



OECD GUIDELINE FOR TESTING OF CHEMICALS

Adopted by the Council on 30th March 1989

Partition Coefficient (n-octanol/water), High Performance Liquid Chromatography (HPLC) Method

Introduction

1. The partition coefficient (P) is defined as the ratio of the equilibrium concentrations of a dissolved substance in a two-phase system consisting of two largely immiscible solvents. In the case of n-octanol and water,

$$P_{ow} = \frac{C_{n-octanol}}{C_{water}}$$

The partition coefficient being the quotient of two concentrations, is dimensionless and is usually given in the form of its logarithm to base ten.

2. P_{ow} is a key parameter in studies of the environmental fate of chemical substances. A highly-significant relationship between the P_{ow} of substances and their bioaccumulation in fish has been shown. It has also been shown that P_{ow} is a useful parameter in the prediction of adsorption on soil and sediments and for establishing quantitative structure-activity relationships for a wide range of biological effects.

3. P_{ow} values in the range $\log P_{ow}$ between -2 and 4 can be experimentally determined by the Shake-Flask method (OECD Test Guideline 107). P_{ow} values in the range $\log P_{ow}$ between 0 and 6 can be estimated using high performance liquid chromatography (HPLC) (1) (2) (3) (4). The HPLC method requires a preliminary estimation of P_{ow} , generally done through calculation. Calculation methods are briefly discussed in the Annex to this guideline.

4. The original proposal for this guideline was based on an article by C.V. Eadsforth and P. Moser (1). The development of the guideline and an OECD inter-laboratory comparison test were coordinated by the Umweltbundesamt of the Federal Republic of Germany during 1986 (5).

Principle of the method

5. HPLC is performed on analytical columns packed with a commercially available solid phase containing long hydrocarbon chains (e.g. C_8 , C_{18}) chemically bound onto silica.

Chemicals injected onto such a column move along it by partitioning between the mobile solvent phase and the hydrocarbon stationary phase. The chemicals are retained in proportion to their hydrocarbon-water partition coefficient, with water-soluble chemicals eluted first and oil-soluble chemicals last. This enables the relationship between the retention time on a reverse-phase column and the n-octanol/water partition coefficient to be established. The partition coefficient is deduced from the capacity factor k , given by the expression

$$k = \frac{t_R - t_0}{t_0}$$

where, t_R is the retention time of the test substance, and t_0 is the dead-time, i.e. the average time a solvent molecule needs to pass the column. Quantitative analytical methods are not required and only the determination of retention times is necessary. If standard reference compounds are available and standard experimental conditions are used, the HPLC method can be performed faster than the Shake-Flask method.

6. The HPLC method enables partition coefficients to be estimated in the $\log P_{ow}$ range between 0 and 6. The method is not applicable to strong acids and bases, metal complexes, substances which react with the eluent, or surface-active agents. Measurements should be made on ionisable substances in their non-ionised form (free acid or free base) only by using an appropriate buffer with a pH below the pK for a free acid or above the pK for a free base (e.g. phosphoric acid for pH = 2 and 0.01 - 0.02 M phosphate buffer for pH = 7.5).

7. The HPLC method is less sensitive to the presence of impurities in the test substance than the Shake-Flask method. Nevertheless, in some cases impurities can make the interpretation of the results difficult due to uncertainty in peak assignments. For mixtures which result in an unresolved band, upper and lower limits of $\log P$ should be stated (3).

Information on the test substance

8. The structural formula and the dissociation constant should be known before using the method. Information on solubility and hydrolysis characteristics is useful.

Repeatability and accuracy

9. In order to increase the confidence in the measurement, duplicate determinations must be made. The values of $\log P_{ow}$ derived from the different measurements should fall within a range of ± 0.1 log units.

10. The inter-laboratory comparison test has shown that with the HPLC method $\log P_{ow}$ values can be obtained to within ± 0.5 units of the Shake-Flask values (5). Other comparisons can be found in the literature (3) (4) (6) (7) (8). Correlation graphs based on structurally related reference compounds give the most accurate results (9).

Reference compounds

11. In order to correlate the measured capacity factor k of a compound with its P_{ow} , a calibration graph using at least 6 points has to be established. It is up to the user to select the appropriate reference compounds. It is preferable that these should be structurally related to the test substance. Whenever possible, at least one reference compound should have a P_{ow} above that of the test substance, and another a P_{ow} below that of the test substance. For $\log P_{ow}$ values below 4, the calibration can be based on data obtained by the Shake Flask method. For $\log P_{ow}$ values above 4, the calibration can be based on literature values if they correspond to calculated values.

12. Extensive lists of $\log P_{ow}$ values for many groups of chemicals are available (10) (11). If data on the partition coefficients of structurally related compounds are not available, a more general calibration, established with other reference compounds, may be used. Recommended reference compounds and their P_{ow} values are listed in Table 1. For ionisable substances the values given apply to the non-ionised form. The values were checked for plausibility and quality during the inter-laboratory comparison test.

Description of the method

Preliminary estimate of the partition coefficient

13. The partition coefficient of the test substance is estimated preferably by using a calculation method (see Annex), or where appropriate, by using the ratio of the solubilities of the test substance in the pure solvents (12).

Table 1: Recommended Reference Compounds

Reference substance	$\log P_{ow}$	pKa
2-Butanone	0,3	
4-Acetylpyridine	0,5	
Aniline	0,9	
Acetanilide	1,0	
Benzil alcohol	1,1	
4-Methoxyphenol	1,3	pKa = 10,26
Phenoxyacetic acid	1,4	pKa = 3,12
Phenol	1,5	pKa = 9,92
2,4-Dinitrophenol	1,5	pKa = 3,96
Benzonitrile	1,6	
Phenylacetoneitrile	1,6	
4-Methylbenzyl alcohol	1,6	
Acetophenone	1,7	
2-Nitrophenol	1,8	pKa = 7,17
3-Nitrobenzoic acid	1,8	pKa = 3,47
4-Chloraniline	1,8	pKa = 4,15
Nitrobenzene	1,9	
Cinnamic alcohol	1,9	
Benzoic acid	1,9	pKa = 4,19
p-Cresol	1,9	pKa = 10,17

**Table 1: Recommended Reference Compounds
(Continued)**

Reference substance	log P _{ow}	pKa
Cinnamic acid	2,1	pKa = 3,89 cis 4,44 trans
Anisole	2,1	
Methyl benzoate	2,1	
Benzene	2,1	
3-Methylbenzoic acid	2,4	pKa = 4,27
4-Chlorophenol	2,4	pKa = 9,1
Trichloroethene	2,4	
Atrazine	2,6	
Ethyl benzoate	2,6	
2,6-Dichlorobenzonitrile	2,6	
3-Chlorobenzoic acid	2,7	pKa = 3,82
Toluene	2,7	
1-Naphthol	2,7	pKa = 9,34
2,3-Dichloroaniline	2,8	
Chlorobenzene	2,8	
Allyl phenyl ether	2,9	
Bromobenzene	3,0	
Ethylbenzene	3,2	
Benzophenone	3,2	
4-Phenyl phenol	3,2	pKa = 9,54
Thymol	3,3	
1,4-Dichlorobenzene	3,4	
Diphenylamine	3,4	pKa = 0,79
Naphthalene	3,6	
Phenyl benzoate	3,6	
Isopropylbenzene	3,7	
2,4,6-Trichlorophenol	3,7	pKa = 6
Biphenyl	4,0	
Benzyl benzoate	4,0	
2,4-Dinitro-6 sec. butyl phenol	4,1	
1,2,4-Trichlorobenzene	4,2	
Dodecanoic acid	4,2	
Diphenyl ether	4,2	
Phenanthrene	4,5	
n-Butylbenzene	4,6	
Fluoranthene	4,7	
Dibenzyl	4,8	
2,6-Diphenylpyridine	4,9	
Triphenylamine	5,7	
DDT	6,2	

Apparatus

14. A liquid-phase chromatograph, fitted with a pulse-free pump and a suitable detection device is required. The use of an injection valve with injection loops is recommended. The

presence of polar groups in the stationary phase may seriously impair the performance of the HPLC column. Therefore, stationary phases should have a minimal percentage of polar groups (13). Commercial microparticulate reverse-phase packings or ready-packed columns can be used. A guard column may be positioned between the injection system and the analytical column.

Mobile phase

15. HPLC-grade methanol and distilled water are used to prepare the eluting solvent, which is degassed before use. Isocratic elution should be employed. Methanol/water ratios with a minimum water content of 25% should be used. Typically a 3:1 (v/v) methanol-water mixture is satisfactory for eluting compounds with a log P of 6 within an hour, at a flow rate of 1 ml/min. For compounds with a log P above 6 it may be necessary to shorten the elution time (and those of the reference compounds) by decreasing the polarity of the mobile phase or the column length.

16. The test substance and the reference compounds should be soluble in the mobile phase in sufficient concentration to allow their detection. Additives may be used with the methanol-water mixture in exceptional cases only, since they will change the properties of the column. In these cases a separate column of the same type should be used. If methanol-water is not appropriate, other organic solvent-water mixtures can be used, e.g. ethanol-water, and acetonitrile-water.

17. The pH of the eluent is critical for ionisable compounds. It should be within the operating pH range of the column, usually between 2 and 8. Buffering is recommended. Care must be taken to avoid salt precipitation and column deterioration which occur with some organic phase/buffer mixtures. HPLC measurements with silica-based stationary phases above pH 8 are not advisable since the use of an alkaline mobile phase may cause rapid deterioration in the performance of the column.

Solutes

18. The test and reference compounds should be the purest available. Compounds to be used for test or calibration purposes are dissolved in the mobile phase if possible.

Test conditions

19. The temperature during the measurements should not vary by more than ± 2 K.

Determination of dead time t_0

20. The dead time t_0 can be measured by using unretained organic compounds (e.g. thiourea or formamide). It can also be derived from the retention times measured for a set of approximately seven members of a homologous series (e.g. n-alkyl methyl ketones) (14). The retention times $t_R(n_c+1)$ are plotted against $t_R(n_c)$, where n_c is the number of carbon atoms. A straight line, $t_R(n_c+1) = A t_R(n_c) + (1-A)t_0$, is obtained, where A, representing $k(n_c+1)/k(n_c)$, is constant. The dead time t_0 is obtained from the intercept $(1-A)t_0$ and the slope A.

Calibration graph

21. The next step is to plot a correlation graph of $\log k$ versus $\log P$ for appropriate reference compounds with $\log P$ values near the value expected for the test substance. In practice, from 5 to 10 reference compounds are injected simultaneously. The retention times are determined, preferably on a recording integrator linked to the detection system. The corresponding logarithms of the capacity factors, $\log k$, are calculated and plotted as a function of $\log P$. The calibration is performed at regular intervals, at least once daily, so that account can be taken of possible changes in column performance.

Determination of the P_{ow} of the test substance

22. The test substance is injected in the smallest quantity possible. The retention time is determined in duplicate. The partition coefficient of the test substance is obtained by interpolation of the calculated capacity factor on the calibration graph. For very low and very high partition coefficients extrapolation is necessary. Especially in these cases attention must be given to the confidence limits of the regression line.

Report

23. The following should be included in the report:

- test and reference substances, and their purity;
- description of equipment and operating conditions: analytical column, guard column,
- mobile phase, means of detection, temperature range, pH;
- elution profiles;
- deadtime and how it was measured;
- quantities of test and reference substances introduced in the column;
- retention data and literature $\log P$ values for reference compounds used in calibration;
- details on fitted regression line ($\log k$ versus $\log P$);
- preliminary estimate of the partition coefficient and the method used; and if a calculation method was used, its full description including identification of the data base and detailed information on the choice of fragments;
- average retention data and interpolated $\log P$ value for the test substance.

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A N N E X

P_{ow} Calculation Methods

Introduction

This annex provides a short introduction to the calculation of P_{ow}. For further information the reader is referred to textbooks (1) (2).

Calculated values of P_{ow} are used for:

- deciding which experimental method to use: Shake Flask method for log P_{ow} between -2 and 4 and HPLC method for log P_{ow} between 0 and 6;
- selecting conditions to be used in HPLC (reference compounds, methanol/water ratio);
- checking the plausibility of values obtained through experimental methods;
- providing an estimate when experimental methods cannot be applied.

Principle of calculation methods

Calculation methods are based on the theoretical fragmentation of the molecule into suitable substructures for which reliable log P_{ow} increments are known. The log P_{ow} is obtained by summing the fragment values and the correction terms for intramolecular interactions. Lists of fragment constants and correction terms are available, [(1) to (6)]. Some are regularly updated (3).

Reliability of calculated values

In general, the reliability of calculation methods decreases as the complexity of the compound under study increases. In the case of simple molecules of low molecular weight and with one or two functional groups, a deviation of 0.1 to 0.3 log P_{ow} units between the results of the different fragmentation methods and the measured value can be expected. The margin of error will depend on the reliability of the fragment constants used, the ability to recognise intramolecular interactions (e.g. hydrogen bonds) and the correct use of correction terms. In the case of ionising compounds the charge and degree of ionisation must be taken into consideration (10).

Fujita-Hansch π - method

The hydrophobic substituent constant, π , originally introduced by Fujita et al. (7) is defined as:

$$\pi_X = \log P_{ow}(\text{PhX}) - \log P_{ow}(\text{PhH}),$$

where PhX is an aromatic derivative and PhH the parent compound

$$[\text{e.g. } \pi \text{ Cl} = \log P_{\text{ow}}(\text{C}_6\text{H}_5\text{Cl}) - \log P_{\text{ow}}(\text{C}_6\text{H}_6) = 2.84 - 2.13 = 0.71].$$

The π - method is primarily of interest for aromatic compounds. π - Values for a large number of substituents are available (4) (5).

Rekker method

Using the Rekker method (8) the $\log P_{\text{ow}}$ value is calculated as:

$$\log P_{\text{ow}} = \sum_i a_i f_i + \sum (\text{interaction terms})$$

where a_i is the number at which a given fragment is present in the molecule and f_i is the $\log P_{\text{ow}}$ increment of the fragment. The interaction terms can be expressed as an integral multiple of one single constant C_m (so-called "magic constant"). The fragment constants f_i and C_m have been determined from a list of 1054 experimental P_{ow} values of 825 compounds using multiple regression analysis (6) (8). The determination of the interaction terms is carried out according to set rules (6) (8) (9).

Hansch-Leo method

Using the Hansch and Leo method (4), the $\log P_{\text{ow}}$ value is calculated as:

$$\log P_{\text{ow}} = \sum_i a_i f_i + \sum_j b_j F_j$$

where f_i is a fragment constant, F_j a correction term (factor), a_i and b_j the corresponding frequency of occurrence. Lists of atomic and group fragmental values and of correction terms F_j were derived by trial and error from experimental P_{ow} values. The correction terms have been divided into several different classes (1) (4). Software packages have been developed to take into account all the rules and correction terms (3).

Combined method

The calculation of $\log P_{\text{ow}}$ of complex molecules can be considerably improved, if the molecule is dissected into larger substructures for which reliable $\log P_{\text{ow}}$ values are available, either from tables (3) (4) or by existing measurements. Such fragments (e.g. heterocycles, anthraquinone, azobenzene) can then be combined with the Hansch- π values or with Rekker or Leo fragment constants.

Remarks

- i) The calculation methods are only applicable to partly or fully ionised compounds when the necessary correction factors are taken into account.

- ii) If the existence of intramolecular hydrogen bonds can be assumed, the corresponding correction terms (approx. +0.6 to +1.0 $\log P_{ow}$ units) must be added (1). Indications on the presence of such bonds can be obtained from stereo models or spectroscopic data.
- iii) If several tautomeric forms are possible, the most likely form should be used as the basis of the calculation.
- iv) The revisions of lists of fragment constants should be followed carefully.

LITERATURE ON CALCULATION METHODS

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