., 2005; Ranson and Hemingway, 2005). Some individual enzymes also have structural roles instead of, or in addition to, their catalytic activity. This diversity in the function of each enzyme family is accomplished by a mixture of highly specialised enzymes, often with specific substrates and strictly regulated expression profiles, and more generalist, ubiquitously expressed enzymes. Many insect species show an amazing diversity of detoxification enzymes. As insect genomes have been sequenced, and the detoxification genes annotated, it has become

apparent that these detoxification gene families are very rapidly evolving and each insect has a unique complement of detoxification genes, with very few orthologs across insect species (Ranson et al., 2002; Claudianos et al., 2006). The rapid expansion and diversification of detoxification genes likely facilitated the adaptation of insects to their particular ecological niches, and, on a more recent evolutionary timescale, has enabled them to survive various man-made xenobiotics, including insecticides. A small subset of the detoxification genes has been previously described in *Ae. aegypti* (Sieglaff, Duncan and Brown, 2005; David et al., 2006; Lumjuan et al., 2007).

Target-site resistance

Target-site resistance is the second most common mechanism of resistance to insecticides encountered in insects. Insecticides (e.g. organophosphates, carbamates, DDT and pyrethroids) generally act at a specific site within the insect, typically within the nervous system. The site of action can be modified in resistant insect strains such that the insecticide no longer binds effectively.

Reduced sensitivity of the target receptors to insecticide results from non-silent point mutations in the gene encoding the protein constituting the target site. For example, the target site for organophosphate and carbamate insecticides is AChE in the nerve cell synapses. Several mutations in the AChE gene have been found in insects (Fournier, 2005), which result in reduced sensitivity to inhibition of the enzyme by these insecticides (Weill et al., 2003).

Alterations in the target site that cause resistance to pyrethroids and DDT are often referred to as *knockdown resistance* (kdr), in reference to the ability of insects with relevant alleles to withstand prolonged exposure to insecticides without being 'knocked-down'. The kdr is conferred principally by non-synonymous mutations in the voltage-gated sodium channel gene that reduce insecticide binding to this channel in the insect nerve sheath, thereby preventing the loss of co-ordinated activity and paralysis in the insect (Soderlund and Knipple, 2003; Davies et al., 2007; Rinkevich, Du and Dong, 2013).

Worldwide, numerous kdr-conferring voltage-gated sodium channel allele mutations (e.g. S989P; I1,011M; I1,011V; V1,016I; V1,016G; F1,534C; and D1,794Y) have been described in *Ae. aegypti* (Vontas et al., 2012; Rinkevich, Du and Dong, 2013). Multiple kdr mutations had been reported from Mexico, the Caribbean, and Central America (Saavedra-Rodriguez et al., 2007). Subsequent studies reported the presence of kdr conferring mutations in *Ae. aegypti* collections from Brazil (Martins et al., 2009a, 2009b; Lima et al., 2011; Belinato, Martin and Valle, 2012), Mexico (Ponce-Garcia et al., 2009; Siller et al., 2011; Aponte et al., 2013) and the Caribbean (Marcombe et al., 2009, 2012, 2013; Harris, Rajatileka and Ranson, 2010; Bariami et al., 2012; McAllister, Godsey and Scott, 2012; Maestre-Serrano et al., 2014; Alvarez et al., 2015).

Reduced penetration

Reduced penetration (and behavioural resistance by reduced penetration) occurs when insects develop a heritable mechanism(s) that reduces or prevents the entry of a toxin into the insect's body. Modifications in the cuticle or digestive tract linings that prevent or slow the penetration/absorption of insecticides can be found in some resistant insects. This resistance mechanism is non-specific and can affect the effectiveness of a broad range of insecticides. Reduced uptake of insecticide, often referred to as cuticular resistance, is frequently described as a minor resistance mechanism. Certainly, for pests

where the major route of insecticide delivery is via ingestion, this is likely to be the case. However, for dengue control, where insecticides are typically applied spatially or on wall surfaces, uptake of insecticides is primarily through the appendages. An increase in the thickness of the tarsal cuticle, or a reduction in its permeability to lipophilic insecticides, could have a major impact on the bioavailability of an insecticide *in vivo*.

Reduced cuticle penetration is the least understood resistance mechanism. Though it may have a primary role in resistance (Valles, Dong and Brenner, 2000; Ahmad, Denholm and Bromilow, 2006; Puinean et al., 2010), it more often acts in combination with the other mechanism(s).

Behavioural avoidance

Insecticide resistance in mosquitoes may also be conferred by behavioural changes in response to prolonged exposure to an insecticide. Behavioural avoidance does not have the same importance as physiological resistance but may be considered to be a contributing factor, leading to the avoidance of lethal doses of an insecticide (Chandre et al., 2000; Grieco et al., 2007). This type of response can be further divided into direct contact excitation (sometimes referred to as "irritancy"), and non-contact spatial repellency when insects move away from the insecticide-treated area before making direct contact (Chareonviriyaphap et al., 1997; Grieco et al., 2007).

To better approximate insect behaviour in natural field settings, numerous experiments have been made over many decades using specially constructed experimental huts (Smith, 1965; Rozendaal et al., 1989; Roberts and Alecrim, 1991; Bangs, 1999; Grieco et al., 2000, 2007; Polsomboon et al., 2008; Malaithong et al., 2010). Most experimental hut studies have been conducted to observe the behaviour of *Anopheles* mosquitoes; however, Grieco et al. (2007) successfully demonstrated that chemical actions could be observed in experimental huts using *Ae. aegypti* as a model system. The results obtained from both laboratory and field studies can help facilitate the choice of the most effective chemicals and measures to control house-frequenting adult mosquitoes.

Conclusion on resistance to insecticides

Insecticide resistance develops in an insect population when individuals carrying genes that allow them to survive exposure to the insecticide survive, mate and pass these genes onto the next generation. Thus, any activities that control the individuals with the resistance trait will delay the spread of the resistance genes in the population. Some elements on prevention and management of insecticide resistance are given in Annex A.

Genetics of vector competence in *Ae. aegypti*

Vector competence, a key factor

The vector competence (VC) is defined as the intrinsic permissiveness of an arthropod vector for infection, dissemination and transmission of a pathogen (Black et al., 2002; Dickson et al., 2014). The full competence of a vector is determined not only by its ability to become infected, but also by its ability to transmit a pathogen.

VC in *Ae. aegypti* is an element of primary importance to consider because of this mosquito's public health impact as main potential transmitter of dengue, yellow fever, Zika and chikungunya viruses. In a few restricted areas, *Ae. aegypti* is also a vector of

Wuchereria bancrofti and *Brugia malayi*, both of which cause lymphatic filariasis or elephantiasis (Service, 2012; Powell and Tabachnick, 2013)

Consequently, *Ae. aegypti* has been the subject of numerous vector competence and population genetic studies (Aitken, Downs and Shope, 1977; Gubler et al., 1979; Tabachnick and Powell, 1979; Rosen et al., 1985; Tabachnick et al., 1985; Tardieux et al., 1990; Miller and Mitchell, 1991; Apostol, Reiter and Miller, 1996; Bosio and Beaty, 1998; Vazeille-Falcoz et al., 1999; Bosio, Fulton and Salasek, 2000; Bennett et al., 2002b; Gorrochotegui-Escalante et al., 2002; Mercado-Curiel, Black and Muñoz, 2008; Lozano-Fuentes et al., 2009; Sylla et al., 2009; Lambrechts, 2011; Lambrechts et al., 2011; Guo et al., 2013; Muñoz et al., 2013b; Chepkorir et al., 2014; Diagne et al., 2014; Dickson et al., 2014; Gonçalves et al., 2014; Vega-Rúa et al., 2014).

Susceptibility of **Ae. aegypti** *populations to viruses*

Infection with dengue virus

Ae. aegypti becomes infected with a viral disease, dengue for example, when the mosquito bites and acquires a blood meal from a dengue virus (DENV)-infected human, the primary host of the virus. The mosquito infection depends on factors such as DENV virulence, physical barriers and innate immunity that can confer resistance or susceptibility of an *Ae. aegypti* population to viruses.

The relationships between DENV and its arthropod vector *Ae. aegypti* are crucial, and an analysis of host cell responses to flavivirus infection of mosquito vectors is very important for understanding the maintenance and transmission of the disease.[1] Mosquito populations differ in their susceptibility to flavivirus development (i.e. VC), reflecting the different barriers encountered by the virus from its entry into the mosquito to its release in the saliva. Factors such as specific mosquito epithelial cells receptors, as well as differential viral replication in the mosquito, are critical for VC - as are other genes as exhibited by quantitative trait loci (QTL) studies (Gomez-Machorro, Bennett and Muñoz, 2004).

Three *Ae. aegypti* strains with different susceptibilities to DENV infection have been reported (namely DS3, DMEB and IBO-11), and these have been used to study whether midgut cell receptors for DENV may be markers for VC (Bennett and Beaty, 2005). *Ae. aegypti* susceptibility to DENV, attributable to multiple genetic factors, is found to be usually very high, particularly against DENV-2,[2] as elicited by DS3 and DMEB strains.

It has been observed that the three strains showed a difference in the degree of infection of their midgut (MG) cells, depending on their susceptibility to DENV. For example, the IBO-11 strain expressed almost no infection remaining after 26 days post-infection. DMEB strain showed increase in infection up to 26 hours in all the three MG regions, having the maximal virus accumulation in the posterior MG which then diminished by 14 days post-infection, compared to the other susceptible strain DS3 that has maximal virus accumulation in anterior MG at 14 days post-infection. These results also display a statistically-significant MG infection increase from the first five hours post-infection to 26 hours in DS3 and DMEB strains ($p < 0.05$). Moreover, IBO-11 strain exhibited a significant decrease ($p < 0.05$) of MG infection from 5 to 336 hours post-infection. Virus infection of IBO-11 strain was almost completely abolished ($p < 0.05$) from 13 to 336 hours post-infection (Mercado-Curiel, Black and Muñoz, 2008). The susceptibility, resistance and refractoriness depend on multiple genetic factors (Miller and Mitchell, 1991).

Anatomic barriers to infection (midgut cells)

The VC for arboviruses is associated with a number of anatomic barriers to productive vector infection. These include a midgut infection barrier (MIB), a midgut escape barrier (MEB) and a salivary gland barrier (Black et al., 2002). In potential vectors provided with an MIB, a virus cannot infect and/or replicate in the mosquito MG cells. This may be due to a lack of specific cell surface receptors for the virus or to MG cells being non-permissive for infection with the virus (Mercado-Curiel, Black and Muñoz, 2008). Potential vectors provided with an MEB may allow virus replication in the MG, even to high titres (concentrations), but the virus is then unable to exit the MG to cause a disseminated infection. The VC for flaviviruses in *Ae. aegypti* is thought to be controlled by at least two genes or sets of genes, one controlling the MIB and the other controlling the MEB (Miller and Mitchell, 1991; Bosio and Beaty, 1998; Bosio, Fulton and Salasek, 2000). A study by Bennett et al. (2002b) also concluded that these barriers are probably major determinants of VC to DENV in nature and during experimental infections.

Bosio and Beaty (1998) proposed a significant additive genetic effect in MIB and demonstrated that the DENV titre in the mosquito MG and head did not correlate with the rate of infection. They also showed that the heritability for virus titres in tissues (MG or head) were almost identical in different strains of *Ae. aegypti formosus* and showed that the amount of virus in the MG did not determine if the virus was disseminated, which hypothetically may be due to the presence or absence of DENV receptors (in the MG, in particular).

Barriers to infection can vary widely in prevalence among *Ae. aegypti* populations, leading to large intraspecific variation of *Ae. aegypti* VC that may influence the epidemiology of DENV and other flaviviruses (Black et al., 2002).

Climatic factors affecting the infection susceptibility

Susceptibility of *Ae. aegypti* mosquito to DENV varies geographically and can be influenced by climatic factors such as temperature, which affect the incidence, seasonality and distribution of vector-borne diseases.

The VC has shown to be affected by temperature, which impacts biological processes of mosquitoes including their interaction with viruses (Watts et al., 1987; Lambrechts et al., 2011). Chepkorir et al. (2014) demonstrated a significantly higher infection rate at high temperatures for mosquitoes collected in Nairobi and Kilifi (Kenya), which is consistent with previous results (Watts et al., 1987). The 2014 study showed that the Nairobi *Ae. aegypti* population is a relatively inefficient vector for DENV-2 compared to that from Kilifi with the former showing high infection, but low dissemination rates in low- and high-temperature settings. These results also suggested a weak MIB and a strong MEB for the Nairobi population, and a moderate MIB but weak MEB for the Kilifi population.

Genetic variability and geographical variations in* Ae. aegypti *vector competence

A number of genetic studies on VC were conducted worldwide, which demonstrated a great VC variability. The results of some of these works performed in different countries are presented hereinafter. All in all, they indicate that vector control strategies should be adapted to the available data for each region. Further analysis should be

conducted to better understand the reasons for this large variability in VC and how these parameters correlate with epidemiological findings (Gonçalves et al., 2014).

Ae. aegypti populations exhibit considerable genetic variability in VC for flaviviruses, including DENV-2 viruses. The range of VCs shown suggests that the ability to overcome the MIB and MEB to transmit DENV-2 JAM1409 is a quantitative trait with multiple genes that likely condition VC and collectively determine the infection rates of mosquito populations. This theory has been studied using crosses of susceptible and refractory mosquito lines (Miller and Mitchell, 1991; Bosio and Beaty, 1998).

Significant genetic variation in *Ae. aegypti* on a smaller scale has been demonstrated in Puerto Rico (Apostol, Reiter and Miller, 1996) and in Mexico (Gorrochotegui-Escalante et al., 2000). Subsequently, the potential variation in VC on a regional geographic scale was addressed in Mexico. The major aim of this research was to determine if genetic variability in *Ae. aegypti* populations conditions the incidence and severity of dengue fever and dengue haemorrhagic fever outbreaks. Such study can help identify genetic biomarkers for mosquito populations that pose undue risk for severe diseases and allow control programmes to focus their resources on areas at greatest risk.

Several studies have shown that *Ae. aegypti* has a continuous variation in its competence to transmit flavivirus (Bennett et al., 2002a; Black et al., 2002; Severson et al., 2004; Gorrochotegui-Escalante et al., 2005). *Ae. aegypti* from 24 collections in Mexico and the United States were challenged orally with DENV-2 JAM1409, and the VC of the populations ranged from 24% to 83%. In general, the *Ae. aegypti* collections from throughout Mexico exhibited considerable variability in VC, and collections from the Yucatan Peninsula were generally more competent than those from other geographic regions (Bennett et al., 2002b). Lozano-Fuentes et al. (2009) showed that the Neovolcanic Axis (NVA) in Mexico is a natural barrier to *Ae. aegypti* VC for DENV, as a much lower VC (20%) prevails for mosquito populations from south of the NVA compared to mosquitoes collected from north of the NVA (55%).

Ae. aegypti populations from Belo Horizonte, Brazil, exhibited wide variation in VC to transmit dengue. Most Brazilian states are infested with *Ae. aegypti* and are consequently at risk of dengue transmission (Figueiredo et al., 2008). Moncayo et al. (2004) studied populations from various geographical locations and showed that *Ae. aegypti* from Galveston, Texas (United States) were more susceptible than those from Bolivia but were less susceptible than mosquitoes from Thailand. This concurred with the observations made by Bennett et al. (2002b) on *Ae. aegypti* collected from various locations in Mexico and by Chepkorir et al. (2014) on populations from two different Kenyan sites, that all differed significantly in their MG susceptibility to infection.

The VC studies on *Ae. aegypti* from West Africa have shown that mosquitoes are more refractory for both DENV (Tabachnick et al., 1985; Bosio and Beaty, 1998) and yellow fever virus (YFV) (Tabachnick et al., 1985; Miller and Mitchell, 1991) compared to *Ae. aegypti* collected from the Americas or Asia (Diallo et al., 2005, 2008).

In studies directly comparing collections within Senegal, wide variation in VC was shown for both high-passage (Sylla et al., 2009) and low-passage field isolates of DENV-2 (Diallo et al., 2005, 2008). Especially, sylvatic collections from south-eastern Senegal were more refractory than other collections from throughout the country. However, it should be noted that the study by Sylla et al. (2009) only examined the highly passaged DENV-2 Jam1409 isolate (reinforcing the importance of using strains circulating in the geographic area of study). Previous studies demonstrated that geographically-distinct

collections of *Ae. aegypti* from Senegal are genetically diverse and documented the great variability in VC for both DENV-2 and YFV across the country. The northwest-southeast decline in the susceptibility to YFV BA-55 is very similar to that seen with DENV-2 JAM1409 (Huber et al., 2008; Sylla et al., 2009).

Furthermore, it has been demonstrated that VC of *Ae. aegypti* for DENV is dependent on the interactions between the mosquito strain and virus genotype in natural collections (Lambrechts et al., 2009). Using viruses and vectors that are geographically proximate and genetically diverse is important in order to make strong conclusions about VC between collections. Assessing VC in *Ae. aegypti* with a viral isolate collected in proximity seems to be the most informative approach (Lambrechts et al., 2009). Diallo et al. (2005, 2008) followed this process by examining the VC of *Ae. aegypti* from Senegal with multiple local isolates of DENV-2. The 2008 study reported low levels of MG infection (0.0–26.3%) and variable disseminated infection (0-100%) in six collections from Senegal regardless of geographic location. Both studies demonstrated variability in infection rates based on the isolate of DENV-2 and the collection site, confirming the local adaptation between the virus and the mosquito vector. They also showed that although collections of sylvatic *Ae. aegypti* presented lower infection rates than sylvatic *Aedes* from other species, some sylvatic *Ae. aegypti* mosquitos developed nevertheless a disseminated infection.

Similarly, Lambrechts et al. (2009) demonstrated that differences in VC among three *Ae. aegypti* collections from Thailand infected with three genotypes of DENV-1 was a result of mosquito and virus genotypes interactions. Dickson et al. (2014) also highlighted interactions between mosquito and virus genotypes in sylvatic *Ae. aegypti*, and with YFV in West Africa. Overall, VC was dependent upon both viral and vector strains. Importantly, and contrary to previous studies, the study by Dickson et al. (2014) reported that sylvatic collections of *Ae. aegypti* showed high levels of disseminated infection for local isolates of both DENV-2 and YFV.

Recently, VC of *Ae. aegypti* for chikungunya virus (CHIKV) has been investigated and it was found that *Ae. aegypti* populations from Cape Verde and Kedougou (Senegal) were competent for CHIKV, but *Ae. aegypti* from Dakar (Senegal) presented a low susceptibility to the virus. The virus strains belonging to the West African lineage were the only ones disseminated by the domestic population of *Ae. aegypti* from Dakar and transmitted by those from Cape Verde (Diagne et al., 2014). And as previously demonstrated in Kerala, India (Kumar et al., 2012), it has been also observed that *Ae. albopictus* was a better vector for this virus (CHIKV) than *Ae. aegypti* (Diagne et al., 2014).

In addition to DENV, CHIKV and YFV, *Ae. aegypti* is also a vector for Zika virus (ZIKV), a single-stranded RNA virus belonging to the Flaviviridae family (Faye et al., 2014; Abushouk, Negida and Ahmed, 2016). More details on ZIKV and related infection are given in Annex B, section “Virus infection vectored by mosquitoes”. The VC for ZIKV in *Ae. aegypti* is variable, depending on the source of the mosquitoes and the virus strain. *Ae. aegypti* collected from French Polynesia displayed high ZIKV infection rate, but late ability to transmit the virus (Richard, Paoaafaite and Cao-Lormeau, 2016). *Ae. aegypti* from Singapore infected with ZIKV demonstrated high MG infection, resulting in the salivary glands of more than half of the mosquitoes being tested positive for ZIKV (62%) by Day 5, and all mosquitoes potentially infective by Day 10 (Li et al., 2012).

Summary on vector competence

In summary, *Ae. aegypti* is a most efficient vector for several deadly (e.g. dengue, yellow fever) and debilitating (e.g. chikungunya, Zika) arthropod-borne diseases. The vector competence (VC) for arboviruses is associated with a number of anatomic barriers to productive vector infection. The VC for flaviviruses in *Ae. aegypti* is thought to be controlled by at least two genes or sets of genes, one controlling the midgut infection barrier (MIB) and the other controlling the midgut escape barrier (MEB). The mosquito susceptibility to DENV, attributable to multiple genetic factors, is usually very high, particularly against DENV-2 as elicited by DS3 and DMEB strains. The ability to overcome the MIB and MEB to transmit DENV-2 JAM1409 is a quantitative trait with multiple genes that likely condition VC and collectively determine the infection rates of mosquito populations. The rate of midgut infection, midgut escape and salivary gland infection generally increases at higher temperature, though may vary amongst different populations. Natural geographic features may also act as a barrier to gene flow in varied *Ae. aegypti* populations (for VC) for DENV-2.

VC differences among different populations infected with genotypes of DENV result from interactions between mosquito and virus genotypes. At subspecies level, *Ae. aegypti aegypti* is generally far more efficient in transmitting DENV in urban agglomerations than the sylvatic *Ae. aegypti formosus*, albeit with a variation in VC.

While *Ae. aegypti* is unquestionably a much stronger potential transmitter for dengue, yellow fever and Zika viruses, it seems that *Ae. albopictus* would be a more efficient vector for CHIKV, especially in sylvatic and rural settings. Both mosquitoes are day biters, multiple feeders and capable to transmit several pathogens.

Notes

[1] More information on flavivirus infection can be found in Annex B. Human and animal health affected by mosquitoes.

[2] The four viral serotypes of DENV are explained in Annex B. Human and animal health affected by mosquitoes.

References

Abushouk, A.I., A. Negida and H. Ahmed (2016), "An updated review of Zika virus", *Journal of Clinical Virology*, Vol. 84, pp. 53-58.

Ahmad, M., I. Denholm and R.H. Bromilow (2006), "Delayed cuticular penetration and enhanced metabolism of deltamethrin in pyrethroid-resistant strains of *Helicoverpa armigera* from China and Pakistan", *Pest Management Science*, Vol. 62, pp. 805-810.

Aitken, T.H., W.G. Downs and R.E. Shope (1977), "*Aedes aegypti* strain fitness for yellow fever virus transmission", *The American Journal of Tropical Medicine and Hygiene*, Vol. 26, No. 5, pp. 985-989.

Alvarez, L.C. et al. (2015), "Frequency of V1016I and F1534C mutations in the voltage-gated sodium channel gene in *Aedes aegypti* in Venezuela", *Pest Management Science*, Vol. 71, No. 6, pp. 863-869.

Aponte, H.A. et al. (2013), "The pyrethroid resistance status and mechanisms in *Aedes aegypti* from the Guerrero state, Mexico", *Pesticide Biochemistry and Physiology*, Vol. 107, pp. 226-234.

Apostol, B.L., P. Reiter and B.R. Miller (1996), "Population genetics with RAPD-PCR markers: The breeding structure of *Aedes aegypti* in Puerto Rico", *Heredity*, Vol. 76, No. 4, pp. 325-334.

Arensburger, P. et al. (2011), "The mosquito *Aedes aegypti* has a large genome size and high transposable element load but contains a low proportion of transposon-specific piRNAs", *BMC Genomics*, Vol. 12, pp. 606.

Bangs, M.J. (1999), *The Susceptibility and Behavior of Anopheles Albimanus Weidemann and Anopheles Vestitipennis Dyar and Knab (Diptera: Culicidae) to Insecticides in Northern Belize, Central America*, Uniformed Services University of the Health Sciences, Bethesda (USA).

Bariami, V. et al. (2012), "Gene amplification, ABC transporters and cytochrome P450s: Unraveling the molecular basis of pyrethroid resistance in the dengue vector, *Aedes aegypti*", *PLoS Neglected Tropical Diseases*, Vol. 6, No. 6: e1692.

Belinato, T.A., A.J. Martins and D. Valle (2012), "Fitness evaluation of two Brazilian *Aedes aegypti* field populations with distinct levels of resistance to the organophosphate temephos", *Memórias do Instituto Oswaldo Cruz.*, Vol. 107, pp. 916-922.

Bennett, K.E. and B.J. Beaty (2005), "Selection of D2S3, an *Aedes aegypti* (Diptera: Culicidae) strain with high oral susceptibility to Dengue 2 virus and D2MEB, a strain with a midgut barrier to Dengue 2 escape", *Journal of Medical Entomology*, Vol. 42, No. 2, pp. 110-119.

Bennett, K.E. et al. (2002a), "Flavivirus susceptibility in *Aedes aegypti*", *Archives of Medical Research*, Vol. 33, No. 4, pp. 379-388.

Bennett, K.E. et al. (2002b), "Variation in vector competence for dengue 2 virus among 24 collections of *Aedes aegypti* from Mexico and the United States", *The American Journal of Tropical Medicine and Hygiene*, Vol. 67, No. 1, pp. 85-92.

Biryukova, I. and T. Ye (2015), "Endogenous siRNAs and piRNAs derived from transposable elements and genes in the malaria vector mosquito *Anopheles gambiae*", *BMC Genomics*, Vol. 16, pp. 278.

Black, W.C. 4th et al. (2002), "Flavivirus susceptibility in *Aedes aegypti*", *Archives of Medical Research*, Vol. 33, No. 4, pp. 379-388.

Bosio, C.F. et al. (2005), "Genetic structure of *Aedes aegypti* populations in Thailand using mitochondrial DNA", *The American Journal of Tropical Medicine and Hygiene*, Vol. 72, No. 4, pp. 434-442.

Bosio, C.F. and B.J. Beaty (1998), "Quantitative genetics of vector competence for dengue-2 virus in *Aedes aegypti*", *The American Journal of Tropical Medicine and Hygiene*, Vol. 59, No. 6, pp. 965-970.

Bosio, C.F., R.E. Fulton and M.L. Salasek (2000), "Quantitative trait loci that control vector competence for dengue-2 virus in the mosquito *Aedes aegypti*", *Genetics*, Vol. 156, No. 2, pp. 687-698.

Bracco, J.E. et al. (2007), "Genetic variability of *Aedes aegypti* in the Americas using a mitochondrial gene: Evidence of multiple introductions", *Memórias do Instituto Oswaldo Cruz*, Vol. 102, No. 5, pp. 573-580.

Brogdon, W.G. and J.C. McAllister (1998), "Insecticide resistance and vector control", *Emerging Infectious Diseases*, Vol. 4, pp. 605-613.

Brown, S.E. et al. (2001), "Integration of the *Aedes aegypti* mosquito genetic linkage and physical maps", *Genetics*, Vol. 157, No. 3, pp. 1299-1305.

Brown, S.E. et al. (1995), "Toward a physical map of *Aedes aegypti*", *Insect Molecular Biology*, Vol. 4, No. 3, pp. 161-167.

Campos, J., C.F. Andrade and S.M. Recco-Pimentel (2003), "A technique for preparing polytene chromosomes from *Aedes aegypti* (Diptera, Culicinae)", *Memórias do Instituto Oswaldo Cruz.*, Vol. 98, No. 3, pp. 387-390.

Chandre, F. et al. (2000), "Modifications of pyrethroid effects associated with kdr mutation in *Anopheles gambiae*", *Medical and Veterinary Entomology*, Vol. 14, pp. 81-88.

Chareonviriyaphap, T. et al. (1997), "Pesticide avoidance behavior in *Anopheles albimanus*, a malaria vector in the Americas", *Journal of the American Mosquito Control Association*, Vol. 13, pp. 171-183.

Chepkorir, E. et al. (2014), "Vector competence of *Aedes aegypti* populations from Kilifi and Nairobi for dengue 2 virus and the influence of temperature", *Parasit Vectors*, Vol. 7, No. 1, pp. 435.

Christophers, S.R. (1960), Aedes aegypti *(L.) The Yellow Fever Mosquito. Its Life History, Bionomics and Structure*, Cambridge University Press, Cambridge.

Claudianos, C. et al. (2006), "A deficit of detoxification enzymes: Pesticide sensitivity and environmental response in the honeybee", *Insect Molecular Biology*, Vol. 15, pp. 615-663.

Costa-da-Silva, A.L. (2005), "Genetic lineages in the yellow fever mosquito *Aedes* (Stegomyia) *aegypti* (Diptera: Culicidae) from Peru", *Memórias do Instituto Oswaldo Cruz*, Vol. 100, No. 6, pp. 539-544.

Craig, G.B. Jr and W.A. Hickey (1967), "Current status of the formal genetics of *Aedes aegypti*", *Bulletin of the World Health Organization*, Vol. 36, No. 4, pp. 559-562.

David, J.P. et al. (2006), "Involvement of cytochrome P450 monooxygenases in the response of mosquito larvae to dietary plant xenobiotics", *Insect Biochemistry and Molecular Biology*, Vol. 36, pp. 410-420.

Davies, T.G.E. et al. (2007), "DDT, pyrethrins, pyrethroids and insect sodium channels", *IUBMB Life*, Vol. 59, pp. 151-162.

Despres, L., J.P. David and C. Gallet (2007), "The evolutionary ecology of insect resistance to plant chemicals", *Trends in Ecolpgy & Evolution*, Vol. 22, pp. 298-307.

Diagne, C.T. et al. (2014), "Vector competence of *Aedes aegypti* and *Aedes vittatus* (Diptera: Culicidae) from Senegal and Cape Verde archipelago for West African lineages of chikungunya virus", *The American Journal of Tropical Medicine and Hygiene*, Vol. 91, No. 3, pp. 635-641.

Diallo, M. et al. (2008), "Vector competence of *Aedes aegypti* populations from Senegal for sylvatic and epidemic dengue 2 virus isolated in West Africa", *Transactions of the Royal Society of Tropical Medicine and Hygiene*, Vol. 102, No. 5, pp. 493-498.

Diallo, M. et al. (2005), "Potential role of sylvatic and domestic African mosquito species in dengue emergence", *The American Journal of Tropical Medicine and Hygiene*, Vol. 73, No. 2, pp. 445-449.

Dickson, L.B. et al. (2014), "Vector competence in West African *Aedes aegypti* Is Flavivirus species and genotype dependent", *PLoS Neglected Tropical Diseases*, Vol. 8, No. 10: e3153.

Dueñas, J.C. et al. (2009), "Two different routes of colonization of *Aedes aegypti* in Argentina from neighboring countries", *Journal of Medical Entomology*, Vol. 46, No. 6, pp. 1344-1354.

Esu, E. et al. (2010), "Effectiveness of peridomestic space spraying with insecticide on dengue transmission; Systematic review", *Tropical Medicine and International Health*, Vol. 15, pp. 619-631.

Eto, M. (1974), *Organophosphorus Pesticides: Organic and Biological Chemistry*, CRC Press, Boca Raton.

Failloux, A.B. et al. (2002), "Differentiation of *Aedes aegypti* populations in French Guiana", *Medical and Veterinary Entomology*, Vol. 16, No. 4, pp. 456-460.

Faye, O. et al. (2014), "Molecular evolution of Zika virus during its emergence in the 20th century", *PLoS Neglected Tropical Diseases*, Vol. 8: e2636.

Feinson, F.M. and A. Spielman (1980), "Nutrient mediated juvenile hormone secretion in mosquitoes", *Journal of Insect Physiology*, Vol. 26, pp. 113-117.

Feyereisen, R (2005), "Insect cytochrome P450", in L.I. Gilbert, K. Latrou and S.S. Gill (eds.), *Comprehensive Molecular Insect Science*, Elsevier, Oxford, pp. 1-77.

Figueiredo, R.M. et al. (2008), "Dengue virus type 4, Manaus, Brazil", *Emerging Infectious Diseases*, Vol. 14, No. 4, pp. 667-669.

Flores, A.E. et al. (2006), "Mechanisms of insecticide resistance in field populations of *Aedes aegypti* (L.) from Quintana Roo, Southern Mexico", *Journal of the American Mosquito Control Association*, Vol. 22, No. 4, pp. 672-677.

Flores, A.E. et al. (2005), "Elevated alfa-esterases levels associated with permethrin tolerance in *Aedes aegypti* (L.) from Baja California, Mexico", *Pesticide Biochemistry and Physiology*, Vol. 82, pp. 66-78.

Fournier, D. (2005), "Mutations of acetylcholinesterase which confer insecticide resistance in insect populations", *Chemico-Biological Interactions*, pp. 157-158 and 257-261.

García-Franco, F. et al. (2002), "Large genetic distances among *Aedes aegypti* populations along the South Pacific coast of Mexico", *The American Society of Tropical Medicine and Hygiene*, Vol. 6, pp. 594-598.

Gomez-Machorro, C., K.E. Bennett and M.deL. Muñoz (2004), "Quantitative trait loci affecting dengue midgut infection barriers in an advanced intercross line of *Aedes aegypti*", *Insect Molecular Biology*, Vol. 13, No. 6, pp. 637-648.

Gonçalves, C.M. et al. (2014), "Distinct variation in vector competence among nine field populations of *Aedes aegypti* from a Brazilian dengue-endemic risk city", *Parasites and Vectors*, Vol. 7, pp. 320.

Gorrochotegui-Escalante, N. et al. (2005), "Association mapping of segregating sites in the early trypsin gene and susceptibility to dengue-2 virus in the mosquito *Aedes aegypti*", *Insect Biochemistry and Molecular Biology*, Vol. 35, No. 7, pp. 771-788.

Gorrochotegui-Escalante, N. et al. (2002), "Breeding structure of *Aedes aegypti* populations in Mexico varies by region", *American Journal of Tropical Medicine and Hygiene*, Vol. 66, pp. 213–222.

Gorrochotegui-Escalante, N. et al. (2000), "Genetic isolation by distance among *Aedes aegypti* populations along the northeastern coast of Mexico", *The American Journal of Tropical Medicine and Hygiene*, Vol. 62, No. 2, pp. 200-209.

Grieco, J.P. et al. (2007), "A new classification system for the actions of IRS chemicals traditionally used for malaria control", *PLoS ONE*, Vol. 2, No. 8: e716.

Grieco, J.P. et al. (2000), "A comparison study of house entering and exiting behavior of *Anopheles vestitipennis* (Diptera: Culicidae) using experimental huts sprayed with DDT or deltamethrin in the southern district of Toledo, Belize, C.A.", *Journal of Vector Ecology*, Vol. 25, pp. 62–73.

Gubler, D.J. et al. (1979), "Variation in susceptibility to oral infection with dengue viruses among geographic strains of *Aedes aegypti*", *The American Journal of Tropical Medicine and Hygiene*, Vol. 28, No. 6, pp. 1045-1052.

Guillemaud, T. et al. (1997), "Esterase gene amplification in *Culex pipiens*", *Insect Molecular Biology*, Vol. 6, pp. 319-327.

Guo, X.X. et al. (2013), "Vector competence of *Aedes albopictus* and *Aedes aegypti* (Diptera: Culicidae) for DEN2-43 and New Guinea C virus strains of dengue 2 virus", *Acta Tropica*, Vol. 128, No. 3, pp. 566-570.

Harris, A.F., S. Rajatileka and H. Ranson (2010), "Pyrethroid resistance in *Aedes aegypti* from Grand Cayman", *The American Journal of Tropical Medicine and Hygiene*, Vol. 83, pp. 277-284.

Hawkes, N.J. and J. Hemingway (2002), "Analysis of the promoters for the beta-esterase genes associated with insecticide resistance in the mosquito *Culex quinquefasciatus*", *Biochimica et Biophysica Acta (BBA)*, Vol. 1574, pp. 51-62.

Hemingway, J. and H. Ranson (2000), "Insecticide resistance in insect vectors of human disease", *Annual Review of Entomology*, Vol. 45, pp. 371-391.

Hemingway, J. et al. (2004), "The molecular basis of insecticide resistance in mosquitoes", *Insect Biochemistry and Molecular Biology*, Vol. 34, pp. 653-665.

Herrera, F. et al. (2006), "Population genetic structure of the dengue mosquito *Aedes aegypti* in Venezuela", *Memórias do Instituto Oswaldo Cruz*, Vol. 101, No. 6, pp. 625-633.

Hlaing, T. et al. (2009), "Mitochondrial pseudogenes in the nuclear genome of *Aedes aegypti* mosquitoes: Implications for past and future population genetic studies", *BMC Genetics*, Vol. 10, pp. 11.

Horstick, O. et al. (2010), "Dengue vector-control services: how do they work? A systematic literature review and country case studies", *Transactions of the Royal Society of Tropical Medicine and Hygiene*, Vol. 104, pp. 379-386.

Huber, K. et al. (2008), "*Aedes aegypti* in Senegal: Genetic diversity and genetic structure of domestic and sylvatic populations", *The American Journal of Tropical Medicine and Hygiene*, Vol. 79, No. 2, pp. 218-229.

Khambay, B.P.S. (2002), "Pyrethroid insecticides", *Pesticide Outlook*, Vol. 13, pp. 49-54.

Kumar, N.P. et al. (2012), "Detection of Chikungunya virus in wild populations of *Aedes albopictus* in Kerala State, India", *Vector-Borne and Zoonotic Diseases*, Vol. 12, No. 10, pp. 907-911.

Lambrechts, L. (2011), "Quantitative genetics of *Aedes aegypti* vector competence for dengue viruses: Towards a new paradigm?", *Trends in Parasitology*, Vol. 27, No. 3, pp. 111-114.

Lambrechts, L. et al. (2011), "Impact of daily temperature fluctuations on dengue virus transmission by *Aedes aegypti*", *Proceedings of the National Academy of Sciences of the United States of America*, Vol. 108, No. 18, pp. 7460-7465.

Lambrechts, L. et al. (2009), "Genetic specificity and potential for local adaptation between dengue viruses and mosquito vectors", *BMC Evolutionary Biology*, Vol. 9, pp. 160.

Li, M.I. et al. (2012), "Oral susceptibility of Singapore *Aedes* (Stegomyia) *aegypti* (Linnaeus) to Zika virus", *PLOS Neglected Tropical Diseases*, Vol. 6, No. 8: e1792.

Lima, E.P. et al. (2011), "Insecticide resistance in *Aedes aegypti* populations from Ceara, Brazil", *Parasites and Vectors*, Vol. 4, pp. 5.

Lima, R.S. Jr and V.M. Scarpassa (2009), "Evidence of two lineages of the dengue vector *Aedes aegypti* in the Brazilian Amazon, based on mitochondrial DNA ND4 gene sequences", *Genetics and Molecular Biology*, Vol. 32, No. 2, pp. 414-422.

Liu, N. et al. (2006), "Pyrethroid resistance in mosquitoes", *Insect Science*, Vol. 13, pp. 159-166.

Lozano-Fuentes, S. et al. (2012), "The dengue virus mosquito vector *Aedes aegypti* at high elevation in Mexico", *The American Journal of Tropical Medicine and Hygiene*, Vol. 87, No. 5, pp. 902-909.

Lozano-Fuentes, S. et al. (2009), "The neovolcanic axis is a barrier to gene flow among *Aedes aegypti* populations in Mexico that differ in vector competence for Dengue 2 virus", *PLoS Neglected Tropical Diseases*, Vol. 3, No. 6: e468.

Lumjuan, N. et al. (2007), "The *Aedes aegypti* glutathione transferase family", *Insect Biochemistry and Molecular Biology*, Vol. 37, pp. 1026-1035.

Madhu, S.K. and V.A. Vijayan (2009), "Laboratory evaluation of a juvenile hormone mimic, pyriproxyfen on *Culex quinquefasciatus* Say and *Aedes aegypti* Linn. at Mysore, India", *The Journal of Communicable Diseases*, Vol. 41, No. 3, pp. 169-174.

Maestre-Serrano, R. et al. (2014), "Susceptibility to insecticides and resistance mechanisms in *Aedes aegypti* from the Colombian Caribbean Region", *Pesticide Biochemistry and Physiology*, No. 116, pp. 63-73.

Malaithong, N. et al. (2010), "Human landing patterns of *Anopheles* dirus sensu lato (Diptera: Culicidae) in experimental huts treated with DDT or deltamethrin", *Journal of Medical Entomology*, Vol. 47, pp. 823-832.

Marcombe, S. et al. (2013), "Insecticide-driven patterns of genetic variation in the dengue vector *Aedes aegypti* in Martinique Island", *PLoS One*, Vol. 8: e77857.

Marcombe, S. et al. (2012), "Insecticide resistance in the dengue vector *Aedes aegypti* from Martinique: Distribution, mechanisms and relations with environmental factors", *PLoS One*, Vol. 7: e30989.

Marcombe, S. et al. (2009), "Exploring the molecular basis of insecticide resistance in the dengue vector *Aedes aegypti*: A case study in Martinique Island (French West Indies)", *BMC Genomics*, Vol. 10, pp. 494.

Martins, A.J. et al. (2009a), "Frequency of Val1016Ile mutation in the voltage-gated sodium channel gene of *Aedes aegypti* Brazilian populations", *Tropical Medicine & International Health*, Vol. 14, pp. 1351-1355.

Martins, A.J. et al. (2009b), "Voltage-gated sodium channel polymorphism and metabolic resistance in pyrethroid-resistant *Aedes aegypti* from Brazil", *The American Journal of Tropical Medicine and Hygiene*, Vol. 81, pp. 108-115.

McAllister, J.C., M.S. Godsey and M.L. Scott (2012), "Pyrethroid resistance in *Aedes aegypti* and *Aedes albopictus* from Port-au-Prince, Haiti", *Journal of Vector Ecology*, Vol. 37, pp. 325-332.

McClelland, G.A.H. (1962), "Sex-linkage in *Aedes aegypti*", *Transaction of the Royal Society of Tropical Medicine and Hygine*, Vol. 56, No. 4.

McDonald, P.T. and K.S. Rai (1970), "Correlation of linkage groups with chromosomes in the mosquito, *Aedes aegypti*", *Genetics*, Vol. 66, pp. 475–485.

Mercado-Curiel, R.F., W.C. Black 4th and M.deL. Muñoz (2008), "A dengue receptor as possible genetic marker of vector competence in *Aedes aegypti*", *BMC Microbiology*, Vol. 8, pp. 118.

Miller, B.R. and C.J. Mitchell (1991), "Genetic selection of a flavivirus-refractory strain of the yellow fever mosquito *Aedes aegypti*", *The American Journal of Tropical Medicine and Hygiene*, Vol. 45, No. 4, pp. 399-407.

Moncayo, A.C. et al. (2004), "Dengue emergence and adaptation to peridomestic mosquitoes", *Emerging Infectious Diseases*, Vol. 10, No. 10, pp. 1790-1796.

Moore, M. et al. (2013), "Dual African origins of global *Aedes aegypti* s.l. populations revealed by mitochondrial DNA", *PLoS Neglected Tropical Diseases*, Vol. 7, No. 4: e2175.

Mousson, L. et al. (2005), "Phylogeography of *Aedes (Stegomyia) aegypti* (L.) and *Aedes (Stegomyia) albopictus* (Skuse) (Diptera: Culicidae) based on mitochondrial DNA variations", *Genetic Research* , Vol. 86, pp. 1–11.

Muñoz, M.deL. et al. (2013a), "Gene flow pattern among *Aedes aegypti* populations in Mexico", *Journal of the American Mosquito Control Association*, Vol. 29, No. 1, pp. 1-18.

Muñoz, M.deL. et al. (2013b), "Proteomic identification of dengue virus binding proteins in *Aedes aegypti* mosquitoes and *Aedes albopictus* cells", *Biomed Res Int*, Article ID: 875958.

Munstermann, L.E. and G.B. Craig Jr. (1979), "Genetics of *Aedes aegypti*: Updating the linkage map", *Journal of Heredity*, Vol. 70, pp. 291-296.

Nauen, R. (2006), "Insecticide mode of action: Return of the ryanodine receptor", *Pest Management Science*, Vol. 62, pp. 690-692.

Nauen, R. and T. Bretschneider (2002), "New modes of action of insecticides", *Pesticide Outlook*, Vol. 13, pp. 241-245.

Navarro, J.C. et al. (2010), "Highest mosquito records (Diptera: Culicidae) in Venezuela", *Revista de Biología Tropical*, Vol. 58, No. 1, pp. 245-1254.

Nene, V. et al. (2007), "Genome sequence of *Aedes aegypti*, a major arbovirus vector", *Science*, Vol. 316, No. 5832, pp. 1718-1723.

Oakeshott, J.G. et al. (2005), "Biochemical genetics and genomics of insect esterases", in L.I. Gilbert, K. Latrou and S.S. Gill (eds.), *Comprehensive Molecular Insect Science Pharmacology*, Vol. 5, pp. 309-381.

Paduan, K.S. and P.E. Ribolla (2008), "Mitochondrial DNA polymorphism and heteroplasmy in populations of *Aedes aegypti* in Brazil", *Journal of Medical Entomology*, Vol. 45, No. 1, pp. 59-67.

PAHO (1994), *Dengue and Dengue Hemorrhagic Fever in the Americas: Guidelines for Prevention and Control*, PAHO Scientific Publication 548, Pan American Health Organization, Washington DC.

Paupy, C. et al. (2012), "Genetic structure and phylogeography of *Aedes aegypti*, the dengue and yellow-fever mosquito vector in Bolivia", *Infection, Genetics and Evolution*, Vol. 12, No. 6, pp. 1260-1269.

Paupy, C. et al. (2008), "Gene flow between domestic and sylvan populations of *Aedes aegypti* (Diptera: Culicidae) in North Cameroon", *Journal of Medical Entomology*, Vol. 45, No. 3, pp. 391-400.

Paupy, C. et al. (2004), "Comparisons of amplified fragment length polymorphism (AFLP), microsatellite, and isoenzyme markers: Population genetics of *Aedes aegypti* (Diptera: Culicidae) from Phnom Penh (Cambodia)", *Journal of Medical Entomology*, Vol. 41, No. 4, pp. 664-671.

Polsomboon, S. et al. (2008), "Biting patterns of *Anopheles minimus complex* (Diptera: Culicidae) in experimental huts treated with DDT and deltamethrin", *Journal of Vector Ecology*, Vol. 33, pp. 285-292.

Ponce-Garcia, G. et al. (2009), "Recent rapid rise of a permethrin knock down resistance allele in *Aedes aegypti* in Mexico", *PLoS Neglected Tropical Diseases*, Vol. 3: e531.

Powell, J.R. and W.J. Tabachnick (2013), "History of domestication and spread of *Aedes aegypti* – A review", *Memórias do Instituto Oswaldo Cruz*, Vol. 108, pp. 11-17.

Puinean, A.M. et al. (2010), "Amplification of a cytochrome P450 gene is associated with resistance to neonicotinoid insecticides in the aphid *Myzus persicae*", *PLoS Genet*, Vol. 6: e1000999.

Rai, K.S. (1963), "A comparative study of mosquito karyotypes", *Annals of the Entomological Society of America*, Vol. 56, pp. 160-170.

Ranson, H. and J. Hemingway (2005), "Mosquito glutathione transferases", *Methods in Enzymology*, Vol. 401, pp. 226-241.

Ranson, H. et al. (2002), "Evolution of supergene families associated with insecticide resistance", *Science*, Vol. 298, pp. 179-181.

Reiter, P. and D.J. Gubler (1997), "Surveillance and control of urban dengue vectors", in D.J. Gubler and G. Kuno (eds.), *Dengue and Dengue Hemorrhagic Fever*, CABI Publishing, Cambridge, pp. 425-462.

Ribeiro, J.M. et al. (2007), "An annotated catalogue of salivary gland transcripts in the adult female mosquito, *Aedes aegypti*", *BMC Genomics*, Vol. 8, No. 6.

Richard, V., T. Paoaafaite and V-M. Cao-Lormeau (2016), "Vector competence of French Polynesian *Aedes aegypti* and *Aedes polynesiensis* for Zika virus", *PLoS Neglected Tropical Diseases*, Vol. 10, No. 9: e0005024.

Rinkevich, F.D., Y.Z. Du and K. Dong (2013), "Diversity and convergence of sodium channel mutations involved in resistance to pyrethroids", *Pesticide Biochemistry and Physiology*, Vol. 106, pp. 93-100.

Roberts, D.R. and W.D. Alecrim (1991), "Behavioral response of *Anopheles darlingi* to DDT-sprayed house walls in Amazonia", *Bulletin of the Pan American Health Organization*, Vol. 25, pp. 210-217.

Rosen, L. et al. (1985), "Comparative susceptibility of mosquito species and strains to oral and parenteral infection with dengue and Japanese encephalitis viruses", *The American Journal of Tropical Medicine and Hygiene*, Vol. 34, No. 3, pp. 603-615.

Rozendaal, J.A. et al. (1989), "Behavioral responses of *Anopheles darlingi* in Suriname to DDT residues on house walls", *Journal of the American Mosquito Control Association*, Vol. 5, pp. 339-350.

Saavedra-Rodriguez, K. et al. (2007), "A mutation in the voltage-gated sodium channel gene associated with pyrethroid resistance in Latin American *Aedes aegypti*", *Insect Molecular Biology*, Vol. 16, No. 6, pp. 785-798.

Saito, K. and M.C. Siomi (2010), "Small RNA-mediated quiescence of transposable elements in animals", *Developmental Cell*, Vol. 19, No. 5, pp. 687-697.

Senti, K.A. and J. Brennecke (2010), "The piRNA pathway: A fly's perspective on the guardian of the genome", *Trends in Genetics*, Vol. 26, No. 12, pp. 499-509.

Service, M. (2012), *Medical Entomology for Students, 5th Ed*., Cambridge University Press, New York, pp. 303.

Severson, D.W. and S.K. Behura (2012), "Mosquito genomics: Progress and challenges", *Annual Review of Entomology*, Vol. 57, pp. 143-166.

Severson, D.W. et al. (2004), "Comparative genome analysis of the yellow fever mosquito *Aedes aegypti* with *Drosophila melanogaster* and the malaria vector mosquito *Anopheles gambiae*", *Journal of Heredity*, Vol. 95, No. 2, pp. 103-113.

Severson, D.W. et al. (2002), "Linkage map organization of expressed sequence tags and sequence tagged sites in the mosquito, *Aedes aegypti*", *Insect Molecular Biology*., Vol. 11, No. 4, pp. 371-378.

Severson, D.W. et al. (1995), "Restriction fragment length polymorphism mapping of quantitative trait loci for malaria parasite susceptibility in the mosquito *Aedes aegypti*", *Genetics*, Vol. 139, No. 4, pp. 1711-1717S.

Severson, D.W. et al. (1994), "Chromosomal mapping of two loci affecting filarial worm susceptibility in *Aedes aegypti*", *Insect Molecular Biology*, Vol. 3, No. 2, pp. 67-72.

Severson, D.W. et al. (1993), "Linkage map for *Aedes aegypti* using restriction fragment length polymorphisms," *Journal of Heredity*, Vol. 84, No. 4, pp. 241-247.

Sharakhova, M.V. et al. (2011), "Arm-specific dynamics of chromosome evolution in malaria mosquitoes", *BMC Evolutionary Biology*, Vol. 11, pp. 91.

Sharma, G.P. et al. (1978), "A preliminary map of the salivary gland chromosomes of *Aedes* (Stegomyia) *aegypti* (Culicidae, Diptera)", *Cytobios*, Vol. 22, pp. 169-178.

Sieglaff, D.H., K.A. Duncan and M.R. Brown (2005), "Expression of genes encoding proteins involved in ecdysteroidogenesis in the female mosquito, *Aedes aegypti*", *Insect Biochemistry and Molecular Biology*, Vol. 35, pp. 471-490.

Siller, Q. et al. (2011), "Update on the frequency of Ile1016 mutation in voltage-gated sodium channel gene of *Aedes aegypti* in Mexico", *Journal of the American Mosquito Control Association*, Vol. 27, pp. 357-362.

Smith, A. (1965), "A verandah-trap hut for studying the house-frequenting habits of mosquitos and for assessing insecticides. 2. The effect of dichlorvos (DDVP) on egress and mortality of *Anopheles gambiae* Giles and *Mansonia uniformis* (Theo.) entering naturally", *Bulletin of Entomological Research*, Vol. 56, pp. 275-282.

Soderlund, D.M. and D.C. Knipple (2003), "The molecular biology of knockdown resistance to pyrethroid insecticides", *Insect Biochemistry and Molecular Biology*, Vol. 33, pp. 563-577.

Sylla, M. et al. (2009), "Gene flow, subspecies composition, and dengue virus-2 susceptibility among *Aedes aegypti* collections in Senegal", *PLoS Neglected Tropical Diseases*, Vol. 3, No. 4: e408.

Tabachnick, W.J. and J.R. Powell (1979), "A world-wide survey of genetic-variation in the yellow-fever mosquito, *Aedes aegypti*", *Genetical Research*, Vol. 34, No. 3, pp. 215-229.

Tabachnick, W.J. et al. (1985), "Oral infection of *Aedes aegypti* with yellow-fever virus – Geographic variation and genetic considerations", *American Journal of Tropical Medicine and Hygiene*, Vol. 34, No. 6, pp. 1219-1224.

Tardieux, I. et al. (1990), "Variation among strains of *Aedes aegypti* in susceptibility to oral infection with dengue virus type 2", *The American Journal of Tropical Medicine and Hygiene*, Vol. 43, No. 3, pp. 308-313.

Timoshevskiy, V.A. et al. (2014), "Genomic composition and evolution of *Aedes aegypti* chromosomes revealed by the analysis of physically mapped supercontigs", *BMC Biology*, Vol. 12, pp. 27.

Timoshevskiy, V.A. et al. (2013), "An integrated linkage, chromosome, and genome map for the yellow fever mosquito *Aedes aegypti*", *PLoS Neglected Tropical Diseases*, Vol. 7, No. 2, pp. 2052.

Urdaneta-Marquez, L. et al. (2008), "Genetic relationships among *Aedes aegypti* collections in Venezuela as determined by mitochondrial DNA variation and nuclear single nucleotide polymorphisms", *The American Journal of Tropical Medicine and Hygiene*, Vol. 78, No. 3, pp. 479-491.

Valles, S.M., K. Dong and R.J. Brenner (2000), "Mechanisms responsible for cypermethrin resistance in a strain of German cockroach, Blattella germanica", *Pesticide Biochemistry and Physiology*, Vol. 66, pp. 195-205.

Vazeille-Falcoz, M. et al. (1999), "Variation in oral susceptibility to dengue type 2 virus of populations of *Aedes aegypti* from the islands of Tahiti and Moorea, French Polynesia", *The American Journal of Tropical Medicine and Hygiene*, Vol. 60, No. 2, pp. 292-299.

Vega-Rúa, A. et al. (2014), "High level of vector competence of *Aedes aegypti* and *Aedes albopictus* from ten American countries as a crucial factor in the spread of Chikungunya virus", *Journal of Virology*, Vol. 88, No. 11, pp. 6294-6306.

Vontas, J. et al. (2012), "Insecticide resistance in the major dengue vectors *Aedes albopictus* and *Aedes aegypti*", *Pesticide Biochemistry and Physiology*, Vol. 104, pp. 126-131.

Watts, D.M. et al. (1987), "Effect of temperature on the vector efficiency of *Aedes aegypti* for dengue 2 virus", *The American Journal of Tropical Medicine and Hygiene*, Vol. 36, No. 1, pp. 143-152.

Weill, M. et al. (2003), "Comparative genomics: Insecticide resistance in mosquito vectors", *Nature*, Vol. 423, pp. 136-137.

WHO (2012a), *WHO Recommended Insecticides for Space Spraying Against Mosquitoes*, World Health Organization, Geneva, www.who.int/whopes/Insecticides_for_space_spraying_Jul_2012.pdf.

WHO (2012b), *Global Plan for Insecticide Resistance Management in Malaria Vectors (GPIRM)*, World Health Organization, Geneva.

WHO (2009), *WHO Recommended Insecticides for Indoor Residual Spraying Against Malaria Vectors*, World Health Organization, Geneva, www.who.int/whopes/Insecticides_IRS_Malaria_09.pdf.

WHOPES (2005), *Guidelines for Laboratory and Field Testing of Mosquito Larvicides*, World Health Organization Pesticide Evaluation Scheme, Geneva.

Zhong, D. et al. (2006), "Amplified fragment length polymorphism mapping of quantitative trait loci for malaria parasite susceptibility in the yellow fever mosquito *Aedes aegypti*", *Genetics*, Vol. 173, No. 3, pp. 1337-1345.

Chapter 4. **Ecology of the mosquito *Ae. aegypti***

This chapter considers the ecology of Aedes aegypti. *Elements are provided on niche modelling and the species distribution modelling. The limited trophic interactions of this mosquito and its development in anthropic habitats and urban environments are detailed, followed with considerations on its abiotic requirements and tolerance in aquatic and terrestrial conditions. Information is also provided on the* Ae. Aegypti *biotic interactions in the landscape, its life history traits and fitness influencing dispersal patterns and population density and distribution. Elements are given on population models being developed (spatial-temporal dynamics).*

Ecological niche/species distribution modelling of *Ae. aegypti*

The ecological niche of a species can be defined as the range of environmental and biotic conditions within which its populations can persist without immigration (Hutchinson, 1957). The range of environmental and biotic conditions can be assessed through niche modelling, providing evidence for geographic isolation between populations (either based on conserved or divergent ecological niches). By mapping the spatial distribution of environmental suitability of climatic variables (Raxworthy et al., 2007), the niche modelling provides a much stronger case for geographic isolation for populations isolated by intervening unsuitable regions reducing gene flow.

The population structure of *Ae. aegypti* is complex, varies by region and scale, and can be influenced by environment and geography (Yan, Chadee and Severson, 1998; Urdaneta-Marquez et al., 2008). Urban estimates of genetic differentiation have varied in part due to environmental conditions and dispersal patterns (Huber et al., 2002; da Costa-Ribeiro, Lourenço-de-Oliveira and Failloux, 2006). *Ae. aegypti* population dynamics in urban areas are subject to daily as well as seasonal meteorological variability (Halstead, 2008). Effects of seasonal climatic factors on mosquito life-history traits are well documented, particularly on adult distribution, survival and availability of oviposition sites. Several supportive studies have also been made on physiologic aspects such as decreased embryonic (e.g. Trpis, Haufe and Shemanchuk, 1973) and larval (e.g. Teng and Apperson, 2000) development times as well as decreased size of adults (e.g. Rueda et al., 1990) being associated with higher temperature.

It may be an oversimplified assumption that climate change will independently lead to an increased range for this species and a concomitant expansion of the risk of dengue infections around the world. A range of dynamic factors must be considered when predicting future global distribution trends. Constraining the focus of models to a local and/or regional scale rather than aspiring for global models may increase their predictive capacity. In light of climate change, the major drivers of future dengue susceptible areas will likely include unprecedented population growth, particularly in urban areas in the tropics; an increase in the movement of both vector and virus reservoirs via modern transport; and a lack of effective mosquito management (Mackenzie, Gubler and Petersen, 2004). More details are given in the “Abiotic requirements and tolerance” section below.

Ae. aegypti niche and trophic interactions

The mosquito *Ae. aegypti* has a relatively narrow niche with limited trophic interactions. The anthropophilic form of *Ae. aegypti*, *Ae. aegypti aegypti*, utilises flooded artificial containers as habitat for larvae and pupae. It is this form that has become established in most tropical and subtropical areas globally and is the primary vector of dengue, Zika and several other viruses. The original sylvan form of *Ae. aegypti*, *Ae. aegypti formosus*, occurs in sub-Saharan Africa where natural containers such as flooded tree holes are the dominant larval habitat (Lounibos, 1981). A plethora of man-made objects composed of plastic, rubber, metal, concrete, masonry and ceramics have been shown to hold water, capture nutrients and produce *Ae. aegypti* (Ritchie, 2014). Within these flooded containers, larvae graze on the surface of the container, feeding on fallen detritus (typically leaves) and bacteria and algae that have grown on it. Protein sources such as insects, seeds, fruit and even dead conspecific mosquito larvae are fed upon. However, many of these containers are nutrient poor, especially covered containers such as water storage tanks, and typically produce stunted adults. The restriction of nutrients, coupled

with high larval populations beyond the carrying capacity of the container, reduce larval growth and pupation via density-dependent regulation (Hancock et al., 2016). Indeed, it is the sudden input of protein from, for example, a cricket that falls into and drowns within the container, which can lead to a surge in larval growth, pupation and adult emergence.

There is a very limited number of species known to feed upon *Ae. aegypti* larvae and pupae. Most artificial containers are small to medium in size, only intermittently flooded and thus do not maintain populations of predaceous aquatic insects or vertebrates such as fish and amphibians. While many of these aquatic predators can eat mosquito larvae, they are uncommon in most *Ae. aegypti* habitat. In some larger containers, dytiscid beetles and dragonfly naiads can occur and feed upon mosquito larvae, while fish and tadpoles have been propagated and released in large water storage containers to control *Ae. Aegypti*. Among the many kinds of mosquito that do not consume blood, mosquitoes of the genus *Toxorhynchites* oviposit in artificial containers and selectively feed upon mosquito larvae (Trpis, 1973) being *Ae. aegypti* larvae as well as from other container mosquito species such as *Ae. albopictus* and *Ae. notoscriptus*. Copepods of the genus *Mesocyclops* will actively predate first instar larvae of *Ae. aegypti*.

Adult *Ae. aegypti* are also restricted to largely "artificial habitats" created by man. This "cockroach of mosquitoes", as it is often called, prefers to harbour inside buildings and houses in urban areas where it has ready access to humans for blood feeding. In some instances, all life stages of *Ae. aegypti* (egg, larvae, pupae and adult) can occur inside, especially in areas where water is stored indoors for domestic use. However, in many areas *Ae. aegypti* adults do spend considerable time outdoors where they seek flooded containers in which to oviposit. Predation of adult *Ae. aegypti* is poorly studied. Spiders, especially saltidae (jumping spiders), are known to actively stalk and feed upon adult mosquitoes indoors (Sulaiman et al., 1990) and can be a major predator in semi-field cages (S. Ritchie personal observation). Most other animals purportedly linked to adult mosquito predation, such as bats, geckoes and dragonflies, often feed either crepuscularly or at night, and would likely miss day active *Ae. aegypti*. Ants and cockroaches are known to feed upon *Ae. aegypti* eggs in containers (see below the section on "Biotic interactions in the landscape"), and mites and booklice often predate eggs in laboratory colonies and thus potentially would in the field. Ants readily consume dead adult mosquitoes on the ground and even stranded larvae in recently dried containers. As *Ae. aegypti* occurs in relatively low numbers (generally < 10 adults per house), the biomass of this mosquito is small (an estimated 2 g/ha in Cairns, Queensland, Australia [S. Ritchie, unpublished data]) and it is usually considered that it does not make a large trophic contribution.

In summary, urbanised *Ae. aegypti* (*Ae. aegypti aegypti*) is largely restricted to artificial, man-made habitats in geographic areas outside of its native range. Endemic species within "natural" tropical ecosystems are not trophically connected with *Ae. aegypti aegypti*, or in a limited way. Thus, it is assumed that they are at minimal risk should the species be eliminated from those areas.

Anthropic habitats

Increase in the size and population density of major cities place increasing demands on infrastructure and essential services, particularly in developing countries. The response to these demands may dramatically alter the suitability of a locality for urban mosquito breeding. An absence or irregularity of water supply will lead to an increase in domestic

water storage practices which, in turn, will alter the landscape of potential *Ae. aegypti* habitat, perhaps providing a far more regular or abundant supply of larval sites.

The effects of topographic features of urban environments on *Ae. aegypti* behaviour are not fully understood; however, Reiter et al. (1995) noted that buildings were not an impediment to *Ae. aegypti* flight. Certain results indicate that urban landscape does contain barriers to dispersal (Reiter et al., 1995; Chadee, 2004; Valerio et al., 2012), and this affects the mosquito population structure.

Such information can be useful to agencies in charge of vector control for better targeting mosquito populations and areas of higher risk within control zones. Understanding the role of landscape features on population dispersal is likely critical to achieving success with any *Ae. aegypti* control strategy (more information is given in Annex A. Control of the mosquito *Ae. aegypti*).

Abiotic requirements and tolerance

Considerable variation in adult size occurs as a result of habitat conditions such as water quality, food availability, and crowding during mosquito larval breeding (Nasci, 1991). The adult size strongly influences various aspects of mosquito life history: survivorship (Pumpuni and Walker, 1989), mating success (Yuval, Wekesa and Washino, 1993), blood meal size (Xue, Edman and Scott, 1995), parous rate (Haramis, 1983), fecundity (Packer and Corbet, 1989), dispersal (Renshaw, Service and Birley, 1994) and longevity (Feinson and Spielman, 1980). Among abiotic and biotic factors, high temperature and low nutrition in the developing stages of mosquitoes generally result in small adults. While temperature, humidity and rainfall have overt impacts on mosquito adult survival and ecology, other climatic factors such as photoperiod and wind velocity may also be influential. Importantly, it is necessary to consider that these meteorological conditions have a combined effect on the survival and development of mosquitoes and that it is difficult to examine the potential impact of these factors independently as a consequence (Jansen and Beebe, 2010).

Aquatic

Ae. aegypti prefers clean water found in many types of domestic containers inside or near human dwellings (Nazri et al., 2013). The *Aedes* mosquito larvae require standing water to complete their growth cycle, therefore, any body of standing water represents a potential *Aedes* mosquito breeding site for mosquito larvae to mature. Water quality affects the productivity of a potential mosquito breeding habitat. Typically, greater numbers of mosquitoes are produced in water bodies with poor circulation, higher temperatures and higher organic content than in water bodies having good circulation, lower temperatures and lower organic content (Focks et al., 1993; Murrell and Steven, 2008).

Aquatic habitats for *Ae. aegypti* are containers in which eggs develop into adult mosquitoes. Mosquitoes lay eggs on the walls of water-filled containers in or around the house. The eggs hatch when submerged in water and can survive desiccation for months (see section on Morphology in Chapter 1). There is a great variety of man-made containers on backyards or patios that collect rainwater or that are filled with water by people. Artificial or natural water containers (water storage containers, flower pots, discarded tires, plates under potted plants, cemetery vases, flower pots, buckets, tin cans, clogged rain gutters, ornamental fountains, drums, water bowls for pets, birdbaths, etc.)

that are within or close to places where humans live are ideal larval habitats for this mosquito.

Terrestrial

Studies of associations between climate parameters and *Ae. aegypti* are complicated by the dependence of the mosquito on humans, especially its preference for human blood and its adaptation to use artificial containers as larval development sites (Focks and Alexander, 2006; Tun-Lin et al., 2009).

Ae. aegypti is the major urban vector of DENV worldwide. Over the last 25 years, there has been a global increase in both the distribution of *Ae. aegypti* and epidemic DENV activity (Mackenzie, Gubler and Petersen, 2004). Historically, *Ae. aegypti* has been thought to be able to establish in regions between the northern January and southern July 10°C isotherms, while more recent studies suggest that the 15°C yearly isotherm is a better estimate (see Chapter 1 section on "Origin and current geographic distribution").

Although *Ae. aegypti* is generally considered a tropical mosquito (Christophers, 1960), it should be noted that its distribution in some temperate regions of the world does appear to be influenced by climate variables (Liu-Helmersson et al., 2016).

The potential effects of climate and environmental change on *Ae. aegypti* and DENV transmission have generated much debate (Jetten and Focks, 1997; Patz et al., 1998; Hales et al., 2002; Barclay, 2008; Beebe et al., 2009; Ooi and Gubler et al., 2009; Banu et al., 2011; Brady et al., 2013, 2014). Part of this controversy relates to modelling future climate-driven change for the vector or disease without accounting for human-related factors, which also impact the vector itself (e.g. availability of water-filled artificial containers as larval development sites) or DENV transmission dynamics (e.g. serotype-specific susceptibility of the human population). Several reports consider that the domestic nature of this species probably exerts more influence on its distribution than climate variables. These confounding factors can, thus, modulate the effects of climate change on the mosquito distribution. It is also recognised that the effects of climate and environmental change are location-specific and likely to impact *Ae. aegypti* and, potentially, also DENV transmission to a greater extent in some geographic areas than others (Lozano-Fuentes et al., 2012). Studies in Australia suggest that future changes in *Ae. aegypti* distribution in the country may not be directly caused by climate change but rather, by human response to changing rainfall patterns by increased or decreased use of water storage containers (Beebe et al., 2009; Russell et al., 2009; Williams et al., 2010, 2014, 2015; Bannister-Tyrrell et al., 2013).

Biotic interactions in the landscape

Biological interactions between species occupying similar niches may also influence the distribution and abundance of *Ae. aegypti*. Whilst a number of underlying processes including interspecific larval resource competition has been suggested (Lounibos et al., 2002; Juliano and Lounibos, 2005), it is most likely that multiple factors determine the current distributions of each species. Examples of these interconnected factors include the potentially asymmetrical effects of abiotic factors (including climate) on different life cycle stages as underlined above, apparent competition induced by parasites, mating interference and variation between the microclimates in given locations (Lounibos et al., 2002; Juliano and Lounibos, 2005).

In the aquatic environment, the larvae have a number of predators including other invertebrates, tadpoles and fish. Aquatic invertebrate predators from the Coleoptera (beetles), Diptera (flies including the predaceous mosquito *Toxorhynchites* spp.), Hemiptera (true bugs) and Odonata (dragonflies and damselflies) orders prey on all mosquito larvae in the same environment (Shaalan and Canyon, 2009). Because *Ae. aegypti* usually uses man-made containers as breeding sites, it does not seem to have specific predators but rather "opportunistic" ones that feed on larvae if encountering them, as detailed under a previous section dealing with trophic interactions. Predators can significantly affect the survival, development, and recruitment levels of mosquitoes in their aquatic breeding sites. There is also some evidence that the occasional presence of predators in vessels can favour oviposition by *Ae. aegypti*, the mosquitoes being attracted to predator kairomones[1] (Albeny-Simões et al., 2014). Mogi (2007), however, reviewed mosquito invertebrate predators and concluded that they are usually absent or sparse in man-made containers in residential areas.

Russell, Kay and Shipton (2001) placed filter-paper strips containing *Ae. aegypti* eggs within flooded telecommunication pits and surface containers in Charters Towers (Australia), and found that no subterranean eggs and only 1% of surface-placed eggs, respectively, survived the 4-month dry season despite the egg capacity to survive desiccation for months (see Chapter 2, section on Life cycle). In this case, predation was primarily by cockroaches. Attack by a fungus (*Penicillium citrinum*) also resulted in high mortality within the flooded subterranean site. The high mortality of eggs in subterranean sites led the authors to conclude that subterranean egg refugia were not responsible for the reintroduction of *Ae. aegypti* into surface containers at the onset of the wet season.

Ants are also a significant predator of *Ae. aegypti* eggs in colonies, and probably also in the field (Focks et al., 1993; Russell, Kay and Shipton, 2001; Ritchie, 2014).

Life history traits and fitness

The body size of mosquitoes can influence a number of bionomic factors, such as their blood-feeding ability, host attack rate and fecundity (Klowden and Lea, 1978; Xue, Edman and Scott, 1995; Farjana and Tuno, 2012). All of these traits are important determinants of their potential to transmit diseases (Farjana and Tuno, 2013).

Ae. aegypti, the container-breeding mosquito, is closely associated with humans and highly anthropophilic, tending to predominate in densely populated urban areas. They are commonly found indoors, breeding in artificial containers, with female needing to feed on blood to produce eggs, as described above. Studies have demonstrated high anthropophily, with over 90% of the ingested blood being human, and the rest from pets, such as dogs and cats (Scott et al., 1993). Multiple feeding in a gonotrophic cycle can increase the risk of disease transmission by increasing the frequency of contact with hosts (Garrett-Jones, 1964; Garrett-Jones and Shidrawi, 1969; Dye, 1986). Two types of multiple feeding have been recognised: supplementary feeding owing to nutritional reserve depletion in teneral females (Scott et al., 1993; Xue, Edman and Scott, 1995; Scott et al., 2000; Reyes-Villanueva, 2004) and interrupted feeding owing mainly to host defence (Clements, 1999). For more detailed information, see Chapter 2 section on "Physiology of reproduction".

Dispersal

Landscape fragmentation and human demography can influence dispersal patterns of mosquitoes. The degree and nature of modification can affect the flow of genes

conditioning vector competence and insecticide resistance (Hemme et al., 2010). Generally anthropic habitats minimise climatic variation where *Ae. aegypti* distribution is dependent on human behaviour (Jansen and Beebe, 2010).

Mosquito dispersal patterns are non-random and influenced by environmental factors as reported by Sheppard et al. (1969) and Hausermann, Fay and Hacker (1971) in *Ae. aegypti* mosquitoes using mark-release-recapture method. Ecological features including accessible water, vegetation patterns, humidity, contribute to determining the mosquito distribution. The range of dispersal is dependent upon a mosquito's ability to remain in flight and the availability and abundance of shelter, food sources, hosts for blood meals and suitable oviposition sites (Sheppard et al., 1969). Suitable host availability may reduce dispersal as reported by Suwonkerd et al. (2006) where fewer *Ae. aegypti* mosquitoes exited a hut when a human host was present compared to controls with the presence a dog, or with no human host.

Given that dispersal range is an important aspect of dengue transmission, much research has been conducted attempting to determine how far *Ae. aegypti* adults travel. A characteristic feature of *Ae. aegypti* is that they rarely disperse far from where they eclose (i.e. emergence as an adult from the pupa) (Getis et al., 2003), therefore, the presence of adult forms is for practical purposes an accurate indication of the proximity of breeding sites. Adults only disperse further when a vital requirement is limiting or absent or there is a disturbance. Typically, adult *Ae. aegypti* mosquitoes travel relatively short distances of up to 100 m, although longer dispersal estimates of about 800 m have been observed, particularly when host density is low and female mosquitoes are starved (McDonald, 1977; Honório et al., 2003; Harrington et al., 2005).

Overall, most studies show a very short dispersal distance for *Ae. aegypti*. This species has been reported to usually fly from 50 m to 300 m during its lifetime, with mean dispersal distances of 28 m to 199 m (Harrington et al., 2005). Experiments in different parts of the world involving the release and recapture of adults suggest that most are recovered within 20 m to 50 m of the release point, with a small percentage reaching distances greater than 170 m and not more than 200 m (Morlan and Hayes, 1958; Sheppard et al., 1969; McDonald, 1977; Trpis and Häusermann, 1986; Rodhain and Rosen, 1997; Muir and Kay, 1998; Ordoñez-Gonzalez et al., 2001; Harrington et al., 2005; Russell et al., 2005; Maciel-de-Freitas, Codeço and Lourenço-de-Oliveira, 2007a, 2007b; Valerio et al., 2012).

Even if important variations in mosquito daily and lifetime dispersal rates have been reported, however, the examination of the mean distance travelled (MDT) and the flight range of mosquitoes, as opposed to the maximum distance travelled, may be a more epidemiologically-important parameter (Harrington et al., 2005). Many studies using mark-release-recapture methods (above-mentioned) have reported a flight range for *Ae. aegypti* shorter than the largest observed dispersal of 800 m. And the majority of re-captured mosquitoes were collected at the house of release or neighbouring houses, suggesting females are rarely expected to visit more than two or three houses in their lifetime. In a Kenyan village, McDonald (1977) recaptured a majority of mosquitoes within the house where they were released over 12 days. Marked mosquitoes released in a tire dump in New Delhi, India, dispersed with maximum distances from 50 m to 200 m, but most were recaptured within 50 m of the release point (Reuben, Yasuno and Panicker, 1972). Similarly, Muir and Kay (1998) reported females having a MDT of 56 m.

It has also been observed that females are less likely to disperse from houses with a large number of available oviposition sites (Edman et al., 1998). Given that most *Ae. aegypti*

do not disperse very far, containers in close proximity to other productive vessels are more likely to be oviposition sites and to receive a large number of eggs. Holding other attributes constant, containers in areas of dense larval habitat will have a greater probability of being productive with a greater abundance of pupae than areas where suitable wet containers are rare and thus have a spatially-dispersed distribution. This low dispersal is a limit to the use of the autodissemination technique[2] for control in large areas, which would require a high density of dissemination stations (Devine et al., 2009).

In some studies, released mosquitoes tended to cluster around houses with some dispersal towards adjacent houses, and mosquitoes released on the perimeter of villages moved towards the centre of the village (Sheppard et al., 1969; Trpis and Hausemann, 1986; Getis et al., 2003; Harrington et al., 2005; Maciel-de-Freitas et al., 2006). The relatively large numbers and duration of DENV infected females captured in houses with confirmed dengue cases in Merida, Mexico may further indicate high fidelity between *Ae. aegypti* mosquitoes and place of pupal emergence (García Rejón et al., 2008).

The rate at which *Ae. aegypti* spreads to new areas outside of its native range is highly correlated with human activities that aid in its dispersal, including modes of transport. Boats, planes and terrestrial vehicles (e.g. cars, trucks, buses) also play a role on long-range human-mediated dispersal of adults and eggs. *Ae. aegypti* can "hitch a ride" in these vehicles, resulting in long-distance transport (Ritchie, 2014). In the Peruvian Amazon the incidence of *Ae. aegypti* coincides with interconnecting roads and highways and to a lesser extent, routes of boat traffic between ports (Guagliardo et al., 2014). Abandoned bottles, tires and other containers resulting from human activities along these travel routes provide a favoured habitat for the larval development of *Ae. aegypti* (Flores et al., 2005) and likely play a role in expanding its range. Furthermore, Chadee, Doon and Severson (2007) indicated that prevailing weather patterns may potentially influence dispersion.

Results from two classes of markers (SNPs) show strong evidence of limited gene flow across Uriah Butler Highway (UBH) in Trinidad island (Trinidad and Tobago), effectively fragmenting the populations on the east and west side of the highway (Hemme et al., 2010). Although the distance across the highway is well within dispersal estimates for *Ae. aegypti*, lack of cover and shade may have made the UBH a harsh environment for mosquitoes to transect. This is supported by Tun-Lin, Kay and Barnes (1995) who reported shade as a significant factor impacting the presence of *Ae. aegypti* in premise surveys and Russell et al. (2005) confirmed that released *Ae. aegypti* dispersal patterns were non-random with more mosquitoes being recaptured along a corridor with heavy shading from trees and vegetation. Furthermore, oviposition sites were most likely minimal, even along peripheral ditches and absence of blood meal hosts may have dissuaded migration across the UBH and prevented a stepping stone model of colonisation from occurring over UBH.

Population density and distribution

A primary determinant of adult mosquito population density concerns the types and number of containers in a given environment. Adult production is unevenly distributed across potential larval development sites.

In most cases, a few key types of containers are responsible for a large proportion of the pupal, and thus adult, production (Morrison et al., 2004; Focks and Alexander, 2006; Koenraadt et al., 2008). Protective measures such as lids, larvicide, removal of discarded and unused containers or biological agents have reduced adult vector population density

(Kay and Nam, 2005; Morrison et al., 2008). Container capacity, water temperature, source of water and container location, all of which can vary seasonally (Strickman and Kittayapong, 2002; Lenhart et al., 2006; Koenraadt et al., 2008), have been cited as important ecological factors affecting the production of adult *Ae. aegypti* (Morrison et al., 2004; Barrera, Amador and Clark, 2006a). Access to humans for blood feeding is additionally important for the production of *Ae. aegypti* adults (Ritchie, 2014).

A number of studies have also found that *Ae. aegypti* abundance is not homogeneous among households, with disproportionate numbers of immature and adult mosquitoes clustered in key premises (Tun-Lin, Kay and Barnes, 1995; Getis et al., 2003; Barrera, Amador and Clark, 2006b). A study of *Ae. aegypti* production in American Samoa found that containers were more productive on average in houses with a large number of containers (Lambdin et al., 2009). To this point, the relationship between productivity and the spatial distribution of containers has not been rigorously examined.

Population modelling

Spatial models of *Ae. aegypti* could provide an important advance toward model-guided vector control and risk assessment (Williams et al., 2008; Xu et al., 2010). One of the key challenges in modelling *Ae. aegypti* is the lack of adequate data for validation. Most models seek to represent the temporal dynamic response to climate and endogenous forces (Focks et al., 1993; Ferreira and Yang, 2003; Otero, Solari and Schweigmann, 2006; Williams et al., 2013), while others consider the spatial-temporal dynamic by introducing dispersal mechanisms (Otero, Schweigmann and Solari, 2008; Magori et al., 2009; Almeida et al., 2010).

Models describing the population dynamics of *Ae. aegypti* are either deterministic (Ferreira and Yang, 2003) or stochastic (Otero, Solari and Schweigmann, 2006) and share a common structure based on the framework of System Theory (Bertalanffy, 1975). Few available computational models simulate *Ae. aegypti* spatial-temporal dynamics. Otero, Schweigmann and Solari (2008) proposed a stochastic spatially-explicit model, based on their previous temporal model (Otero, Solari and Schweigmann, 2006), in which space is modelled as cells which are occupied by autonomous mosquito populations interconnected by flying individuals. Dispersal between cells is modulated by the availability of breeding sites. A similar approach considered both the spatial distribution of breeding sites and the dynamics of the aquatic stage of the mosquitoes (larvae and pupae) (Focks et al., 1993; Magori et al., 2009).

Notes

[1] Kairomones are semiochemicals similar to pheromones but differing by the fact that they send signals between different species.

[2] See more information on this technique in Annex A. Section: Chemical control.

References

Albeny-Simões, D. et al. (2014), "Attracted to the enemy: *Aedes aegypti* prefers oviposition sites with predator-killed conspecifics", *Oecologia*, Vol. 175, pp. 481–492.

Almeida, J.S. et al. (2010), "Multi-agent modelling and simulation of an *Aedes aegypti* mosquito population", *Environmental Modelling and Software*, Vol. 25, No. 12, pp. 1490-1507.

Bannister-Tyrrell, M. et al. (2013), "Weather-driven variation in dengue activity in Australia examined using a process-based modelling approach", *American Journal of Tropical Medicine and Hygiene*, Vol. 88, No. 1, pp. 65-72.

Banu, S. et al. (2011), "Dengue transmission in the Asia-Pacific region: Impact of climate change and socio-environmental factors", *Tropical Medicine & International Health.*, Vol. 16, pp. 598-607.

Barclay, E. (2008), "Is climate change affecting dengue in the Americas?", *Lancet*, Vol. 371, No. 9617, pp. 973-974.

Barrera, R., M. Amador and G.G. Clark (2006a), "Ecological factors influencing *Aedes aegypti* (Diptera: Culicidae) productivity in artificial containers in Salinas, Puerto Rico", *Journal of Medical Entomology*, Vol. 43, pp. 484-492.

Barrera, R., M. Amador and G.G. Clark (2006b), "Use of the pupal survey technique for measuring *Aedes aegypti* (Diptera: Culicidae) productivity in Puerto Rico", *The American Journal of Tropical Medicine and Hygiene.*, Vol. 74, pp. 290-302.

Beebe, N.W. et al. (2009), "Australia's dengue risk driven by human adaptation to climate change", *PLoS Neglected Tropical Diseases*, Vol. 3: e429.

Bertalanffy, L.V. (1975), *Perspectives on General Systems Theory. Scientific-Philosophical Studies*, E. Taschdjian (eds.), New York: George Braziller, ISBN 0-8076-0797-5 - (in Portuguese), Vozes.

Brady, O.J. et al. (2014), "Global temperature constraints on *Aedes aegypti* and *Ae. albopictus* persistence and competence for dengue virus transmission", *Parasites and Vectors*, Vol. 7, pp. 338.

Brady, O.J. et al. (2013), "Modelling adult *Aedes aegypti* and *Aedes albopictus* survival at different temperatures in laboratory and field settings", *Parasit Vectors*, Vol. 6, pp. 351.

Chadee, D.D. (2004), "Observations on the seasonal prevalence and vertical distribution patterns of oviposition by *Aedes aegypti* (L.) (Diptera: Culicidae) in urban high-rise apartments in Trinidad, West Indies", *Journal of Vector Ecology*, Vol. 29, No. 2, pp. 323-330.

Chadee, D.D., R. Doon and D.W. Severson (2007), "Surveillance of dengue fever cases using a novel *Aedes aegypti* population sampling method in Trinidad, West Indies: the cardinal points approach", *Acta Tropica*, Vol. 104, pp. 1-7.

Clements, A.N. (1999), *The Biology of Mosquitoes, Volume II. "Sensory Reception and Behaviour"*, CABI Publishing, Oxon.

Da Costa-Ribeiro, M.C.V., R. Lourenço-de-Oliveira and A.B. Failloux (2006), "Geographic and temporal genetic patterns of *Aedes aegypti* populations in Rio de Janeiro, Brazil", *Tropical Medicine & International Health*, Vol. 11, pp. 1276-1285.

Devine, G.J. et al. (2009), "Using adult mosquitoes to transfer insecticides to *Aedes aegypti* larval habitats", *Proceedings of the National Academy of Sciences of the United States of America*, Vol. 106, pp. 11530-11534.

Dye, C. (1986), "Vectorial capacity: Must we measure all its components?", *Parasitology*, Vol. 2, pp. 203-209.

Edman, J. et al. (1997), "Attractant resting boxes for rapid collection and surveillance of *Aedes aegypti* (L.) inside houses", *Journal of the American Mosquito Control Association*, Vol. 13, pp. 24-27.

Farjana, T. and N. Tuno (2013), "Multiple blood feeding and host-seeking behavior in *Aedes aegypti* and *Aedes albopictus* (Diptera: Culicidae)", *Journal of Medical Entomology*, Vol. 50, No. 4, pp. 838-846.

Farjana, T. and N. Tuno (2012), "Effect of body size on multiple blood feeding and egg retention of *Aedes aegypti* (L.) and *Aedes albopictus* (Skuse) (Diptera: Culicidae)", *Medical Entomology and Zoology*, Vol. 63, pp. 1-9.

Ferreira, C. and H. Yang (2003), "Study population dynamics of *Aedes aegypti* mosquito (in Portuguese)", *Tendências em Matemática Aplicada e Computacional* [*Trends in Applied and Computational Mathematics*], Vol. 4, No. 2, pp. 187-196.

Flores, A.E. et al. (2005), "Elevated alfa-esterases levels associated with permethrin tolerance in *Aedes aegypti* (L.) from Baja California, Mexico", *Pesticide Biochemistry and Physiology*, Vol. 82, pp. 66-78.

Focks, D.A. and N. Alexander (2006), *A Multi-Country Study on the Methodology for Surveys of* Aedes aegypti *Pupal Productivity: Findings and Recommendations*, World Health Organization, Geneva.

Focks, D.A. et al. (1993), "Dynamic life table model for *Aedes aegypti* (Diptera: Culicidae): Simulation results and validation", *Journal of Medical Entomology*, Vol. 30, pp. 1003-1028.

García-Rejón, J.E. et al. (2008), "Dengue virus-infected *Aedes aegypti* in the home environment", *American Journal of Tropical Medicine and Hygiene*, Vol. 79, No. 6, pp. 940-950.

Garrett-Jones, C. (1964), "Prognosis for interruption of malaria transmission through assessment of the mosquito's vectorial capacity", *Nature*, Vol. 204, pp. 1173-1175.

Garrett-Jones, C. and G.R. Shidrawi (1969), "Malaria vectorial capacity of a population of *Anopheles gambiae*", *Bulletin of the World Health Organization*, Vol. 40, pp. 531-545.

Getis, A. et al. (2003), "Characteristics of the spatial pattern of the dengue vector, *Aedes aegypti*, in Iquitos, Peru", *The American Journal of Tropical Medicine and Hygiene*, Vol. 69, pp. 494-505.

Guagliardo, S.A. et al. (2014), "Patterns of geographic expansion of *Aedes aegypti* in the Peruvian Amazon", *PLoS Neglected Tropical Diseases*, Vol. 8, No. 8: e3033.

Hales, S. et al. (2002), "Potential effect of population and climate changes on global distribution of dengue fever: An empirical model", *Lancet*, Vol. 360, pp. 830-834.

Halstead, S.B. (2008), "Dengue virus - Mosquito interactions", *Annual Review of Entomology*, Vol. 53, pp. 273-291.

Hancock, P.A. et al. (2016), "Density dependent population dynamics in *Aedes aegypti* slow the spread of wMel *Wolbachia*", *Journal of Applied Ecology*, Vol. 53, No. 3, pp. 785-793.

Haramis, L.D. (1983), "Increased adult size correlated with parity in *Aedes triseriatus*", *Mosquito News*, No. 43, pp. 77-79.

Harrington, L.C. et al. (2005), "Dispersal of the dengue vector *Aedes aegypti* within and between rural communities", *American Journal of Tropical Medicine and Hygiene*, Vol. 72, No. 2, pp. 209-220.

Hausermann, W., R.W. Fay and C.S. Hacker (1971), "Dispersal of genetically marked female *Aedes aegypti* in Mississippi", *Mosquito News*, Vol. 31, pp. 37-51.

Hemme, R.R. et al. (2010), "Influence of urban landscapes on population dynamics in a short-distance migrant mosquito: Evidence for the dengue vector *Aedes aegypti*", *PLoS Neglected Tropical Diseases*, Vol. 4, No. 3: e634.

Honório, N.A. et al. (2003), "Dispersal of *Aedes aegypti* and *Aedes albopictus* (Diptera: Culicidae) in an urban endemic dengue area in the state of Rio de Janeiro, Brazil", *Memórias do Instituto Oswaldo Cruz*, Vol. 98, pp. 191-198.

Huber, K. et al. (2002), "Genetic differentiation of the dengue vector, *Aedes aegypti* (Ho Chi Minh City, Vietnam) using microsatellite markers", *Molecular Ecology*, Vol. 11, No. 9, pp. 1629-1635.

Hutchinson, G.E. (1957), "Concluding remarks", *Cold Spring Harbor Symposia on Quantitative Biology*, Vol. 22, pp. 415-427.

Jansen, C.C. and N.W. Beebe (2010), "The dengue vector *Aedes aegypti*: What comes next", *Microbes and Infection*, Vol. 12, pp. 272-279.

Jetten, T.H. and D.A. Focks (1997), "Potential changes in the distribution of dengue transmission under climate warming", *The American Journal of Tropical Medicine and Hygiene*, Vol. 57, pp. 285-297.

Juliano, S.A. and L.P. Lounibos (2005), "Ecology of invasive mosquitoes: Effects on resident species and on human", *Ecology Letters*, Vol. 8, pp. 558-574.

Kay, B. and V.S. Nam (2005), "New strategy against *Aedes aegypti* in Vietnam", *Lancet*, Vol. 365, No. 9459, pp. 613-617.

Klowden, M.J. and A.O. Lea (1978), "Blood meal size as a factor affecting continued host-seeking by *Aedes aegypti* (L.)", *American Journal of Tropical Medicine and Hygiene*, Vol. 27, pp. 827-831.

Koenraadt, C.J.M. et al. (2008), "Spatial and temporal patterns in pupal and adult production of the dengue vector *Aedes aegypti* in Kamphaeng Phet, Thailand", *The American Journal of Tropical Medicine and Hygiene*, Vol. 79, pp. 230-238.

Lambdin, B.H. et al. (2009), "Dry Season production of filariasis and dengue vectors in American Samoa and comparison with wet season production", *The American Journal of Tropical Medicine and Hygiene*, Vol. 81, pp. 1013-1019.

Lenhart, A.E. et al. (2006), "Use of the pupal/demographic-survey technique to identify the epidemiologically important types of containers producing *Aedes aegypti* (L.) in a dengue-endemic area of Venezuela", *Annals of Tropical Medicine and Parasitology*, Vol. 100 (Supple. 1), pp. S1-S7.

Liu-Helmersson, J. et al. (2016), "Climate change and *Aedes* vectors: 21st century projections for dengue transmission in Europe", *EBioMedicine*, Vol. 7, pp. 267–277.

Lounibos, L.P. et al. (2002), "Does temperature affect the outcome of larval competition between *Aedes aegypti* and *Aedes albopictus*?", *Journal of Vector Ecology*, Vol. 27, pp. 86-95.

Lounibos, L.P. (1981), "Habitat segregation among African treehole mosquitoes", *Ecological Entomology*, Vol. 6, pp. 129-154.

Lozano-Fuentes, S. et al. (2012), "The dengue virus mosquito vector *Aedes aegypti* at high elevation in Mexico", *The American Journal of Tropical Medicine and Hygiene*, Vol. 87, No. 5, pp. 902-909.

Maciel-de-Freitas, R., C.T. Codeço and R. Lourenço-de-Oliveira (2007a), "Daily survival rates and dispersal of *Aedes aegypti* females in Rio de Janeiro, Brazil", *American Journal of Tropical Medicine and Hygiene*, Vol. 76, No. 4, pp. 659-665.

Maciel-de-Freitas, R., C.T. Codeço and R. Lourenço-de-Oliveira (2007b), "Body size-associated survival and dispersal rates of *Aedes aegypti* in Rio de Janeiro", *Medical and Veterinary Entomology*, Vol. 21, pp. 284-292.

Maciel-de-Freitas, R. et al. (2006), "Movement of dengue vectors between human the human modified environment and an urban forest in Rio de Janeiro", *Journal of Medical Entomology*, Vol. 43, pp. 1112-1120.

Mackenzie, J.S., D.J. Gubler and L.R. Petersen (2004), "Emerging flaviviruses: The spread and resurgence of Japanese encephalitis, West Nile and dengue viruses", *Nature Medicine*, Vol. 10, S98-S109.

Magori, K. et al. (2009), "A stochastic, spatially explicit modelling tool for studying *Aedes aegypti* population replacement and population suppression strategies", *PLoS Negleted Tropical Diseases*, No. 3, No. 9, pp. 1-18.

McDonald, P.T. (1977), "Population characteristics of domestic *Aedes aegypti* (DipteraL Culicidae) in villages on the Kenya coast: II. Dispersal within and between villages", *Journal of Medical Entomology*, Vol. 14, pp. 49-53.

Mogi, M. (2007), "Insects and other invertebrate predators", *Journal of the American Mosquito Control Association*, Vol. 23, No. sp2, pp. 93-109.

Morlan, H.B. and R.O. Hayes (1958), "Urban dispersal and activity of *Aedes aegypti*", *Mosquito News*, Vol. 18, pp. 137-144.

Morrison, A.C. et al. (2008), "Defining challenges and proposing solutions for control of the virus vector *Aedes aegypti*", *PLoS Medicine*, Vol. 5: e68.

Morrison, A.C. et al. (2004), "Temporal and geographic patterns of *Aedes aegypti* production in Iquitos, Peru", *Journal of Medical Entomology*, Vol. 42, pp. 502-510.

Muir, L.E. and B.H. Kay (1998), "*Aedes aegypti* survival and dispersal estimated by mark-release-recapture in northern Australia", *The American Journal of Tropical Medicine and Hygiene*, Vol. 58, pp. 277-282.

Murrell, E.G. and A.J. Steven (2008), "Detritus type alters the outcome of interspecific competition between *Aedes aegypti* and *Aedes albopictus* (Diptera: Culicidae)", *Journal of Medical Entomology*, Vol. 45, No. 3, pp. 375-385.

Nasci, R.S. (1991), "Influence of larval and adult nutrition on biting persistence in *Aedes aegypti* (Diptera: Culicidae)", *Journal of Medical Entomology*, Vol. 28, pp. 522-526.

Nazri, C.D., A.H. Ahmad and R. Ismail (2013), "Habitat Characterization of *Aedes* Sp. Breeding in Urban Hotspot Area", *Procedia - Social and Behavioral Sciences*, Vol. 85, pp. 100-109.

Ooi, E.E. and D.J. Gubler (2009), "Global spread of epidemic dengue: The influence of environmental change", *Future Virology*, Vol. 4, pp. 571-580.

Ordóñez-Gonzalez, J.G. et al. (2001), "The use of sticky ovitraps to estimate dispersal of *Aedes aegypti* in northeastern Mexico", *Journal of the American Mosquito Control Association*, Vol. 17, No. 2, pp. 93-97.

Otero, M., N. Schweigmann and H.G. Solari (2008), "A stochastic spatial dynamical model for *Aedes aegypti*", *Bulletin of Mathematical Biology*, Vol. 70, No. 5, pp. 1297-1325.

Otero, M., H.G. Solari and N. Schweigmann (2006), "A stochastic population dynamics model for *Aedes aegypti*: Formulation and application to a city with temperate climate", *Bulletin of Mathematical Biology*, Vol. 68, No. 8, pp. 1945-1974.

Packer, M.J. and P.S. Corbet (1989), "Size variation and reproductive success of female *Aedes punctor* (Diptera: Culicidae)", *Ecological Entomology*, Vol. 14, pp. 297-309.

Patz, J.A. et al. (1998), "Dengue fever epidemic potential as projected by general circulation models of global climate change", *Environmental Health Perspectives*, Vol. 106, pp. 147-153.

Pumpuni, C.B. and E.D. Walker (1989), "Population size and survivorship of adult *Aedes triseriatus* in a scrap tireyard in northern Indiana", *Journal of the American Mosquito Control Association*, Vol. 5, pp. 166-172.

Raxworthy, C.J. et al. (2007), "Applications of ecological niche modelling for species delimitation: A review and empirical evaluation using day geckos (*Phelsuma*) from Madagascar", *Systematic Biology*, Vol. 56, No. 6, pp. 907-923.

Reiter, P. et al. (1995), "Dispersal of *Aedes aegypti* in an urban area after blood feeding as demonstrated by rubidium-marked eggs", *The American Journal of Tropical Medicine and Hygiene*, Vol. 52, pp. 177-179.

Renshaw, M., M.W. Service and M.H. Birley (1994), "Size variation and reproductive success in the mosquito *Aedes cantans*", *Medical and Veterinary Entomology*, Vol. 8, pp. 179-186.

Reuben, R., M. Yasuno and K.N. Panicker (1972), "Studies on the dispersal of *Aedes aegypti* at two localities in Delhi", WHO/VBC/72.388, World Health Organization.

Reyes-Villanueva, F. (2004), "Egg development may require multiple bloodmeals among small *Aedes aegypti* (Diptera: Culicidae) field collected in northeastern Mexico", *Florida Entomologist*, Vol. 87, pp. 630-632.

Ritchie, S.A. (2014), "Dengue vector bionomics: Why *Aedes aegypti* is such a good vector", in D. Gubler et al. (eds), *Dengue and Dengue Hemorrhagic Fever*, CAB International, Oxfordshire.

Rodhain, F. and L. Rosen (1997), "Mosquito vectors and dengue virus vector relationships", in D.J. Gubler, and G. Kuno (eds), *Dengue and Dengue Hemorrhagic Fever*, CAB International, New York, pp. 45-60.

Rueda, L.M. et al. (1990), "Temperature-dependent development and survival rates of *Culex quinquefasciatus* and *Aedes aegypti* (Diptera. Culicidae)", *Journal of Medical Entomology*, Vol. 27, No. 5, pp. 892-898.

Russell, R.C. et al. (2009), "Dengue and climate change in Australia: Predictions for the future should incorporate knowledge from the past", *Medical Journal of Australia*, Vol. 190, pp. 265-268.

Russell, R.C. et al. (2005), "Mark-release-recapture study to measure dispersal of the mosquito *Aedes aegypti* in Cairns, Queensland, Australia", *Medical and Veterinary Entomology*, Vol. 19, pp. 451-457.

Russell, B.M., B.H. Kay and W. Shipton (2001), "Survival of *Aedes aegypti* (Diptera: Culicidae) eggs in surface and subterranean breeding sites during the northern Queensland dry season", *Journal of Medical Entomology*, Vol. 38, pp. 441–445.

Scott, T.W. et al. (2000), "Longitudinal studies of *Aedes aegypti* (Diptera: Culicidae) in Thailand and Puerto Rico: Population dynamics", *Journal of Medical Entomology*, Vol. 37, No. 1, pp. 77-88.

Scott, T.W. et al. (1993), "Blood-feeding patterns of *Aedes aegypti* (Diptera: Culicidae) collected in a rural Thai village", *Journal of Medical Entomology*, Vol. 30, No. 5, pp. 922-927.

Shaalan, E.A. and D.V. Canyon (2009), "Aquatic insect predators and mosquito control", *Tropical Biomedicine*, Vol. 26, No. 3, pp. 223-261.

Sheppard, P. et al. (1969), "The dynamics of an adult *Aedes aegypti* in relation to dengue hemorrhagic fever in Bangkok", *The Journal of Animal Ecology*, Vol. 38, pp. 661-702.

Strickman, D. and P. Kittayapong (2002), "Dengue and its vectors in Thailand: Introduction to the study and seasonal distribution of *Aedes* larvae", *American Journal of Tropical Hygiene and Medicine*, Vol. 67, pp. 247-259.

Sulaiman, S. et al. (1990), "Detection of *Aedes aegypti* (L.) (Diptera: Culicidae) predators in urban slums in Malaysia using the precipitin test", *Mosquito Borne Diseases Bulletin*, Vol. 7, pp. 123–126.

Suwonkerd, W. et al. (2006), "The effect of host type on movement patterns of *Aedes aegypti* (Diptera: Culicidae) into and out of experimental huts in Thailand", *Journal of Vector Ecology*, Vol. 31, pp. 311-318.

Teng, H.J. and C.S. Apperson (2000), "Development and survival of immature *Aedes albopictus* and *Aedes triseriatus* (Diptera: Culicidae) in the laboratory: Effects of density, food, and competition on response to temperature", *Journal of Medical Entomology*, Vol. 37, No. 1, pp. 40-52.

Trpis, M. (1973), "Interaction between the predator *Toxorhynchites brevipalpis* and its prey *Aedes aegypti*", *Bulletin of the World Health Organization*., Vol. 49, No. 4, pp. 359.

Trpis, M. and W. Hausermann (1986), "Dispersal and other population parameters of *Aedes aegypti* in an African village and their possible significance in epidemiology of vector-borne diseases", *The American Journal of Tropical Medicine and Hygiene*, Vol. 35, pp. 1263-1279.

Trpis, M., W.O. Haufe and J.A. Shemanchuk (1973), "Embryonic development of *Aedes* (O.) *sticticus* (Diptera: Culicidae) in relation to different constant temperatures", *The Canadian Entomologist*, Vol. 105, pp. 43-50.

Tun-Lin, W., B.H. Kay and A. Barnes (1995), "The premise condition index: A tool for streamlining surveys of *Aedes aegypti*", *American Journal of Tropical Medicine and Hygiene*, Vol. 53, pp. 591-594.

Tun-Lin, W. et al. (2009), "Reducing costs and operational constraints of dengue vector control by targeting productive breeding places: A multi-country non-inferiority cluster randomized trial", *Tropical Medicine & International Health*, Vol. 14, pp. 1143-1153.

Urdaneta-Marquez, L. et al. (2008), "Genetic relationships among *Aedes aegypti* collections in Venezuela as determined by mitochondrial DNA variation and nuclear single nucleotide polymorphisms", *The American Journal of Tropical Medicine and Hygiene*, Vol. 78, No. 3, pp. 479-491.

Valerio, L. et al. (2012). "Dispersal of male *Aedes aegypti* in a coastal village in southern Mexico", *American Journal of Tropical Medicine and Hygiene*, Vol. 86, No. 4, pp. 665-676.

Williams, C.R. et al. (2015), "Testing the impact of virus importation rates and future climate change on dengue activity in Malaysia using a mechanistic entomology and disease model", *Epidemiol. Infect*, pp. 1-9.

Williams, C.R. et al. (2014), "Bionomic response of *Aedes aegypti* to two future climate change scenarios in far north Queensland, Australia: Implications for dengue outbreaks", *Parasites and Vectors*, Vol. 7, pp. 447.

Williams, C.R. et al. (2013), "Productivity and population density estimates of the dengue vector mosquito *Aedes aegypti* (*Stegomyia aegypti*) in Australia", *Medical and Veterinary Entomology*, Vol. 27, pp. 313–322.

Williams, C.R. et al. (2010), "The extinction of dengue through natural vulnerability of its vectors", *PLoS Neglected Tropical Diseases*, Vol. 4, No. 12: e922.

Williams, C.R. et al. (2008), "Rapid estimation of *Aedes aegypti* population size using simulation modeling, with a novel approach to calibration and field validation", *Journal of Medical Entomology*, Vol. 45, No. 6, pp. 1173-1179.

Xu, C. et al. (2010), "Understanding uncertainties in model-based predictions of *Aedes aegypti* population dynamics", *PLoS Neglected Tropical Diseases*, Vol. 4, No. 9: e830.

Xue, R.D., J.D. Edman and T.W. Scott (1995), "Age and body size effects on blood meal size and multiple blood feeding by *Aedes aegypti* (Diptera: Culicidae)", *Journal of Medical Entomology*, Vol. 32, pp. 471-474.

Yan, G., D.D. Chadee and D.W. Severson (1998), "Molecular population genetics of the yellow fever mosquito: Evidence for genetic hitchhiking effects associated with insecticide resistance", *Genetics*, Vol. 148, pp. 793-800.

Yuval, B., J.W. Wekesa and R.K. Washino (1993), "Effect of body size on swarming behavior and mating success of male *Anopheles freeborni* (Diptera: Culicidae)", *Journal of Insect Behaviour*, Vol. 6, pp. 333-342.

Annex A. **Control of the mosquito *Ae. aegypti***

This annex describes the current strategies put in place to limit or eradicate mosquitoes that transmit disease pathogens: chemical control using larvicides and insecticides, biological control based on introduction of other organisms, use of Wolbachia *bacteria in methods for controlling virus transmission through reduction or replacement of the mosquito* Aedes aegypti *population. Research is also conducted on genetic control of* Ae. aegypti. *Then information is given on relevant environmental management aiming to limit its propagation, including integrated control management and the prevention of insecticide resistance.*

Current control strategies

Mosquitoes can be vectors (transmitters) of several infectious diseases to humans and animals and are thus of significant importance to public health. The aim of mosquito control, in general, is to prevent mosquito bites, to maintain mosquito populations at "acceptable" densities, to minimise mosquito-host contact and to reduce the longevity of female mosquitoes (Foster and Walker, 2002).

Vector control is any method to limit or eradicate mosquitoes that transmit disease pathogens. Disease control is the reduction in the incidence, prevalence, morbidity or mortality of an infectious disease to a locally acceptable level or, if possible, its elimination or eradication. In order to be sustainable, a vector control strategy must limit the spread of resistance to insecticides within target mosquito populations.

Aedes aegypti control is generally performed in the context of public health because it is the vector of Zika, dengue, chikungunya and yellow fever, and a number of other diseases. Particularly for Zika, dengue and chikungunya, there are no vaccines, therapeutic treatments or cure. Preventing or reducing Zika, dengue and chikungunya virus transmission depends entirely on control of the mosquito vectors or interruption of human-vector contact (WHO, 2009b). Eradication of *Ae. aegypti* populations may be achievable, but is rarely sustainable, therefore, the present paradigm is to reduce mosquito density below disease transmission threshold levels rather than eliminate entire populations (McCall and Kittayapong, 2006).

Ae. aegypti control largely depends on organised control programmes at the community level administered by ministries of health undertaken together with some self-protection measures. Because *Ae. aegypti* lives in close affinity with humans and human-made ecosystems, it is an ideal candidate for integrated control (utilisation of multiple methods to provide control), which is summarised in the following Table A A.1 and briefly described in the following sections.

Chemicals for mosquito control may only be used in accordance with national legislation and approval of the products. Some of the chemicals mentioned as examples in Table A A.2 might be allowed in some countries but not in others.

Detailed information on the mosquito ecology, dispersal and the distribution of human habitats (see the chapter on Ecology) can be useful to vector control agencies for better targeting populations for suppression. Control programmes can be built on an urban area divided into zones of control along landscape features that are large enough to impede mosquito dispersal. This technique allows for the possibility of local elimination of *Ae. aegypti* mosquitoes, barring or at least minimising re-infestation due to the active transportation of the mosquito. Furthermore, during outbreaks, control agencies can more accurately target areas of higher risk along these same control zones. Understanding the role of landscape features on population dispersal is likely critical to achieving success with any *Ae. aegypti* control strategy.

Chemical control

Immature stages: The control of *Ae. aegypti* larvae and pupae can be effected by treating containers holding water (specifically those that are productive breeding-sites and cannot otherwise be eliminated or managed) with insecticides (larvicides). Larvicides such as diflubenzuron, novaluron pyriproxyfen, fenthion, pirimiphos-methyl, temephos and

spinosad (approved by WHOPES) target the immature mosquitoes living in water before they become biting adults.

Table A A.1. Summary of control tools/strategies available for *Ae. aegypti*

Method	Description	Examples
CHEMICAL CONTROL	Immature stages	Treating containers (breeding-sites) with for e.g. Temephos 1% Sand Granule; biorational larvicides; insect growth regulators (IGR) such as methoprene and pyriproxyfen, spinosad
	Adult in medium/large areas or houses	Aerial treatments, indoor spraying, surface treatments
	Personal protection	Domestic insecticides, repellents (natural or synthetic), insecticide-treated materials and paints
BIOLOGICAL CONTROL	Immature stages and adults (the whole population)	Fish, dragonflies, copepods, *Bti*, *Toxorhynchites*, *Wolbachia*
GENETIC CONTROL (self-limiting)	Immature stages and adults (the whole population)	Self-limiting insects, sterile insect technique, others
GENETIC CONTROL (population replacement)	Forces genes/organism through the whole population	Gene drive systems (i.e. HEGs and CRISPR), *Wolbachia*
ENVIRONMENTAL MANAGEMENT	Modification: permanent transformations in some characteristics to the vector breeding habitats Manipulation: temporal changes (management) to affect the breeding sites (key) behaviour Structural changes in human habitation and human behaviour	Draining/cleaning/recycling/disposal of breeding-sites or potential larval habitats Installation of reliable piped water supply to communities, comprehensive coverage and proper disposal of solid waste collection, filling, draining public spaces Public sensitisation to reduce the availability of breeding sites (source reduction) Installing mosquito screening on windows, doors and other entry points. Using mosquito nets Paints, peridomestic veneering to contribute eliminating natural habitats

Source: Modified from PAHO (1994), *Dengue and Dengue Hemorrhagic Fever in the Americas: Guidelines for Prevention and Control*, PAHO Scientific Publication 548, Pan American Health Organization, Washington, DC, and McCall, P.J. and P. Kittayapong (2006), "Control of dengue vectors: Tools and strategies", in *Report of the Scientific Working Group Meeting on Dengue*, World Health Organization, Geneva, WHO/TDR 2007, pp. 110-119.

The application of larvicides can also be done by ground or aerial treatments. However, the high density of small habitats (< 200 mL) makes it very difficult to treat a reasonable proportion of highly disseminated breeding sites. It has been proposed recently to use auto-dissemination of pyriproxifen by adult females themselves to their breeding sites, after their contamination using dissemination stations (Devine et al., 2009). This approach is very efficient but has a short range of action because of low rates of adult dispersal. It has thus been proposed to release sterile males contaminated with pyriproxifen to contaminate the females through venereal transfer, an approach called the "boosted sterile insect technique" (Bouyer and Lefrançois, 2014). This control method has been successfully demonstrated recently in a field trial against *Ae. albopictus* at

a very small scale (Mains, Brelsfoard and Dobson, 2015), and it is a major research axis to improve larvicidal control at the moment.

Adult: The control of adult vectors with insecticides (adulticides), applied either as residual surface treatments or as space treatments (thermal fogging and ultra-low volume aerosol sprays), is expected to impact mosquito densities, longevity and other transmission parameters. Insecticides from three chemical groups, namely pyrethroids, organophosphates and carbamates, are recommended by WHOPES both for indoor and outdoor spraying (WHO, 2003). The application of adulticides can be done by ground or aerial treatments but has a very short-term and local action.

Indoor residual spraying (IRS) involves the spraying of an insecticide on all the walls inside the house. This is usually done only once or twice a year because the effect is lasting and continues to kill mosquitoes for many months after treatment. Targeted indoor residual spraying involves spraying dark shady areas used by adult *Ae. aegypti* as resting places, such as under beds and tables, inside closets and dark objects such as plastic crates and suitcases. This method uses less pesticide and has been successfully used to protect residences from dengue transmission (Vazquez-Prokopec et al., 2017).

Indoor space-spraying (ISS) involves delivery of an insecticidal fog inside houses. However, space sprays do not leave a residual layer providing long-term control and have found to be ineffective for dengue control (Esu et al., 2010).

Outdoor fogging is the method commonly used in many parts of the world. The insecticide is usually sprayed from vehicles as a cloud of "fog" outside houses, targeting the flying female mosquitoes. Vector populations can be suppressed over large areas by the use of space sprays released from low-flying aircraft, especially where gaining access with ground equipment is difficult and extensive areas must be treated rapidly. It is generally ineffective against *Ae. aegypti* populations that have access to indoor harbourage sites.

Personal protection: *Ae. aegypti* exposure can be avoided with chemical products such as domestic insecticides, repellents (natural or synthetic) and insecticide-treated materials and paints including spatial repellents such as metofluthrin (Ritchie and Devine, 2013).

In general, pyrethroids are the main active ingredients in household aerosol products available to the public. Where indoor biting occurs, household insecticide aerosol products, mosquito coils or other insecticide vaporisers may reduce biting activity (WHO, 2009a).

Numerous insect repellent products are available commercially in a variety of formulations. Some of these products contain active ingredient(s) from botanical origin and some are synthetic organic products, with a vast majority available as sprays. Repellents may be applied to exposed skin or to clothing. Repellents recommended contain DEET (N, N-diethyl-3-methylbenzamide), IR3535 (3-[N-acetyl-N butyl]-aminopropionic acid ethyl ester) or Icaridin (1-piperidinecarboxylic acid, 2-(2-hydroxyethyl)-1 methylpropylester) (WHO, 2009a).

Long-lasting insecticidal netting (LLIN) is factory-produced mosquito netting pre-loaded with synthetic pyrethroid insecticide that is intended to retain its biological activity for at least 20 standard washes under laboratory conditions, and three years of recommended use under field conditions (WHO, 2013). Deployed as bed nets, LLIN potentially can reduce human biting rates and vector longevity at both household and community levels (McCall and Kittayapong, 2006). In Latin America, encouraging results for *Ae. aegypti*

control have also been obtained when LLIN are deployed as window or door screens, curtains or as container covers (Vanlerberghe et al., 2011; Rizzo et al., 2012; Manrique-Saide et al., 2015).

Biological control

Biological control is based on the introduction of organisms that prey upon, parasitise, compete with or otherwise reduce populations of the target species. *Bacillus thuringiensis* var. *israelensis* (*Bti*) is an entomopathogenic bacterium that has demonstrated high efficacy against *Ae. aegypti* larvae and is commercially available in different formulations that can be utilised in a variety of breeding habitats (Lacey, 2007; Boyce et al., 2013). Its strain AM65-52 in a water-dispersible granulated formulation is recommended by WHOPES (2016).

Other biological control agents that have been used for larval control of *Ae. aegypti* include species of larvivorous fish (WHO/EMRO, 2003) e.g. *Poecilia reticulata*, dragonflies (Sebastian et al., 1980, 1990; Venkatesh and Tyagi, 2013) and predatory copepods (Copepoda: Cyclopoidea) (Kay et al., 2012) which have proved effective in operational contexts in specific container habitats, but seldom on a large scale.

Wolbachia *as a biological control method for virus transmission*

Uses of Wolbachia *in control methods*

Wolbachia bacteria can be used to control *Ae. aegypti* and the diseases it spreads in two different ways, population reduction or population replacement:

a) Population reduction: *Ae. aegypti* males infected with *Wolbachia* are released. When the infected males mate with wild females, no offspring are produced, and with such release renewed over a period of time, the mosquito population can be reduced. It is important with this approach that no infected females be released as that could potentially lead to failure of the control programme; the infected females can pass *Wolbachia* onto their offspring, which survive and can spread into the environment. To date, there have been no successful suppression trials using *Wolbachia* for population reduction with *Ae. aegypti*.

b) Population replacement: *Wolbachia* can also be used in a population replacement strategy approach, similar to gene drive systems. In the wild, Wolbachia can spread through a species by a process known as cytoplasmic incompatibility (CI). CI is similar to a gene drive mechanism, which kills any offspring that are not infected with *Wolbachia*, effectively selecting for only offspring that are infected and hence spreading the *Wolbachia* through a population. The following paragraphs detail population replacement strategies being tested in *Wolbachia* and *Ae. aegypti*.

Introducing the *Wolbachia* strain wMelPop into wild populations of *Ae. aegypti* can shorten the adult mosquito lifespan, thereby theoretically reducing but not eliminating the transmission of dengue since it has not fully proven to reduce mosquito longevity shorter to the extrinsic incubation period for dengue virus (DENV). However, high fitness costs have prevented wMelPop from being successfully established in wild populations of *Ae. aegypti* in Australia and Viet Nam (Nguyen et al., 2015).

Two *Wolbachia* strains (wMel and wMelPop-CLA) have shown to confer antiviral properties to *Ae. aegypti* and limit DENV-2 infection in the mosquito by reducing the

virus' ability to disseminate from the midgut (MG) into mosquito saliva and affected mosquito fitness for disease transmission. A major open field trial was conducted in which about 300 000 *Wolbachia* wMel-infected *Ae. aegypti* mosquitoes raised under laboratory conditions were deliberately released in 2011 at 2 locations near Cairns, Australia. The frequency of *Wolbachia*-infected *Ae. aegypti* initially increased to more than 15% in both locations at two-week post-release. After additional releases, frequencies increased to > 60% and reached near fixation levels 5 weeks after releases were terminated, and these high frequencies were maintained through 2017. These observations suggest that *Wolbachia* could potentially become a powerful bio-control agent to suppress DENV transmission by *Ae. aegypti* in endemic areas, though field data demonstrating reduction of DENV transmission has not been shown.

Wolbachia *transfer into* Ae. aegypti *mosquitoes*

Although *Wolbachia* infections are relatively common in mosquitoes (Kittayapong et al., 2000; Ricci et al., 2002) including *Culex pipiens* (Yen and Barr, 1973), *Cx. quinquefasciatus*, *Ae. fluviatilis* (Moreira et al., 2009) and *Ae. albopictus* (Sinkins, Braig and O'Neill, 1995), the main vectors for dengue fever (*Ae. aegypti*) and malaria (*Anopheles* spp.) are not naturally infected by *Wolbachia*. Approaches that use *Wolbachia* for the control of diseases transmitted by uninfected, naive insects rely on the successful establishment of stable *Wolbachia* infections, usually by embryonic microinjection of *Wolbachia*-infected cytoplasm or *Wolbachia* purified from infected insect hosts.

To create stably transinfected lines, embryo injections must target the region near the pole cells in pre-blastoderm embryos in order to incorporate *Wolbachia* into the developing germline and favour the transmission of *Wolbachia* to offspring. Several *Wolbachia* strains have been transferred across sometimes phylogenetically distant insects and, importantly, the phenotypes induced by these strains in their native hosts are generally also expressed in the newly infected hosts. *Wolbachia* transinfection experiments are more likely to be successful when the donor and recipient organisms are closely related.

In line with this, the transfer of wMelPop from its natural host, *Drosophila melanogaster*, into the dengue fever vector *Ae. aegypti* was achieved in the laboratory after *Wolbachia* was first maintained by continuous passage in *Ae. albopictus in vitro* cell culture for almost four years (McMeniman et al., 2008). *Wolbachia* adapted to a mosquito intracellular environment, facilitating transinfection *in vivo*. After microinjection of thousands of *Ae. aegypti* embryos, two stable wMelPop-CLA (cell-line-adapted) lines with maternal transmission rates of approximately 100% were generated (McMeniman et al., 2009). The wMelPop-CLA-infected mosquitoes showed an approximately 50% reduction in adult lifespan, compared with their uninfected counterparts (McMeniman et al., 2009). The halving of adult mosquito lifespan and the high *Wolbachia* maternal transmission rates were also maintained in more genetically diverse outbred mosquitoes and larval nutrition did not affect the life-shortening ability of the wMelPop-CLA strain (Yeap et al., 2010).

The wMelPop-CLA infection is widespread in *Ae. aegypti* tissues, with high bacterial densities in the head (brain and ommatidia), thorax (salivary glands, muscle) and abdomen (fat tissue, reproductive tissues and malpighian tubules) (Moreira et al., 2009). Wide distribution across tissues has been found in other transinfected mosquitoes, such as *Ae. aegypti* infected with the wAlbB strain from *Ae. albopictus* (Bian et al., 2010). By using quantitative PCR and western blot analyses, this strain was also found in reproductive tissues, MG, muscles and heads, in both native *Ae. albopictus* (Dobson

et al., 1999) and the transinfected *Ae. aegypti* (Bian et al., 2010), although the densities are not as high as those found in *Ae. aegypti* infected with wMelPop-CLA.

In addition, there is evidence that *Wolbachia* infection can result in permanent genetic modification of its insect hosts in a process called Lateral gene transfer (LGT). LGT of fragments of the *Wolbachia* genome (total size approximately 1.2 Mb), ranging from 500 base pairs to more than 1 Mb, have been observed in many invertebrates, including beetles (Nikoh et al., 2008), grasshoppers (Funkhouser-Jones, 2015; Toribio-Fernández et al., 2017), wasps (Dunning-Hotopp et al., 2007), fruit flies (Dunning-Hotopp et al., 2007; Klasson et al., 2014; Choi, Bubnell and Aquadro, 2015; Morrow et al., 2015), tsetse flies (Brelsfoard et al., 2014; Nakao et al., 2016), butterflies and moths (Ahmed et al., 2016), kissing bugs (Mesquita et al., 2015), mosquitoes (Klasson et al., 2009; Hou et al., 2014), filarial nematodes (Fenn et al., 2006; Dunning-Hotopp et al., 2007; Keroack et al., 2016) and spiders (Baldo et al., 2008).

Next step

The ability of some *Wolbachia* strains to reduce the lifespan of *Ae. aegypti*, invade mosquito populations through the induction of CI and, in particular, interfere with the replication of a variety of pathogens has distinct implications for disease control. There is some evidence that the *Wolbachia* can spread through a mosquito population as predicted, and the next phase is to prove that this leads to disease reduction.

Genetic control

Many trials have been conducted using classical sterile insect technique (SIT) and self-limiting insects (OX513A transgenic line) (Alphey, 2014). Classical SIT pilot projects have been tested in Indonesia, Malaysia, Mexico, Sri Lanka and Thailand. This technology is based on the mass-rearing production of male mosquitos sterilised under X-rays or by irradiation (Gamma). This technology is very well applied on agricultural pests and other vector species like the tsetse fly (Dicko et al., 2014; Vreysen et al., 2014), and can be very powerful on insect population suppression or even eradication. However, successful population suppression for *Ae. aegypti* using SIT has yet to be demonstrated. In China, *Ae. albopictus-Wolbachia* IIT/SIT strategies that use the introduction of infected males (IIT) and sterile females (SIT) are tested to reduce wild populations (Zhang et al., 2016). In Europe, the classical SIT is considered as a biological control technique and exempted from the "GMO" regulation, unlike self-limiting insects (EFSA Panel on Genetically Modified Organisms (GMO), 2013).

Self-limiting insects are engineered with a gene that causes offspring to die before reaching functional adulthood, a species-specific control approach that has been developed for *Ae. aegypti* but which is applicable to a wide range of insects. Released mosquitoes die along with their offspring and therefore do not persist in the environment (Gorman et al., 2016). Additionally, the self-limiting OX513A mosquitoes and their offspring contain a fluorescent marker (DsRed2) that allows identification of OX513A larvae and pupae under laboratory conditions. Deployment of this technology through the release of self-limiting OX513A mosquitoes has achieved effective population suppression of wild *Ae. aegypti* in multiple trials in Brazil, the Cayman Islands and Panama (Harris et al., 2012; Carvalho et al., 2015; Gorman et al., 2016), and has been positively reviewed by regulatory bodies in Brazil, the European Union and the United States.

Environmental management

Environmental management seeks to change the environment in order to prevent or minimise vector propagation and human contact with the vector of pathogen by destroying, altering, removing or recycling non-essential containers that provide larval habitats. Such actions should be the mainstay of vector control and require important efforts for public sensitisation. Three types of environmental management are defined as follows (WHO, 1982; PAHO, 1994; Erlanger, Keiser and Utzinger, 2008; McCall, Lloyd and Nathan, 2009).

Environmental modification: Long-lasting physical transformations to reduce vector larval habitats such as the installation of reliable piped water supply to communities, including household connections.

Environmental manipulation: Temporary changes to vector habitats involving the management of "essential" containers, such as frequent emptying and cleaning by scrubbing of water-storage vessels, flower vases and desert room coolers, cleaning of gutters, sheltering stored tires from rainfall, recycling or proper disposal of discarded containers and tires, management or removal from the vicinity of homes of plants such as ornamental or wild bromeliads that collect water in the leaf axils. There are a great variety of man-made containers in backyards or patios that collect rainwater or that are filled with water by people. Disposing of unused containers, placing useful containers under a roof or protected with tight covers, and frequently changing the water of animal drinking pans and flower pots will greatly reduce the risk of dengue infections. Water storage containers should be kept clean and sealed so mosquitoes cannot use them as aquatic habitats (CDC, 2010).

Changes to human habitation or behaviour: Actions to reduce human-vector contact, such as installing mosquito screening on windows, doors and other entry points, and using mosquito nets while sleeping during daytime.

Integrated control management

Integrated vector management (IVM) is the strategic approach to vector control promoted by the World Health Organization (WHO, 2008) and includes control of the vectors of dengue. Defined as "a rational decision-making process for the optimal use of resources for vector control", IVM considers five key elements in the management process, namely (McCall, Lloyd and Nathan, 2009):

1. *Advocacy, social mobilisation and legislation* – the promotion of the IVM principles in development policies of all relevant agencies, organisations and civil society; the establishment or strengthening of regulatory and legislative controls for public health; and the empowerment of communities.
2. *Collaboration within the health sector and with other sectors* – the consideration of all options for collaboration within and between public and private sectors; planning and decision-making delegated to the lowest possible administrative level; and strengthening communication among policy-makers, managers of programmes for the control of vector-borne diseases, and other key partners.
3. *Integrated approach to disease control* – ensuring the rational use of available resources through the application of a multi-disease control approach; integration of non-chemical and chemical vector control methods; and integration with other disease control measures.

4. *Evidence-based decision-making* – adaptation of strategies and interventions to local vector ecology, epidemiology and resources, guided by operational research and subject to routine monitoring and evaluation.
5. *Capacity-building* – the development of essential infrastructure, financial resources and adequate human resources at national and local levels to manage IVM programmes, based on a situation analysis.

Prevention and management of insecticide resistance

The evolution and spread of resistance to insecticides is a major concern for the control of the dengue vector *Ae. aegypti*. The reliance by most dengue control programmes on just two classes of insecticide (pyrethroids and organophosphates) available for use in public health, poses additional selection pressure on the mosquito vectors (Ranson et al., 2010).

Alterations in the molecular target sites of insecticides, which reduce the binding of insecticides, are the most understood resistance mechanisms. Several mutations in the sodium channel, the target site of DDT and pyrethroid insecticides, have been reported in *Ae. aegypti* (Brengues et al., 2003). Two alternative substitutions at one of the polymorphic sites, residue 1 016, have been linked to pyrethroid resistance and recently, methodologies to detect these mutations (often referred to as kdr mutations) in individual mosquitoes have been reported (Saavedra-Rodríguez et al., 2007; Rajatileka et al., 2008).

Resistance management strategies generally recommend the rotation of chemicals with different modes of action and the use of non-chemical methods of control. The implicit assumption is that resistance to a chemical will disappear from a population once the selection pressure is removed. Effective IVM will be possible only through an important development of available biological control tools, to be combined with insecticide and physical control.

In order to successfully develop and implement any resistance management strategies based on rotations, mosaics, mixtures or combinations, knowledge of the mode of action, chemical properties and residual life of the available insecticide products is crucial. Focusing on surveillance wherever possible is essential in order to react proactively once a regional population manifests a shift in its susceptibility towards synthetic insecticides.

References

Ahmed, M.Z., J.W. Breinholt and A.Y. Kawahara (2016), "Evidence for common horizontal transmission of Wolbachia among butterflies and moths", *BMC Evolutionary Biology*, Vol.16, No. 1, pp. 118.

Alphey, L. (2014), "Genetic Control of Mosquitoes", *Annual Review of Entomology*, Vol. 59, pp. 205-224.

Baldo, L. et al. (2008), "Insight into the routes of Wolbachia invasion: high levels of horizontal transfer in the spider genus Agelenopsis revealed by *Wolbachia* strain and mitochondrial DNA diversity", *Molecular Ecology*, Vol. 17, pp. 557–569.

Bian, G. et al. (2010), "The endosymbiotic bacterium *Wolbachia* induces resistance to dengue virus in *Aedes aegypti*", *PLoS Pathogens*, Vol. 6: e1000833.

Bouyer, J. and T. Lefrançois (2014), "Boosting the sterile insect technique to control mosquitoes", *Trends in Parasitology*, Vol. 30, pp. 271-273.

Boyce, R. et al. (2013), "Bacillus thuringiensis israelensis (Bti) for the control of dengue vectors: Systematic literature review", *Tropical Medicine & International Health*, Vol. 18, No. 5, pp. 564-577.

Brelsfoard, C. et al. (2014), "Presence of extensive *Wolbachia* symbiont insertions discovered in the genome of its host Glossina morsitans morsitans", *PLoS Neglected Tropical Diseases*, Vol. 8, No. 4: e2728.

Brengues, C. et al. (2003), "Pyrethroid and DDT cross-resistance in *Aedes aegypti* is correlated with novel mutations in the voltage-gated sodium channel gene", *Medical and Veterinary Entomology*, Vol. 17, No. 1, pp. 87-94.

Carvalho, D.O. et al. (2015), "Suppression of a field population of *Aedes aegypti* in Brazil by sustained release of transgenic male mosquitoes", *PLoS Neglected Tropical Diseases*, Vol. 9, No. 7: e0003864.

CDC (2010), *Dengue, Entomology and Ecology website*, Centers for Disease Control and Prevention, U.S. Deptertment of Health and Human Services, Atlanta, United States, www.cdc.gov/dengue/entomologyEcology.

Choi, J.Y., J.E. Bubnell and C.F. Aquadro (2015), "Population genomics of infectious and integrated *Wolbachia pipientis* genomes in *Drosophila ananassae*", *Genome Biology and Evolution*, Vol. 7, No. 8, pp. 2362-2382.

Devine, G.J. et al. (2009), "Using adult mosquitoes to transfer insecticides to *Aedes aegypti* larval habitats", *Proceedings of the National Academy of Science. U.S.A.*, Vol. 106, pp. 11530-11534.

Dicko, A.H. et al. (2014), "Using species distribution models to optimize vector control: The tsetse eradication campaign in Senegal", *Proceedings of the National Academy of Sciences*, Vol. 111, pp. 10149-10154.

Dobson, S.L. et al. (1999), "*Wolbachia* infections are distributed throughout insect somatic and germ line tissues", *Insect Biochemistry and Molecular Biology*, Vol. 29, pp. 153-160.

Dunning-Hotopp, J.C. et al. (2007), "Widespread lateral gene transfer from intracellular bacteria to multicellular eukaryotes", *Science*, Vol. 317, No. 5845, pp. 1753-1756.

EFSA Panel on Genetically Modified Organisms (GMO) (2013), "Guidance on the environmental risk assessment of genetically modified animals", *EFSA Journal*, Vol. 11, No. 5.

Erlanger, T., L. Keiser and J. Utzinger (2008), "Effect of dengue vector control interventions on entomological parameters in developing countries: A systematic review and meta-analysis", *Medical and Veterinary Entomology*, Vol. 22, pp. 203-221.

Esu, E. et al. (2010), "Effectiveness of peridomestic space spraying with insecticide on dengue transmission; Systematic review", *Tropical Medicine and International Health*, Vol. 15, pp. 619-631.

Fenn, K. et al. (2006), "Phylogenetic relationships of the *Wolbachia* of nematodes and arthropods", *PLoS Pathogen*, Vol. 2: e94.

Foster, W.A. and E.D. Walker (2002), "Mosquitoes (Culicidae)", in G. Mullen and L. Durden (eds.), *Medical and Veterinary Entomology*, Academic Press, San Diego, pp. 203-262.

Funkhouser-Jones, L.J. et al. (2015), "*Wolbachia* co-infection in a hybrid zone: Discovery of horizontal gene transfers from two *Wolbachia* supergroups into an animal genome", *PeerJ*., Vol. 3: e1479.

Gorman, K. et al. (2016), "Short-term suppression of *Aedes aegypti* using genetic control does not facilitate *Aedes albopictus*", *Pest Management Sciences*, Vol. 72, No. 3, pp. 618–628.

Harris, A.F. et al. (2012), "Successful suppression of a field mosquito population by sustained release of engineered male mosquitoes", *Nature Biotechnology*, Vol. 30, pp. 828-830.

Hou, Q. et al. (2014), "A case of horizontal gene transfer from *Wolbachia* to *Aedes albopictus* C6/36 cell line", *Mobile Genetic Elements*, Vol. 4: e28914.

Kay, B.H. et al. (2012), "Control of *Aedes* vectors of dengue in three provinces of Vietnam by use of Mesocyclops (Copepoda) and community-based methods validated by entomologic, clinical, and serological surveillance", *The American Journal of Tropical Medicine and Hygiene*, Vol. 66, pp. 40-48.

Keroack, C.D. et al. (2016), "Absence of the filarial endosymbiont *Wolbachia* in seal heartworm (*Acanthocheilonema spirocauda*) but evidence of ancient lateral gene transfer", *Journal of Parasitol*ogy, Vol. 102, pp. 312–318.

Kittayapong, P. et al. (2000), "Distribution and diversity of *Wolbachia* infections in Southeast Asian mosquitoes (Diptera: Culicidae)", *Journal of Medical Entomology*, Vol. 37, pp. 340-345.

Klasson, L. et al. (2014), "Extensive duplication of the *Wolbachia* DNA in chromosome four of *Drosophila ananassae*", *BMC Genomics*, Vol. 15, No. 1: 1097.

Klasson, L. et al. (2009), "Horizontal gene transfer between *Wolbachia* and the mosquito *Aedes aegypti*", *BMC Genomics*, Vol. 10: 33.

Lacey, L.A. (2007), "*Bacillus thuringiensis* serovariety *israelensis* and *Bacillus sphaericus* for mosquito control", *Journal of the American Mosquito Control Association*, Vol. 23 (2 Suppl), pp. 133-163.

Mains, J.W., C.L. Brelsfoard and S.L. Dobson (2015), "Male mosquitoes as vehicles for insecticide", *PLoS Neglected Tropical Diseases*, Vol. 9: e0003406.

Manrique-Saide, P. et al. (2015), "Long-lasting insecticide treated screens significantly reduce indoor *Aedes aegypti* presence and abundance in Acapulco, Mexico", *Transactions of the Royal Society of Tropical Medicine and Hygiene*, Vol. 109, No. 2, pp. 106-115.

McCall, P.J. and P. Kittayapong (2006), "Control of dengue vectors: Tools and strategies", in *Report of the Scientific Working Group Meeting on Dengue*, World Health Organization, Geneva, WHO/TDR 2007, pp. 110-119.

McCall, P.J., L. Lloyd and M.B. Nathan (2009), *Vector Management and Delivery of Vector Control Services Chapter 3 in Dengue Guidelines for Diagnosis, Treatment, Prevention and Control 3rd Edition*, World Health Organization, Geneva.

McMeniman, C.J. et al. (2009), "Stable introduction of a life-shortening *Wolbachia* infection into the mosquito *Aedes aegypti*", *Science*, Vol. 323, pp. 141-144.

McMeniman, C.J. et al. (2008), "Host adaptation of a *Wolbachia* strain after long-term serial passage in mosquito cell lines", *Applied and Environmental Microbiology*, Vol. 74, pp. 6963-6969.

Mesquita, R.D. et al. (2015), "Genome of Rhodnius prolixus, an insect vector of Chagas disease, reveals unique adaptations to hematophagy and parasite infection", *Proceedings of the National Academy of Sciences of the United States of America*, Vol. 112, pp. 14936–14941.

Moreira, L.A. et al. (2009), "A *Wolbachia* symbiont in *Aedes aegypti* limits infection with dengue, Chikungunya, and Plasmodium", *Cell*, Vol. 139, pp. 1268-1278.

Morrow, J.L. et al. (2015), "*Wolbachia* pseudogenes and low prevalence infections in tropical but not temperate Australian tephritid fruit flies: Manifestations of lateral gene transfer and endosymbiont spillover?", *BMC Evolutionary Biology.*, Vol. 15, No. 202.

Nakao, R. et al. (2016), "Horizontally transferred genetic elements in the tsetse fly genome: An alignment-free clustering approach using batch learning self-organising approach using Batch Learning Self-Organising Map (BLSOM)", *BioMed Research International*, Vol. 2016: 3164624.

Nguyen, T.H. et al. (2015), "Field evaluation of the establishment potential of *w*MelPop *Wolbachia* in Australia and Vietnam for dengue control", *Parasites and Vectors*, Vol. 8, No. 1, pp. 1-14.

Nikoh, N. et al. (2008), "*Wolbachia* genome integrated in an insect chromosome: Evolution and fate of laterally transferred endosymbiont genes", *Genome Research*, Vol. 18, pp. 272–280.

PAHO (1994), *Dengue and Dengue Hemorrhagic Fever in the Americas: Guidelines for Prevention and Control*, PAHO Scientific Publication 548, Pan American Health Organization, Washington, DC.

Rajatileka, S. et al. (2008), "Development and application of a simple colorimetric assay reveals widespread distribution of sodium channel mutations in Thai populations of *Aedes aegypti*", *Acta Tropica*, Vol. 108, No. 1, pp. 54-57.

Ranson, H. et al. (2010), "Insecticide resistance in dengue vectors", *TropIKA.net*, Vol. 1, No.1.

Ricci, I. et al. (2002), "Searching for *Wolbachia* (Rickettsiales: Rickettsiaceae) in mosquitoes (Diptera: Culicidae): Large polymerase chain reaction survey and new identifications", *Journal of Medical Entomology*, Vol. 39, pp. 562-567.

Ritchie, S.A. and G.J. Devine (2013), "Confusion, knock-down and kill of *Aedes aegypti* using metofluthrin in domestic settings: A powerful tool to prevent dengue transmission?", *Parasites &Vectors.*, Vol. 6, pp. 1-9.

Rizzo, N. et al. (2012), "Dengue vector management using insecticide treated materials and targeted interventions on productive breeding-sites in Guatemala", *BMC Public Health*, Vol. 2, pp. 931.

Saavedra-Rodriguez, K. et al. (2007), "A mutation in the voltage-gated sodium channel gene associated with pyrethroid resistance in Latin American *Aedes aegypti*", *Insect Molecular Biology*, Vol. 16, No. 6, pp. 785-798.

Sebastian, A. et al. (1990), "Suppression of *Aedes aegypti* (Diptera: Culicidae) using augmentative release of dragonfly larvae (Odonata: Libellulidae) with community participation in Yangon, Myanmar", *Bulletin of Entomological. Research*, Vol. 80, pp. 223-232.

Sebastian, A. et al. (1980), "The use of dragonfly larvae in the control of *Aedes aegypti*", *Southeast Asian Journal of Ttropical Medicine and Public Health*, Vol. 11, No. 1, pp. 104-107.

Sinkins, S.P., H.R. Braig and S.L. O'Neill (1995), "*Wolbachia* superinfections and the expression of cytoplasmic incompatibility", *Proceedings. Biological Sciences*, Vol. 261, pp. 325-330.

Toribio-Fernández, R. et al. (2017), "Chromosomal localization of *Wolbachia* inserts in the genomes of two subspecies of *Chorthippus parallelus* forming a Pyrenean hybrid zone", *Chromosome Research*, Vol. 3-4, pp. 215-225.

Vanlerberghe, V. et al. (2011), "Evaluation of the effectiveness of insecticide treated materials for household level dengue vector control", *PLoS Neglected Tropical Diseases*, Vol. 5, No. 3: e994.

Vazquez-Prokopec, G.M. et al. (2017), "Combining contact tracing with targeted indoor residual spraying significantly reduces dengue transmission", *Science Advances*, No. 3, No. 2: e1602024.

Venkatesh, A. and B.K. Tyagi (2013), "Predatory potential of *Bradinopyga 118 ocalize* and *Ceriagrion coromandelianum* larvae ON dengue vector *Aedes aegypti* under controlled conditions (Anisoptera: Libellulidae; Zygoptera: Coenagrionidae; Diptera: Culicidae)", *Odonatologica*, Vol. 42, No. 2, pp. 139-149.

Vreysen, M.J.B. et al. (2014), "Sterile insects to enhance agricultural development: The case of sustainable tsetse eradication on Unguja Island, Zanzibar using an area-wide integrated pest management approach", *PLoS Neglected Tropical Diseases*, Vol. 8: e2857.

WHO (2016), *Report of the nineteenth WHOPES working group meeting: WHO/HQ, Geneva, 8-11 February 2016: review of Veeralin LN, VectoMax GR, Bactivec SC*, WHO Pesticide Evaluation Scheme, Control of Neglected Tropical Diseases, World Health Organization, Geneva, ISBN 978 92 4 151040 0, http://apps.who.int/iris/bitstream/handle/10665/205588/9789241510400_eng.pdf?sequence=1&isAllowed=y

WHO (2013), *Guidelines for Laboratory and Field Testing of Long-Lasting Insecticidal Nets*, World Health Organization, Geneva.

WHO (2009a), *WHO Recommended Insecticides for Indoor Residual Spraying Against Malaria Vectors*, World Health Organization, Geneva, www.who.int/whopes/Insecticides_IRS_Malaria_09.pdf.

WHO (2009b), *Dengue: Guidelines for Diagnosis, Treatment, Prevention and Control: New Edition*, World Health Organization, Geneva, www.ncbi.nlm.nih.gov/books/NBK143163/.

WHO (2008), *World malaria report 2008*, World Health Organization, Geneva.

WHO (2003), *Space Spray Application of Insecticides for Vector and Public Health Pest Control A Practitioner's Guide, Communicable Disease Control, Prevention and Eradication WHO Pesticide Evaluation Scheme (WHOPES)*, World Health Organization, Geneva, http://apps.who.int/iris/bitstream/handle/10665/68057/WHO_CDS_WHOPES_GCDPP_2003.5.pdf?sequence=1&isAllowed=y

WHO (1982), *Manual on Environmental Management for Mosquito Control with Special Emphasis on Malaria Vectors*, World Health Organization, Geneva.

WHO/EMRO (2003), *Use of Fish for Mosquito Control*, World Health Organization Regional Office for the Eastern Mediterranean, Cairo.

Yeap, H.L. et al. (2010), "Dynamics of the "popcorn" *Wolbachia* infection in outbred *Aedes aegypti* informs prospects for mosquito vector control", *Genetics*, Vol. 187, pp. 583-595.

Yen, J.H. and A.R. Barr (1973), "New hypothesis of the cause of cytoplasmic incompatibility in Culex pipiens L", *Nature*, Vol. 232, pp. 657-658.

Zhang, D. et al. (2016), "Combining the sterile insect technique with the incompatible insect technique: III-robust mating competitiveness of irradiated triple *Wolbachia*-infected *Aedes albopictus* males under semi-field conditions", *PloS ONE*, Vol. 11, No. 3: e0151864.

Annex B. **Human and animal health affected by mosquitoes**

This annex deals with the pathogens and diseases transmitted by mosquitoes to humans and animals. The main arbovirus infections of humans in the diverse regions of the world are summarised. More details are given on the dengue virus including its four viral serotypes, the range of symptoms affecting humans, the past and current epidemics of dengue (mainly vectored by Aedes aegypti *and other aedine species) including its increasing spread over the past fifteen years. Few elements are also provided on virus transmission to animals by* Ae. aegypti*, and on vertical transmission.*

Pathogens and diseases

An arthropod-borne virus or arbovirus is defined as a virus that is maintained in nature principally through biological transmission between susceptible vertebrate hosts by haematophagous arthropods; arboviruses multiply and produce virus in the vertebrate host, multiply in arthropod tissues, and are passed on after a period of extrinsic incubation to other vertebrates once again by the bites of an arthropod (PAHO, 1979). Most arboviruses fulfil the criteria laid down in this definition, but the group is very heterogeneous, containing viruses which, because they have not been fully classified on morphological or physicochemical grounds, are included among the arboviruses for convenience. There are currently 490 known arboviruses and this very large group contains representatives from several different viral families, the most important of which are the families Togaviridae, Flaviviridae, Bunyaviridae, Reoviridae and Rhabdoviridae (Bishop et al., 1980; Rehle, 1989).

The extrinsic incubation period (EIP) is the time necessary for the development of arbovirus in the arthropod host. If the female mosquito longevity is lower than the viral EIP, then the potential for vector transmission is reduced. Average EIP is 15 days at 25°C and 6.5 days at 30°C (Chan and Johansson, 2012).

By definition, arboviruses have at least two different hosts, a vertebrate and an invertebrate arthropod, although many arboviruses have complex life-cycles involving several different vertebrates, and some are capable of transmission by more than one species of vector. All arboviruses, with perhaps very few exceptions, are current or potential zoonoses maintained in nature principally by wild animals and birds.

They have evolved to a state of mutual tolerance or symbiosis with their reservoirs. Since arboviruses rely only on virus production in the vertebrate host for successful transmission, disease in this host would be a disadvantage. Therefore, arboviruses seldom cause recognisable disease in maintenance hosts, and when disease is apparent in man or domesticated animals, it is only overt sign of the presence of these viruses.

Over 80 viruses produce significant human disease which ranges from mild febrile illness, which may or may not be accompanied by a skin rash and sometimes by polyarthritis, to severe and often fatal encephalitis or haemorrhagic fever. The same virus may produce different disease patterns in different subjects and illness often has a biphasic pattern. Mild fever, often not recognised, occurs during the initial viraemic stage. This may be followed by more serious symptoms, at which stage viraemia may have ceased and immunological responses, including antibody formation, have occurred. Frequently, only a small proportion of persons infected with potentially encephalitogenic arboviruses in epidemics develop encephalitis in this second phase. The great majority of infections do not develop past the first phase, which may even be asymptomatic.

Virus infection vectored by mosquitoes

There are 66 members in the flavivirus group, of which 31 are mosquito-borne. 26 flaviviruses can cause human disease but several of them have produced only laboratory-acquired infections or isolated cases of disease in man (Table A B.1). The range of clinical manifestations produced by flaviviruses is similar to those of the alphaviruses – febrile illnesses with or without a rash, or encephalitis. In addition, yellow fever, Kyasanur Forest disease, Omsk haemorrhagic fever, and dengue virus can

cause haemorrhagic symptoms. Only those viruses which produce substantial prevalence are discussed in detail.

The International Committee on Taxonomy of Viruses (ICTV) has assigned the dengue virus (DENV) to the genus *Flavivirus*, of the Flaviviridae family. Based upon biological, immunological and molecular criteria, there are four viral serotypes, namely DENV-1, DENV-2, DENV-3 and DENV-4, which have different antigenic characteristics and serology (Boshell, 1995; Klungthong et al., 2004). Each serotype creates specific lifelong immunity against homologous reinfection, as well as short-term cross-immunity against the other serotypes, which can last several months (Leitmeyer et al., 1999; Monath, 2004). Each serotype has been subdivided into several genotypes (clades): three genotypes for DENV-1 (I, II and III) although two other clades named IV and V have been proposed, six genotypes for DENV-2 (American, Asian/American, Asian I, Asian II, Cosmopolitan and Sylvatic), four for DENV-3 (I, II, III and IV) although a fifth has also been proposed (V) and finally four for DENV-4 (I, II, III and Sylvatic) (Holmes, 2006).

Classic dengue fever affects both adults and older children. Following an infective mosquito bite, there is an incubation period of five to eight days followed by the sudden onset of acute fever, which often becomes biphasic, with a severe headache, pain behind the eyes, backache, chills and generalised pain in muscles and joints. A maculopapular rash generally appears on the thorax between the third and fifth day of illness and may spread later to the face and extremities. Lymphadenopathy, anorexia, constipation and altered taste sensation are common. Occasionally, petechiae are seen on the dorsal surfaces of the feet and the legs, hands, axillae and palate late in the illness. In young children, upper respiratory tract symptoms predominate and dengue fever is rarely suspected. The illness generally lasts for about ten days, after which recovery is usually complete, although convalescence may be prolonged. Laboratory findings reveal leukopenia, a mild thrombocytopenia, and slight lymphocytosis (Brathwaite et al., 2012).

Concerning the dengue haemorrhagic syndrome, fever, upper respiratory symptoms, headache, vomiting and abdominal pain may be present in the initial phase of the disease. Myalgia and arthralgia are uncommon. These symptoms (which are not severe enough for confinement) may last two to four days and many recover without any further symptoms. However, in a proportion of these cases, the initial phase is followed by an abrupt systemic collapse with hypotension, peripheral vascular congestion, petechiae, and sometimes a rash. Different degrees of shock may be evident, with the patient often restless, sweating, and febrile, clammy extremities, and a hot, feverish trunk. The fourth and fifth days are critical and purpura, ecchymoses, epistaxis, haematemesis, melaena, coma, convulsions and severe shock indicate a poor prognosis. Should the patient survive this period, however, recovery is usually complete. Laboratory studies often reveal thrombocytopenia, a prolonged bleeding time, an elevated prothrombin time, a raised haematocrit, hyperproteinemia and a positive tourniquet test. The liver is often enlarged, soft, and tender (Brathwaite et al., 2012). Several hypotheses have been proposed to explain why DENV now causes devastating epidemics, although it previously caused relatively mild illness. The two principal proposals are that either there is an unusual response to infection in the host, or there is an increase in the virus' virulence. Haemorrhagic manifestations are thought to be due to secondary infection with different DENV, with a critical interval of six months between the two infections. The first infection probably sensitises the patient, whereas the second appears to produce an immunological catastrophe (WHO, 2009).

Table A B.1. Some important arbovirus infections of humans in geographic regions of the world

				DISEASE FEATURES IN HUMANS			
Disease	**Geographic region(s)**	**Vectors**	**Vertebrate host(s)**	**Disease pattern**	**Description of diseases**	**Diagnosis**	**Control measure**
Yellow fever urban	New World and Africa	*Ae. aegypti*	Man	Epidemic	Acute onset, high fever, prostration, later jaundice, proteinuria; fatalities common, although ratio of inapparent/apparent infection is high	Virus isolation, CF, HI, N, ELISA test	Vaccination with 17D vaccine, *Ae. aegypti* control
Yellow fever jungle	New World and African tropics	Mosquitoes haemagogus and aedines	Forest primates	Endemic	As above; cases occur sporadically in people exposed in forested regions in Africa and New World	Virus isolation, CF, HI, N, ELISA test	Vaccination with 17D vaccine, mosquito control not practicable
Dengue	New World and Old World tropics and subtropics	*Ae. aegypti* and other aedines	Man, possibly a jungle cycle in primates	Endemic and epidemic	Acute onset with rash in many cases and joint pains; simulates as influenza-like syndrome	Virus isolation, CF, HI, N, ELISA test	Vaccination with Dengvaxia, under conditions[1] Mosquito control and protection against mosquito bites
Dengue haemorrhagic fever	Southeast Asia and South America	*Ae. aegypti*	Man	Endemic and epidemic	Serious illness with haemorrhagic complicates, shock syndrome and high mortality almost exclusively in children and following a second infection with a different DENV	CF, HI, N, ELISA test, cell-culture system	Mosquito control
Japanese encephalitis	Korea to India and East Indies	*Culex tritaeniorhynchus* and other culicines	Wild birds, pigs can serve as amplifying host	Endemic and epidemic	Infection usually mild but encephalitic complications can be serious in young and in elderly, very important disease in the Orient	CF, HI, N, ELISA test	Mosquito control, vaccination with an inactivated vaccine
Murry Valley encephalitis	Australia	*Culex annulirostris*	Birds	Endemic, sporadic, over wide areas	Infection usually mild but encephalitis may occur with greatest probability in children and high fatality rates in the young	CF, HI, N test	Mosquito control measures and protection against mosquito bite

				DISEASE FEATURES IN HUMANS			
Disease	**Geographic region(s)**	**Vectors**	**Vertebrate host(s)**	**Disease pattern**	**Description of diseases**	**Diagnosis**	**Control measure**
Chikungunya	Africa and Asia, tropics and subtropics Cases of autochtonous transmission in Europe	*Ae. aegypti*and *Ae. albopictus*	Possibly primates	Epidemic	Acute onset often with rash, rarely with haemorrhagic manifestations; joint aching and swelling are prominent features	CF, HI, N, ELISA test and virus isolation	Mosquito control
Kyasanur Forest disease	India (Mysore State)	Ticks mainly *Haemaphysalis*	Monkey, possibly also small mammals	Endemics and epidemic	Sudden onset, fever, headache, severe myalgia; there may be a diphase course with second phase	Virus isolation, CF, HI, N ELISA test	Protection against tick bite
Crimean-Congo haemorrhagic fever	Southern former USSR, Bulgaria, Central and South Africa, Pakistan, Iraq	Ticks - *Hyalomma marginatum*	Probably small mammals	Endemics	Sudden onset, chills, fever, headache, nausea, vomiting; haemorrhagic manifestations common; mortality rate 5%-10%	Virus isolation, CF test	Protection against tick bite
Venezuela equine encephalitis	Central and South America and southern United States	Mosquito of several species	Horses, possibly small mammals	Probably endemic, sharply epidemic	Fever, encephalitic signs, usually mild fatalities rate	Virus isolation, CF, HI, N, ELISA test	Mosquito control and protection against mosquito bites; attenuated vaccine exists for equines

Note: [1]WHO recommends that vaccine against dengue should only be used after testing on individuals to assess whether they have ever been exposed to the infection. (WHO Website, 2018)

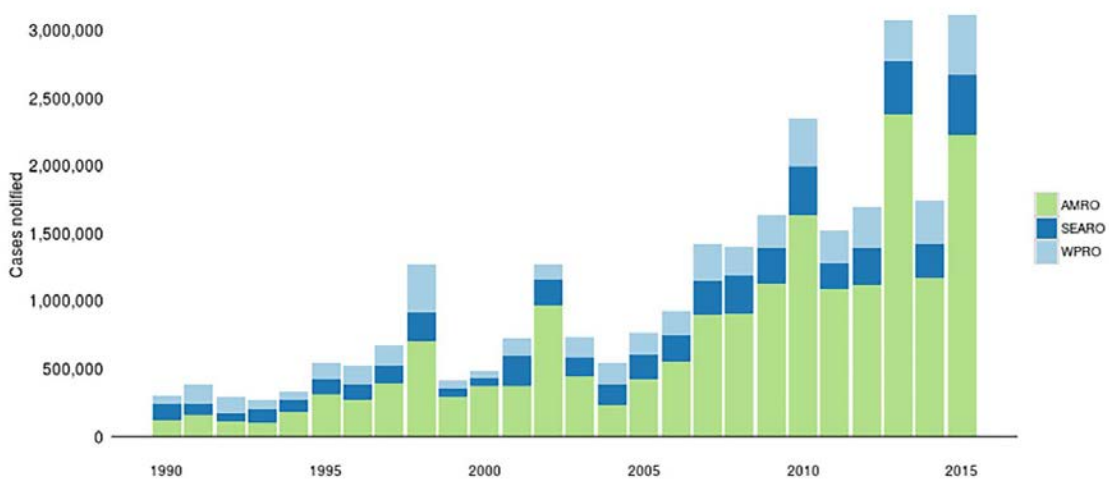
Source: Adapted from Evans, A.S. (Ed.) (1982), Viral Infections of Humans: Epidemiology and Control. Second Edition, Plenum Medical Book Company, New York and London.

Dengue virus serotypes, health effects and epidemics

The dengue disease may be endemic (which is often undiagnosed) or epidemic. In the Americas, there have been four epidemics during the 1963-83 period. The first epidemic in 1963 was caused by DENV-3 in the Caribbean and Venezuela. The second was in 1969, caused by DENV-2, affecting the Caribbean islands and also Colombia. The third epidemic began in 1977 in Jamaica, was caused by DENV-1 and affected more than 60 000 inhabitants, spreading to other Caribbean islands, Mexico, Central America, and Venezuela (Figueroa et al., 1982). In 1981 the fourth epidemic, resulting from DENV-4, began in Saint Barthélemy (French Antilles) and spread to other Caribbean Islands and Belize (PAHO, 2005).

Puerto Rico was seriously affected during all four epidemics, and after relatively high dengue activity in 1981 and 1982, the first epidemic of dengue in Brazil in 50 years began. Most countries reported only sporadic cases during 1983, however, Colombia, El Salvador and Mexico had significant localised outbreaks in 1983 (PAHO, 2005). *Ae. aegypti* reinfestation (1971-99) was caused by the failure of eradication programmes leading to increased dispersal of the mosquito and DENV circulation and a corresponding clear increase in the number of outbreaks over the 2000-10 period. During 2010, more than 1.7 million dengue cases were reported, with 50 235 severe cases and 1 185 deaths (Brathwaite et al., 2012). The epidemic seemed to continue extending globally in the following years; in 2015, the total number of suspected or laboratory-confirmed dengue cases notified to WHO for the Americas, South-East Asia and Western Pacific regions, exceeded three million (Figure A B.1).

Figure A B.1. Number of suspected or laboratory-confirmed dengue cases notified to WHO, 1990-2015



Note: a) AMRO: WHO Regional Office for the Americas
b) SEARO: WHO Regional Office for South-East Asia
c) WPRO: WHO Regional Office for the Western Pacific
Source: WHO (2018), Programmes – Dengue Control – Epidemiology Page, Website, www.who.int/denguecontrol/epidemiology/en/.

Zika virus infection

Zika virus (ZIKV) belongs to *Flavivirus* genus of the Flaviviridae family and it is transmitted to humans by mosquitoes (Gould and Solomon, 2008). However, sexual transmission between humans is another potential form of infection (Moreira et al., 2017). In 2015, ZIKV was shown to be associated with microcephaly and birth defects in children exposed *in utero* following infection of mothers during their pregnancy in Brazil (Zanluca et al., 2015; Calvet et al., 2016; Mlakar et al., 2016). Other studies evidenced the link between ZIKV infection during pregnancy and congenital cerebral malformations in newborns as microcephaly and other dysfunctions (Besnard et al., 2016; Driggers et al., 2016), and this was experimentally supported (Cugola et al., 2016). Moreover, the infection consequences in newborns can cause a range of different pathologies, which were described as the congenital Zika syndrome (Martines et al., 2016). ZIKV may additionally be associated with other neurological complications affecting adults, such as Guillain-Barré Syndrome (Dos Santos et al., 2016). Beyond to newborn disorders, the main symptoms of ZIKV infection are maculopapular rash, fatigue, lethargy, asthenia, fever, arthritis, arthralgia, myalgia, conjunctivitis and headache. The suspected patients can be submitted to RT-PCR assays or serological tests to confirm the ZIKV infection (Musso and Gubler, 2016).

The recent burden of Zika virus outbreaks in many countries is alarming. Although the virus is known since 1947 when it was first isolated from a sentinel *Rhesus* monkey exposed in the Zika Forest (Uganda) (Dick, Kitchen and Haddow, 1952), fewer reports of human infections were described until 2007. In that year, a ZIKV outbreak was first registered in Yap Islands, Federated States of Micronesia and since then, subsequent epidemics were reported in several islands in different Pacific regions between 2013 and 2014. This fast geographic expansion of the viral distribution was achieved in the Americas in 2015, causing important epidemics, mainly in Brazil. Currently, autochthonous transmission of ZIKV is occurring in many countries around the world where potential mosquito vectors are endemic (Musso and Gubler, 2016). The ZIKV emergent scenario caught the attention of the main health authorities mainly because congenital microcephaly and other neurological disorders in newborns were correlated with ZIKV infection in pregnant women, as described above. The WHO declared a state of public health emergency of international concern during almost the entire year of 2016 and launched a document named "Zika Strategic Response Plan" to guide the viral prevention and management by the national governments and communities where activities related to detection, prevention, research, care and support were recommended. This strategical document is constantly updated to provide the key information and progress achieved against ZIKV infections (WHO, 2016).

Entomological studies have demonstrated that Brazilian and other American populations of *Ae. aegypti* and *Ae. albopictus* mosquitoes are competent to ZIKV, but they present different levels of susceptibility (Chouin-Carneiro et al., 2016). Moreover, well-known laboratory strains of *Ae. aegypti* also show vector competence to this pathogen, which can sustain vector-pathogen studies to clarify the interactions between this virus and its invertebrate host (Costa-da-Silva et al., 2017). Recently, a field study demonstrated the occurrence of naturally-infected *Ae. aegypti* in the city of Rio de Janeiro (Brazil), confirming the species potential to transmit ZIKV to humans (Ferreira-de-Brito et al., 2016). The entomological surveillance in endemic regions is an essential activity to monitor the circulation of ZIKV and the potential of new outbreaks to occur.

Ae. Aegypti other characteristics

Transmission to animals

In addition to being a vector for human pathogens, *Ae. aegypti* is capable of spreading disease among animal species that associate with humans, such as cattle and dogs. *Ae. aegypti* female mosquitoes are capable of the mechanical transmission of lumpy skin disease virus (LSDV) from infected to susceptible cattle (Chihota et al., 2001). Canine heartworm is transmitted by *Ae. aegypti* to dogs, which are companion animals frequently associated with the home environment.

Vertical transmission

The virus is transmitted to humans through the bite of the mosquito *Ae. aegypti* as principal vector and *Ae. albopictus* as a secondary vector. The mechanism of transmission of the virus that occurs most commonly involves the human-to-mosquito-to-human cycle.

However, it has been observed that vertical transmission of the virus can occur whereby infected females naturally transmit the virus to their progeny (transovarial transmission), the virus being in this case transmitted to the next generation without an intervening human host. Vertical transmission allows the virus to persist in nature during adverse weather conditions that limit mosquito reproduction, resulting in the appearance of virus-infected mosquitos once desiccated eggs hatch following a subsequent rainfall. Thus, vertical transmissions in vectors could play a role in the endemic maintenance of the viruses. Vertical transmission of dengue viruses in *Ae. aegypti* is documented by several studies (see below) and appears to vary with the vector geographical strains and virus serotypes (Rodhain and Rosen, 1997).

The first findings suggesting that transovarial transmission of DENV can occur in nature was reported by Khin and Than (1983). In this study, DENV-2 serotype was recovered from three of 123 pools of *Ae. aegypti* larvae (6 200 specimen) collected from water containers in Rangoon, Myanmar; the virus was also isolated from two of the 76 pools (7 730 mosquitoes) of male *Ae. aegypti* collected as larvae and reared in the laboratory to adults. In Trinidad and Tobago, the isolation of DENV-4 from adult *Ae. aegypti* reared from eggs and larvae collected in nature was documented by Hull et al. (1984): the virus was recovered in one out of the 158 mosquito pools tested from 10 different localities (10 957 adults processed for virus isolation), giving further evidence that transovarial transmission of DENV occurs in nature. In southern India, DENV-2 and DENV-3 were detected in vertical transmission to males in summer months when dengue infections were high in humans, suggesting how DENV adopted a novel strategy of surviving adverse climatic conditions (Thenmozhi et al., 2000). In Juchitán and Tuxtepec, Oaxaca, Mexico, vertical transmission of DENV in *Ae. aegypti* mosquitoes was recorded in two endemic localities. Although the presence of DENV in larvae could not be demonstrated, DENV- 2, - 3 and -4 serotypes were detected in four out of 43 pools of in-cage born mosquitoes (Günther et al., 2007). In Acapulco, Guerrero, only two (0.9%) of 226 pools of *Ae. aegypti* adults (one pool of adults emerged from field-collected larvae, and another of indoor-collected adults) were positive for DENV-1. This appears to be the first report of evidence on the vertical and transovarial transmission of DENV-1 in field-caught *Ae. aegypti* in Mexico (Martínez et al., 2014).

References

Alfred, S.E. and J.R. Paul (1984), *Viral Infections of Humans: Epidemiology and Control. Second Edition*, Plenum Medical Book Company, New York and London.

Besnard, M. et al. (2016), "Congenital cerebral malformations and dysfunction in fetuses and newborns following the 2013 to 2014 Zika virus epidemic in French Polynesia", *Euro Surveillance*, Vol. 21, No. 13.

Bishop, D.H. et al. (1980), "Bunyaviridae", *Intervirology*, Vol. 14, No. 3-4, pp. 125-143.

Boshell, J. (1995), "El dengue" [Dengue], *Innovación y Ciencia* [*Innovation and Science*], Vol. 4, pp. 46-50.

Brathwaite, D.O. et al. (2012), "The history of dengue outbreaks in the Americas", *the American Journal of Tropical Medicine and Hygiene*, Vol. 87, No. 4, pp. 584-593.

Calvet, G. et al. (2016), "Detection and sequencing of Zika virus from amniotic fluid of fetuses with microcephaly in Brazil: a case study", *Lancet Infectious Diseases*, Vol. 16, No. 6, pp. 653-660.

Chan, M. and M.A. Johansson (2012), "The incubation periods of dengue viruses", *PLoS ONE*, Vol. 7, No. 11: e50972.

Chihota, C.M. et al. (2001), "Mechanical transmission of lumpy skin disease virus by *Aedes aegypti* (Diptera: Culicidae)", *Epidemiology & Infectection.*, Vol. 126, No. 2, pp. 317-321.

Chouin-Carneiro, T. et al. (2016), "Differential susceptibilities of *Aedes aegypti* and *Aedes albopictus* from the Americas to Zika virus", *PLoS Neglected Tropical Diseases*, Vol. 10, No. 3: e0004543.

Costa-da-Silva, A.L. et al. (2017), "Laboratory strains of *Aedes aegypti* are competent to Brazilian Zika virus", *PLoS One.*, Vol. 12, No. 2: e0171951.

Cugola, F.R. et al. (2016), "The Brazilian Zika virus strain causes birth defects in experimental models", *Nature*, Vol. 534, No. 7606, pp. 267-271.

Dick, G.W., S.F. Kitchen and A.J. Haddow **(**1952), "Zika virus. I. Isolations and serological specificity", *Transactions of the Royal Society of Tropical Medicine and Hygiene*, Vol. 46, pp. 509–520.

Dos Santos, T. et al. (2016), "Zika Virus and the Guillain-Barré Syndrome - Case Series from Seven Countries", *New England Journal of Medicine*, Vol. 375, No. 16, pp. 1598-1601.

Driggers, R.W. et al. (2016), "Zika virus infection with prolonged maternal viremia and fetal brain abnormalities", *New England Journal of Medicine*, Vol. 374, No. 22, pp. 2142-2151.

Ferreira-de-Brito, A. et al. (2016), "First detection of natural infection of *Aedes aegypti* with Zika virus in Brazil and throughout South America", *Memórias do Instituto Oswaldo Cruz*, Vol. 111, No. 10, pp. 655-658.

Figueroa, R.M. et al. (1982), "Dengue epidemic in Honduras, 1978-1980", *Bulletin of the Pan American Health Organization*, Vol. 16, No. 2, pp. 130-137.

Gould, E.A. and T. Solomon (2008), "Pathogenic flaviviruses", *Lancet Infectious Diseases*, Vol. 371, No. 9611, pp. 500-509.

Günther, J. et al. (2007), "Evidence of vertical transmission of dengue virus in two endemic localities in the state of Oaxaca, Mexico", *Intervirology*, Vol. 50, pp. 347-352.

Holmes, E.C. (2006), "The evolutionary biology of dengue virus", *Novartis Found Symposium*, Vol. 277, pp. 177-187.

Hull, B. et al. (1984), "Natural transovarial transmission of dengue 4 virus in *Aedes aegypti* in Trinidad", *the American Journal of Tropical Medicine and Hygiene*, Vol. 33, pp. 1248-1250.

Khin, M.M. and K.A. Than (1983), "Transovarial transmission of dengue 2 virus by *Aedes aegypti* in nature", *The American Journal of Tropical Medicine and Hygiene*, Vol. 32, pp. 590-594.

Klungthong, C. et al. (2004), "The molecular epidemiology of dengue virus serotype 4 in Bangkok, Thailand", *Virology*, Vol. 329, pp. 168-179.

Leitmeyer, K.C. et al. (1999), "Dengue virus structural differences that correlate with pathogenesis", *Virology*, Vol. 73, No. 6, pp. 4738-4747.

Martines, R.B. et al. (2016), "Pathology of congenital Zika syndrome in Brazil: A case series", *Lancet*, Vol. 388, No. 10047, pp. 898-904.

Martínez, N.E. et al. (2014), "Natural vertical transmission of dengue-1 virus in *Aedes aegypti* populations in Acapulco, Mexico", *Journal of the American Mosquito Control Association*, Vol. 30, No. 2, pp. 143-146.

Mlakar, J. et al. (2016), "Zika virus associated with microcephaly", *New England Journal of Medicine*, Vol. 374, pp. 951–958.

Monath, T. (2004), "Dengue: The risk to developed and developing countries", *Proceedings of the National Academy of Sciences of the United States of America*, Vol. 91, pp. 2395-2400.

Moreira, J. et al. (2017), "Sexually acquired Zika virus: A systematic review", *Clinical Microbiology and Infection*, Vol. 23, No. 5, pp. 296-305.

Musso, D. and D.J. Gubler (2016), "Zika virus", *Clinical Microbiology and Infection*, Vol. 29, No. 3, pp. 487-524.

PAHO (2005), *Number of Reported Cases of Dengue and Dengue Hemorrhagic Fever (Dhf), Region of the Americas (by Country and Subregion)*, Pan American Health Organization, Washington, DC.

PAHO (1979), *Dengue in the Carribean, 1977*, Proceedings of a Workshop Held in Montego Bay, Jamaica, 8-11 May, 1978, PAHO Scientific Publication 375, Pan American Health Organization, Washington, DC.

Rehle, T.M. (1989), "Classification, distribution and importance of arboviruses", *Tropical Medicine and Parasitology*, Vol. 40, No. 4, pp. 391-395.

Rodhain, F. and L. Rosen (1997), "Mosquito vectors and dengue virus vector relationships", in Gubler, D.J. and G. Kuno (eds), Dengue and Dengue Hemorrhagic Fever, CAB International, New York, pp. 45–60.

Thenmozhi, V. et al. (2000), "Natural vertical transmission of dengue viruses in *Aedes aegypti* in southern India", *Transactions of the Royal Society of Tropical Medicine and Hygiene*, Vol. 94, pp. 507.

WHO (2018), Programmes – Dengue Control – Epidemiology Page, Website, World Health Organization, Geneva, www.who.int/denguecontrol/epidemiology/en/.

WHO (2016), *Zika Strategic Response Framework and Joint Operations Plan, Zika Strategic Response Plan Quarterly Update 2016*, World Health Organization, Geneva, www.who.int/emergencies/zika-virus/quarterly-update-october/en/.

WHO (2009), *Dengue: Guidelines for Diagnosis, Treatment, Prevention and Control: New Edition*, World Health Organization, Geneva, www.ncbi.nlm.nih.gov/books/NBK143163/.

Zanluca, C. et al. (2015), "First report of autochthonous transmission of Zika virus in Brazil", *Memórias do Instituto Oswaldo Cruz*, Vol. 110, pp. 569–572.

List of OECD consensus documents on environmental safety assessment, 1996-2018

	Consensus document	Lead country(ies)	Year of issue	Volume
Facilitating harmonisation	Designation of a Unique Identifier for Transgenic Plants (revised version) (guidance document)	Working Group	2006	Vol. 3
	Introduction to the OECD Biosafety Consensus Documents	Working Group	2005	Vol. 1, 3, 4, 5, 6, 7, 8
	Low-Level Presence of Transgenic Plants in Seed and Grain Commodities: Environmental Risk/Safety Assessment, and Availability and Use of Information	Working Group	2013	Vol. 6
	Molecular Characterisation of Plants Derived from Modern Biotechnology	Canada	2010	Vol. 3
	Points to Consider for Consensus Documents on Biology of Cultivated Plants	Working Group	2006	Vol. 3
Traits	Crop Plants Made Virus Resistant through Coat Protein Gene-Mediated Protection	Task Group	1996	Vol. 1
	Genes and their Enzymes that Confer Tolerance to Glyphosate Herbicide	United States, Germany and Netherlands	1999	Vol. 1
	Genes and their Enzymes that Confer Tolerance to Phosphinothricin Herbicide	United States, Germany and Netherlands	1999	Vol. 1
	Herbicide Metabolism and the Residues in Glufosinate Ammonium (Phosphinothricin)-Tolerant Transgenic Plants	Germany	2002	Vol. 1
	Transgenic Plants Expressing Bacillus thuringiensis Derived Insect Control Protein	United States	2007	Vol. 3
Micro-organisms	***Information Used in the Assessment of Environmental Applications of Micro-organisms***			
	Acidithiobacillus	Canada	2006	Vol. 2
	Acinetobacter	Canada	2008	Vol. 4
	Baculovirus	Germany	2002	Vol. 2
	Pseudomonas	United Kingdom	1997	Vol. 2
	Guidance Documents on Biosafety Aspects of Bacteria			
	Horizontal Gene Transfer Between Bacteria	Germany	2010	Vol. 4
	Methods for Detection of Micro-organisms Introduced into the Environment: Bacteria	Netherlands	2004	Vol. 4
	Use of Information on Pathogenicity Factors: Bacteria	Netherlands and Canada	2011	Vol. 5
	Use of Taxonomy in Risk Assessment of Micro-organisms: Bacteria	Canada and United States	2003	Vol. 4

Biology of crops	Bananas and plantains (*Musa* spp.)	Spain	2009	Vol. 4
	Brassica crops (*Brassica* spp.)	Canada	2012	Vol. 5
	Cassava (*Manihot esculenta*)	Brazil, NEPAD ABNE and ILSI CERA	2014	Vol. 6
	Chili, hot and sweet peppers (*Capsicum annuum*)	Korea, Mexico and United States	2006	Vol. 1
	Common bean (*Phaseolus vulgaris*)	Brazil and ILSI CERA	2015	Vol. 6
	Cotton (*Gossypium* spp.)	Spain	2008	Vol. 4
	Cowpea (*Vigna unguiculata*)	Australia	2015	Vol. 6
	Maize (*Zea mays* subs. *mays*)	Mexico	2003	Vol. 1
	Squashes, pumpkins, zucchinis and gourds (*Cucurbita*)	Mexico and United States	2012	Vol. 5
	Oyster mushroom (*Pleurotus* spp.)	Korea	2005	Vol. 1
	Papaya (*Carica papaya*)	United States	2005	Vol. 1
	Potato (*Solanum tuberosum* subsp. *tuberosum*)	Netherlands and United Kingdom	1997	Vol. 1
	Rice (*Oryza sativa*)	Japan	1999	Vol. 1
	Oilseed rape (*Brassica napus*) [replaced with *Brassica* Crops (2012) in Vol. 5]	Canada	1997	Vol. 1
	Sugar beet (*Beta vulgaris*)	Switzerland	2001	Vol. 1
	Sugarcane (*Saccharum* spp.)	Australia	2013	Vol. 6
	Sunflower (*Helianthus annus*)	France	2004	Vol. 1
	Sorghum (*Sorghum bicolor*)	South Africa and United States	2016	Vol. 7
	Soybean (*Glycine max*)	Canada	2000	Vol. 1
	Tomato (*Solanum lycopersicum*)	Spain and Mexico	2016	Vol. 7
	Wheat (*Triticum aestivum*)	Germany	1999	Vol. 1
Biology of trees	***Timber trees***			
	Birch: European white birch (*Betula pendula*)	Finland	2003	Vol. 2
	Douglas fir (*Pseudotsuga menziesii*)	Canada	2008	Vol. 3
	Eucalyptus (*Eucalyptus* spp.)	Australia	2014	Vol. 6
	Larches: North American larches (*Larix lyalli*, *Larix occidentalis*, *Larix laricina*)	Canada	2007	Vol. 3
	Pines: Eastern white pine (Pinus strobus)	Canada	2002	Vol. 2
	Pines: Jack pine (*Pinus banksiana*)	Canada	2006	Vol. 3
	Pines: Lodgepole pine (*Pinus contorta*)	Canada	2008	Vol. 3
	Pines: White pine (*Pinus monticola*)	Canada	2008	Vol. 3
	Poplars (*Populus* spp.)	Canada	2000	Vol. 2

	Spruces: Black spruce (*Picea mariana*)	Canada	2010	Vol. 3
	Spruces: Norway spruce (*Picea abies*)	Norway	1999	Vol. 2
	Spruces: Sitka spruce (*Picea sitchensis*)	Canada	2002	Vol. 2
	Spruces: White spruce (*Picea glauca*)	Canada	1999	Vol. 2
	Fruit trees			
	Bananas and plantains (*Musa* spp.) [listed above in "Biology of crops"]	Spain	2009	Vol. 4
	Papaya (*Carica papaya*) [listed above in "Biology of crops"]	United States	2005	Vol. 1
	Stone fruits (*Prunus* spp.)	Austria	2002	Vol. 2
Biology of animals	Atlantic salmon (*Salmo salar*)	Finland, Norway and United States	2017	Vol. 7
	Mosquito *Aedes aegypti*	Mexico, Brazil and ILSI RF	2018	Vol. 8

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Harmonisation of Regulatory Oversight in Biotechnology

Safety Assessment of Transgenic Organisms in the Environment, Volume 8

OECD CONSENSUS DOCUMENT OF THE BIOLOGY OF MOSQUITO *AEDES AEGYPTI*

This Series represents a compilation of the science-based consensus documents developed by the OECD Working Group on the Harmonisation of Regulatory Oversight in Biotechnology since 1995. The consensus documents are prepared by authorities of OECD members and other economies associated with this work. They contain information for use during the regulatory assessment of organisms produced from modern biotechnology – transgenic crops, trees, animals and micro-organisms – intended for release in the environment for agriculture, animal farming, forestry or other purposes. Information relevant to environmental risk assessment (biosafety) includes biology, reproduction, genetics, ecology, and other elements. Knowledge of the traits introduced in the organisms, and their biotechnological developments, is also critical. These documents are of value to applicants for commercial uses of transgenic organisms, to regulators and risk assessors in charge of granting approvals to their environmental release, as well as to the wider scientific community. More information on this OECD series can be found at BioTrack Online: *www.oecd.org/biotrack.*

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Consult this publication on line at ***http://dx.doi.org/10.1787/9789264302235-en***.

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